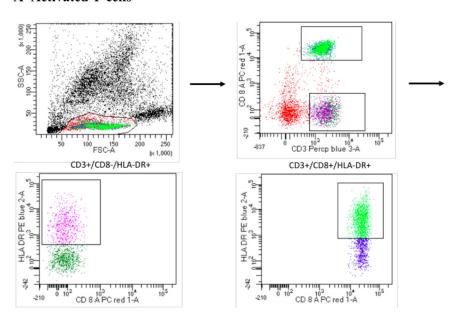
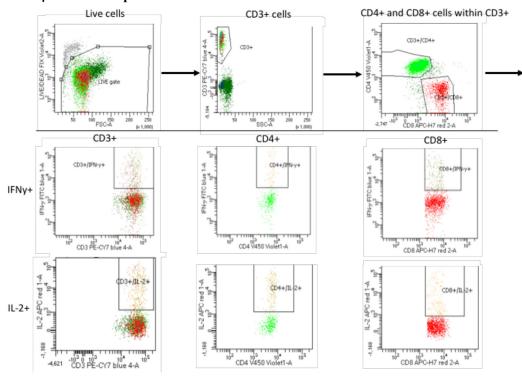
Lenalidomide combined with low-dose cyclophosphamide and prednisone modulates Ikaros and Aiolos in lymphocytes, resulting in immunostimulatory effects in lenalidomiderefractory multiple myeloma patients

## SUPPLEMENTARY MATERIALS

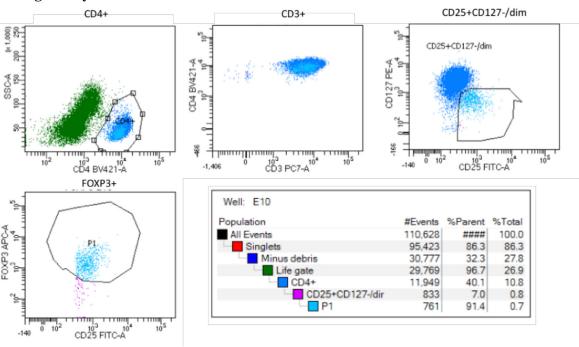
## A Activated T-cells



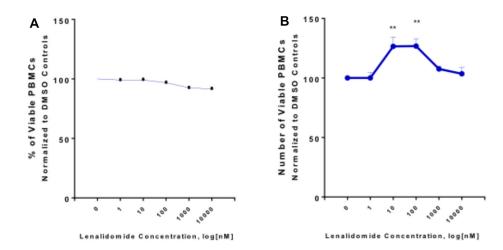
## B IFNy and IL-2 expression in T-cell subsets



## C Regulatory T-cells



Supplementary Figure 1: Representative flow cytometry plots showing the gating strategy to detect different lymphocyte subsets. (A) Activated T-cells, defined as the percentage of T-cells expressing HLA-DR. CD3+/CD8- cells were considered CD4+ T-cells. (B) IFNγ and IL-2 positive T-cells. Cytokine production (IL-2 and IFNγ) of T-cells was measured after stimulating PBMCs with CD3/CD28 Human T-activator Beads (Dynabeads®) in a 1:1 ratio for 5 hours at 37° C in the presence of an inhibitor of intracellular protein transport (Brefeldin A, eBioscience). (C) Regulatory T-cells were defined as CD3+/CD4+/CD25+/CD127-/dim. FOXP3 intracellular staining was used to confirm the FOXP3 positivity of our defined regulatory T-cells. The FOXP3/Transcription Factor Staining Buffer Set (eBioscience) was used for the fixation and intracellular staining according to the manufacturer's protocol. The LIVE/DEAD® Fixable Dead Cell Staining kit (Thermofisher Scientific) was used to determine the viability of the cells prior to the fixation and permeabilization and subsequent intracellular staining.



**Supplementary Figure 2: Effect of lenalidomide on viability of PBMCs.** Following DMSO or lenalidomide treatment and anti-CD3 stimulation for 72 hours, and prior to co-culture, viability of PBMCs was determined by flow cytometry. After 2 washes with PBS, these PBMCs were stained with Annexin-V (BD Biosciences) and To-Pro3 (Invitrogen) according to manufacturer's protocol, and CountBright<sup>TM</sup> Absolute Counting Beads (ThermoFisher) were added for absolute cell count. Live PBMCs were gated as Annexin-Vand Topro- population, and enumerated by the counting beads according to manufacturer's protocol. (A) Percent viable PBMCs of total PBMCs was plotted (n = 4). (B) Viable PBMCs were enumerated with Countbright<sup>TM</sup> absolute counting beads according to manufacturer's protocol (n = 4). P-values were calculated using one-way ANOVA, \*\*P < 0.01.