**Developmental Cell, Volume 47** 

### **Supplemental Information**

### **Combining Developmental and Perturbation-Seq**

#### **Uncovers Transcriptional Modules**

#### **Orchestrating Neuronal Remodeling**

Idan Alyagor, Victoria Berkun, Hadas Keren-Shaul, Neta Marmor-Kollet, Eyal David, Oded Mayseless, Noa Issman-Zecharya, Ido Amit, and Oren Schuldiner





# <u>Figure S1</u> – 71G10-Gal4 is robustly and almost exclusively expressed in MB $\gamma$ neurons throughout development, related to Figure 1

Confocal Z-projections of MB lobe (A<sub>1</sub>-N<sub>1</sub>) and whole brains (A<sub>2</sub>-N<sub>2</sub>) where the  $\gamma$  neurons labeled with mCD8-GFP (GFP) driven by 71G10-Gal4 (71G10), at all the time points taken for the developmental RNA-seq. Whole brain projections are intentionally over exposed in order to emphasize the near absence of labeled neurons outside the MB expressing GFP. Scale bars represent 15 µm for MB-lobes and 30 µm for whole brains.



#### Figure S2 – Quality control analyses of cell isolation, related to Figure 1.

(A) Confocal Z-projections of larval (A-A') and adult (B-B') MB  $\gamma$  neurons labeled with nuclear DsRed (RedStinger) and mCD8-GFP (GFP) driven by 71G10-Gal4 (71G10). Low magnification projections that show whole brains (A''-B'') demonstrate the specificity of 71G10 within the entire brain. Scale bars represent 15  $\mu$ m for lobes and 30  $\mu$ m for whole brains.

(C) FACS gating strategy for isolating DsRed positive MB  $\gamma$  neurons from dissociated brains. Forward scatter – FSC is plotted against Side scatter – SSC (C) or DsRed expression (C').

(D) FACS analysis of DsRed positive cells from dissociated brains before (D) and after (D') positive sorting according to the gating strategy presented in (C). Note that D represents the entire cell population resulting in lower DsRed<sup>+</sup> cell proportion (1.3%) compared to C', where the analysis includes only live cells that passed a certain SSC and FSC threshold (2.5% DsRed<sup>+</sup> cells).



# <u>Figure S3</u> – Gene expression profiles of dynamic genes from selected terms, related to Figure 2.

(A-C) Heat map depicting the relative expression patterns of dynamic genes belonging to the described GO terms. The magenta scale depicts the intensity of the peak expression of each gene relative to that of other genes in the group. The genes are ordered according to their developmental cluster and then by their max expression peak.





# <u>Figure S4</u> – **DNA-binding-protein screen uncovers new genes required for remodeling, related to Figure 3**

(A) Heat map of the relative expression patterns of the DNA binding proteins (DBPs) taken for the screen, sorted by cluster and peak expression levels (magenta). P, R and (-) stand for pruning, regrowth or WT phenotypes, respectively. While known roles of genes are labeled in black, new findings from this study are labeled in orange. Asterisks demarcate examples of genes whose expression is downregulated at the onset of pruning - which we therefore tested by overexpression. § demarcate the fact that while expression of Bap60 RNAi resulted in a pruning defect, we were not able to confirm this using mutant analysis.

(B-L) Confocal Z-projections of MBs of the indicated age in which 71G10-Gal4 drives the expression of mCD8-GFP as well as the indicated RNAi/overexpression transgenes (indicated by 71G10>transgene) OR, brains in which 71G10-Gal4 ( $D_{2-3}$ ,  $E_{2-3}$ ,  $G_{2-3}$ ) or 201Y-Gal4 ( $F_{2-3}$ ) drive the expression of mCD8-GFP within a MARCM clone of the indicated genotype.

Pruning defects are marked by arrows, and regrowth defects are evident when the adult  $\gamma$  lobe does not fully occupy the white dashed line. Scale bars represent 15  $\mu$ m. The numbers (x/n) on the lower left corners depict the number of times the phenotype was observed out of the total hemispheres examined.



# <u>Figure S5</u> – Antibody staining of selected DBPs during development correlate to their RNA expression profile, related to Figure 3

(A-D) Single confocal slices of the cell body region of MB neurons labeled with mCD8-GFP (GFP, green) driven by 71G10-Gal4 co-stained using the indicated antibodies (magenta) at the indicated developmental stages. Protein peak expression within  $\gamma$  neurons that are labeled in green, for each of the DBPs (0h APF for EcR-B1 and Tai, 6h APF for Mamo and 0h APF for Sox14) correlates to the RNA expression peak as referred from the RNA-seq. n>8 lobes for each experiment, scale bars represent 15 µm.







