### **Supplementary materials**

### Endocrine therapy-resistant breast cancer model cells are inhibited by soybean glyceollin I through *Eleanor* non-coding RNA

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### Supplementary Table 1

		Control	Resveratrol	Fr.4	Fr.7	Control	Fr.2	Fr.3	Control	Fr.6
ESR1	Ave.	20.22	26.10	04 77	05.47	10.40	20.42	22.01	20.24	25.24
	(n=3)	20.33	20.10	24.77	25.47	19.40	20.13	22.91	20.24	25.24
	SD	0.95	0.48	0.40	0.81	0.77	0.62	1.50	0.53	0.43
GAPDH	Ave.	15.61	17.02	16.40	17.65	15.67	15.04	16 50	15 50	16.06
	(n=3)	10.01	17.03	16.49	17.05	10.07	15.94	10.00	15.52	10.90
	SD	0.24	0.17	0.35	0.95	0.38	0.12	0.70	0.51	0.33

Ct values of Figure 2B

### Supplementary Table 2

Ct value of Figure 4B

		Control	Resveratrol	Fr.6	Glyceollin I	
ESR1	Ave. (n=3)	17.95	21.41	21.43	21.19	
	SD	0.41	0.34	0.70	1.37	
GAPDH	Ave. (n=3)	15.96	16.15	16.07	15.95	
	SD	0.14	0.50	0.99	0.50	

# Supplementary Figure S1. Estradiol inhibited *Eleanors* and LTED cell proliferation.

(A) FISH analysis showing that the *Eleanor* RNA cloud regressed in LTED cells treated with high concentrations of estradiol. Scale bar, 10 µm. (B) The *ESR1* mRNA level was decreased by estradiol in LTED cells. The *ESR1* mRNA level was measured by qRT-PCR. Values were normalized against *GAPDH* mRNA, and values for cells treated with control (DMSO) in LTED cells were set to 1. The bars represent the means  $\pm$  S.D. n = 3, \*\*p < 0.01. (C) LTED cell proliferation was inhibited by high concentrations of estradiol. The values represent the means  $\pm$  S.D. n = 3, \*\*p < 0.01. (C) LTED cell proliferation was inhibited by high concentrations of estradiol. The values represent the means  $\pm$  S.D. n = 3, \*\*p < 0.01. (D) LTED cell growth was inhibited by estradiol in a dose-dependent manner. The values represent the means  $\pm$  S.D. n = 3. (E) Immunoblot showing that the ER protein levels were decreased in LTED cells by estradiol treatment. Actin was used as an internal control. MW, molecular weight. Full-length blots are presented in Supplementary Fig. S6.

Supplementary Figure S2. Glyceollin I decreased the ER protein level and its transcriptional activity in LTED cells.

(A) Immunoblot showing that the ER level was reduced by glyceollin

I in LTED cells. Actin was used as an internal control. Full-length blots are presented in Supplementary **Fig. S6**. **(B)** Quantitative RT-PCR showing that the expression levels of the ER target genes (*GREB1*, *TFF1*, *PGR*) decreased with glyceollin I treatment. The values for the control treatment were set to 1. The bars represent the means  $\pm$  S.D. n = 3, \*p < 0.05, \*\*p < 0.01.

## Supplementary Figure S3. The effects of resveratrol and glyceollin I on MCF7 and tamoxifen-resistant cells.

(A) MCF7 cells were inhibited with resveratrol and glyceollin I in a dosedependent manner. The effect of glyceollin I in MCF7 cells was lower than in LTED cells, while the effect of resveratrol was comparable in MCF7 and LTED cells (see **Fig. 4E**). (**B**) RNA FISH showing that the *Eleanor* RNA cloud was not detected in TamR cells. Scale bar, 10  $\mu$ m. (**C**) The level of *ESR1* mRNA decreased slightly by the glyceollin I treatment of TamR cells. Quantitative RT-PCR was performed to measure the relative *ESR1* mRNA levels in the indicated cells. Values were normalized against *GAPDH* mRNA, and values for cells treated with DMSO (control) in TamR cells were set to 1. The bars represent the means  $\pm$  S.D. n = 3, \*p < 0.05; \*\*p < 0.01. (**D**) Immunoblot showing that the ER protein levels slightly decreased with the glyceollin I treatment of TamR cells. Actin was used as a loading control. Full-length blots are presented in Supplementary **Fig. S6**. **(E)** TamR cell proliferation was inhibited by glyceollin I. The cell growth was measured with a colorimetric assay at 490 nm (Kit-8). The values represent the means  $\pm$  S.D. n = 3, \*\*\*p < 0.001.

### Supplementary Figure S4. Resveratrol, estradiol and glyceollin I induced apoptosis in LTED cells.

FACS analysis of Annexin V positive cells. Treatment of LTED cells with resveratrol, estradiol and glyceollin I increased a population with intense labeling with Annexin V, as indicated by bars. Data from triplicate experiments are shown.

## Supplementary Figure S5. Glyceollin I exerts its inhibitory effect on *Eleanors* independently of ER.

(A) Immunofluorescences confirmed ER degradation by RNAi knockdown.
LTED cells were treated with siRNA targeted to the *ESR1* mRNA (si*ESR1*) or control siRNA (si*GL3*) for 24 h, and ER was stained with antibodies. Scale bar, 10 μm. (B)
FISH analysis of the *Eleanor* RNA cloud (green) in LTED nuclei (blue). *Eleanors* were inhibited by 50 μM resveratrol, estradiol, or glyceollin I in the control knockdown (left,

from the second to the bottom row). *Eleanors* were recovered by ER knockdown in resveratrol or estradiol-treated LTED cells (right, the second or third row). On the other hand, the inhibition of *Eleanors* by glyceollin I was maintained even in the absence of ER (right, bottom row). All drug treatments were for 24 h. Scale bar, 10 µm. **(C)** The activity of the mTOR pathway components was decreased by resveratrol and glyceollin I treatments in LTED cells. Immunoblotting revealed that the phosphorylation level of the mTOR substrate (phospho-p70 S6 kinases) decreased with glyceollin I treatment (middle panel), while the mTOR protein level did not change (top panel). Actin was used as a loading control (bottom panel). Full-length blots are presented in Supplementary Fig. S6. **(D)** The A to G mutation corresponding to ESR1<sup>Y537C</sup> was detected in a sub-population of LTED cells (black arrow). DNA sequencing profiles obtained from Sanger capillary electrophoreses are shown.

### Supplementary Figure S6. Original immunoblot images.

Uncropped immunoblots for Figs S1E, S2A, S3D and S5C are shown. Separate blots are delineated with black lines.

0

0 h

24 h

48 h

72 h













### Supplementary Figure S5. Yamamoto et al

### Supplementary Figure S1E, S2A

#### 250 - 120 - 100 - 80 - 60 - 60 - 40 - 100 -100 -



Supplementary Figure S5C



### Supplementary Figure S6. Yamamoto et al

### Supplementary Figure S3D