# <u>Figure S6</u> – CRISPR mediated deletion design, and *tai* function at the same epistatic level as EcR, related to Figure 3

(A) A schematic representation of the gRNA design to induce CRISPR/Cas9-mediated deletions in *Sox14*, *chinmo* and *mamo*. Green bars represent coding exons, grey bars represent non-coding exonal sequences and lines represent introns. All annotated isoforms for each gene are displayed – while  $Sox14^{CRISPR\Delta1}$  and *chinmo*<sup>CRISPR\Delta1</sup> alleles are deletions of most of the coding sequence, *mamo*<sup>CRISPR\Delta1</sup> allele induces a frameshift 11 base pairs downstream of the annotated ATG (resulting in a premature stop codon). The gRNAs were designed to delete most of the coding sequence of the gene.

**(B)** Quantification of the pruning severity in Figure 3D, 3H and 3O using blind ranking analysis of the described genotypes. \*\*\* p<0.001, See experimental procedures for details.

(C-D) Single confocal sections of the cell body region of  $tai^{61G1}$  MARCM MB neuroblast clones stained for anti-Sox14 (C, n=6) or anti-Mamo (D, n=4) at 0h APF and 6h APF, respectively. Expression of Sox14 (C) and Mamo (D) is 4.2-fold (p<0.001) and 2.4-fold (p<0.01) decreased within the  $tai^{61G1}$  clone, respectively. Dashed lines demarcate the boundaries of the clone. Green represents mCD8-GFP driven by 71G10-Gal4, magenta represents Sox14 (C) or Mamo (D) antibody staining, which are also shown in grey. Scale bars represent 15 µm.



# <u>Figure S7</u> – Expression profiles of neurons perturbed for key transcription factors uncovers hierarchical regulation of axon pruning by regulatory factors, related to Figure 5

(A-H) Normalized expression of *E75* (A), *tai* (B), *mamo* (C), *Hr3* (D), *chinmo* (E), *Blimp-1* (F), *prospero* (G) or *HmgZ* (H) in WT MB  $\gamma$  neurons and in those expressing the indicated transgene (\* p<0.05, \*\* p<0.01, \*\*\* p<0.001). Error bars indicate SEM, units on the y axis are arbitrary.

(I-N, P) Single confocal slices of MB cell bodies containing MARCM clones labeled with mCD8-GFP (GFP) driven by 71G10 (I,K,N,P) or 201Y (J,L-M) additionally expressing  $EcR^{DN}$  (I, n=6, M, n=4), *E*75-RNAi (J, n=8, L, n=8), homozygous for  $Sox14^{CRISPR\Delta1}$  (K, n=9, N, n=6) or homozygous for *chinmo*<sup>CRISPRA1</sup> (P, n=3) stained using the indicated antibody at the indicated developmental stage. The anti-Tai staining was decreased by 3.1-fold (p<0.001) within clones expressing  $EcR^{DN}$  (I). The anti-Mamo staining was decreased by 2.2-fold (p<0.01) within clones expressing *E*75-RNAi (L) and by 7.1-fold within clones expressing  $EcR^{DN}$  (M). The anti-EcR-B1 staining was decreased by 3.5-fold (p<0.001) within clones expressing *chinmo*<sup>CRISPRA1</sup>. The boundaries of the clones are represented by dashed lines.

**(O)** Confocal Z-projections of an adult MB labeled by 71G10-Gal4 driving the expression of mCD8-GFP and an *Hr*3 transgene. Arrow indicates dorsally projecting unpruned  $\gamma$  axons. The numbers (x/n) on the lower left corners depict the number of times the phenotype was observed out of the total hemispheres examined.

Green represents mCD8-GFP expression driven by the indicated Gal4. Magenta and Gray represent the antibody indicated. Scale bars represent 15  $\mu$ m.

#### <u>Table S2 – Enrichment analysis of selected GO, KEGG and Reactome terms within</u> <u>developmental clusters</u>

Cluster	Genes within cluster	Selected enriched terms	Term enrichment (see	Adjusted p value=
I	266	Synaptic transmission [GO:0007268]	35/184	5 x 10 <sup>-13</sup>
		Regulation of neurotransmitter levels [GO:0001505]	21/86	4 x 10 <sup>-9</sup>
		Voltage-gated cation channel activity [GO:0022843]	10/26	4 x 10 <sup>-6</sup>
П	188	Glycolytic process [GO:0006096]	10/16	7 x 10 <sup>-7</sup>
111	270	Proteasome-mediated ubiquitin-dependent protein catabolic process [GO:0043161]	38/123	4 x 10 <sup>-22</sup>
		Neuron projection morphogenesis [GO:0048812]	35/401	4 x 10 <sup>-4</sup>
IV	404	Programmed cell death [GO:0012501]	32/196	7 x 10 <sup>-4</sup>
		Autophagy [GO:0006914]	29/116	2 x 10 <sup>-7</sup>
		Endosome transport via multivesicular body sorting pathway [GO:0032509]	8/16	8 x 10 <sup>-4</sup>
		Endosomal transport [GO:0016197]	15/65	0.014
V	239	COP9 signalosome [GO:0008180]	6/9	1 x 10⁻⁵
VI	309	Axon guidance [GO:0007411]	24/182	4 x 10 <sup>-4</sup>
		Axon development [GO:0061564]	28/242	3 x 10 <sup>-4</sup>
		Spliceosomal complex [GO:0005681] Spliceosome (map03040)	32/140 25/97	3 x 10 <sup>-13</sup> 2 x 10 <sup>-10</sup>
		Chromatin remodeling [GO:0006338]	19/78	2 x 10 <sup>-7</sup>
VII	247	Protein processing in endoplasmic reticulum [map04141]	13/96	0.002
VIII	316	Cellular respiration [GO:0045333]	21/79	1 x 10 <sup>-7</sup>
		Mitochondrial part [GO:0044429]	46/196	3 X 10 <sup>-13</sup>
IX	333	Neurotransmitter secretion [GO:0007260]	18/80	1 x 10 <sup>-5</sup>
Х	101	Ribosome [GO:0005840]	25/55	1 x 10 <sup>-13</sup>

Enrichment analysis of selected GO, KEGG and Reactome terms that are significantly enriched (p<0.05) after Holm–Bonferroni correction. Term enrichment refers to the number of term related genes within the cluster out of all of the *Drosophila* term related genes has significant expression in the  $\gamma$  neurons.

#### <u>Table S6 – Enrichment analysis of selected GO, KEGG and Reactome terms within</u> <u>sub-clusters</u>

Cluster	Genes within cluster	Selected enriched terms	Term enrichment	Adjusted p value=
la	72	Synaptic transmission [GO:0007268]	12/35	9 x 10 <sup>-4</sup>
lb	75	Plasma membrane proton-transporting V- type ATPase complex [GO:0033181]	6/9	1 x 10 <sup>-6</sup>
lc	43	Voltage-gated cation channel activity [GO:0022843]	5/10	1 x 10 <sup>-4</sup>
lla	45	Glycolytic process [GO:0006096]	8/10	2 x 10 <sup>-10</sup>
llb	38			
llc	61			
Illa	65	Proteasome complex [GO:0000502]	29/32	4 x 10 <sup>-49</sup>
IIIb	32			
IIIc	47	Neuron projection morphogenesis [GO:0048812]	13/35	0.003
IVa	125	Autophagy [GO:0006914]	14/29	1 x 10 <sup>-4</sup>
		ESCRT complex [GO:0036452]	4/7	0.035
		Endosome [GO:0005768]	10/26	0.01
IVb	101	programmed cell death involved in cell development [GO:0010623]	9/21	0.01
		Peptidase family C14A, cysteine active site [IPR033139]	3/3	0.01
IVc	33	Ferritin complex [GO:0070288]	2/2	0.009
Va	39			
Vb	63	RHO GTPases Activate Formins [R-DME- 5663220]	3/3	0.048
Vla	36	Chromatin organization [GO:0006325]	10/41	0.01
Vlb	35	Protein processing in endoplasmic reticulum [map04141]	7/13	3 x 10 <sup>-4</sup>
Vlla	47			
VIIc	40			
VIIIa	122	Mitochondrial part [GO:0044429]	23/46	6 x 10 <sup>-7</sup>
		Citrate cycle TCA cycle [map00020]	6/11	0.02
VIIIb	38			
IXa	135	Synaptic transmission [GO:0007268]	19/34	2 x 10 <sup>-6</sup>
IXb	57	Cation channel complex [GO:0034703]	4/8	0.049

Enrichment analysis of selected GO, KEGG and Reactome terms that are significantly enriched (p<0.05) after Holm–Bonferroni correction. Term enrichment refers to the number of term related genes within the cluster out of the number of genes from the functional group within the parent cluster.