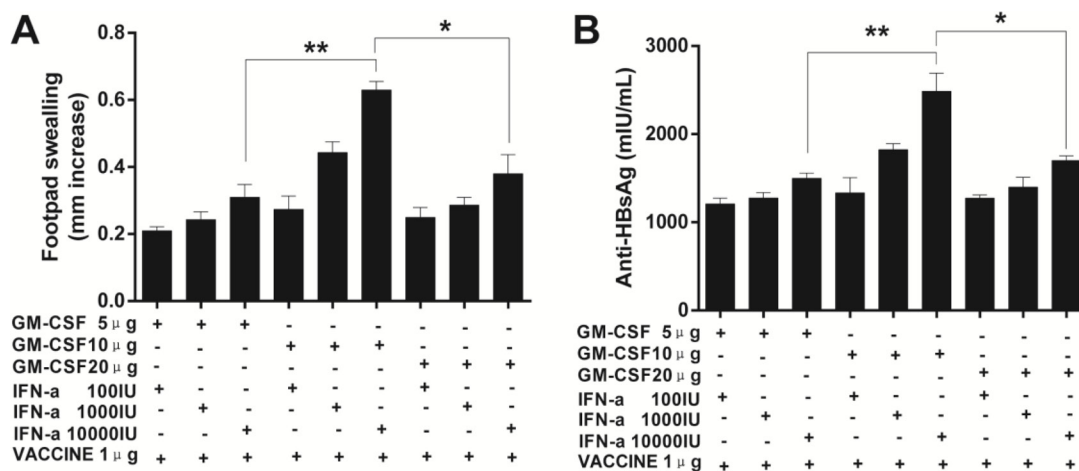
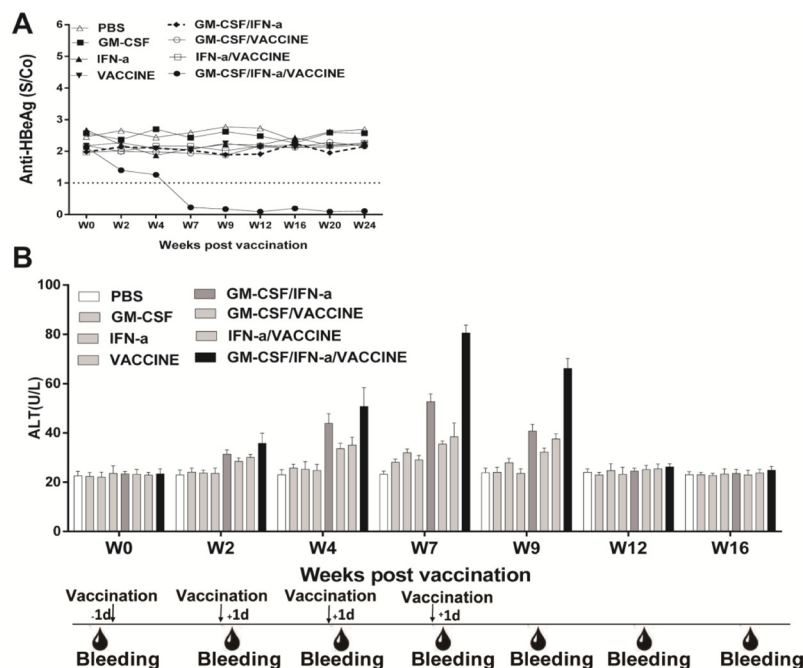


Clearance of HBeAg and HBsAg of HBV in mice model by a recombinant HBV vaccine combined with GM-CSF and IFN- α as an effective therapeutic vaccine adjuvant

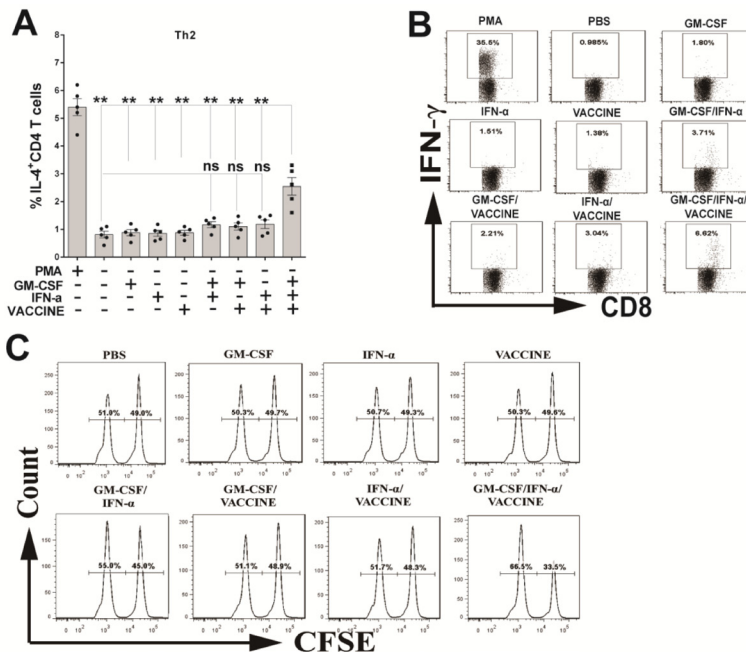
SUPPLEMENTARY MATERIALS



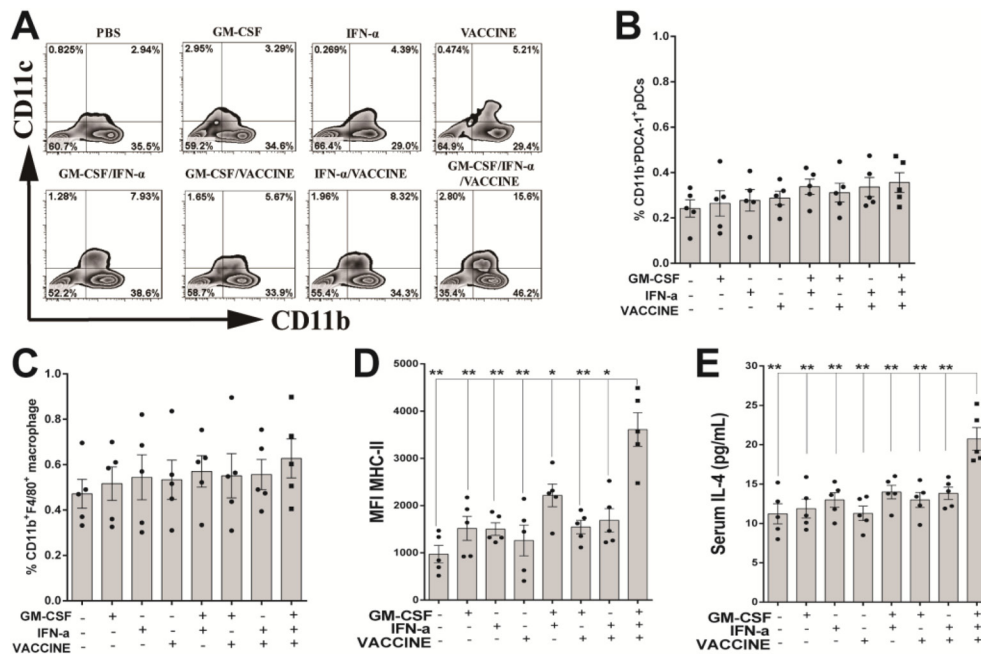
Supplementary Figure 1: GM-CSF combined with IFN- α enhanced immune response of HBV vaccine (VACCINE). The wild type C57BL/6 mice were immunized three times with VACCINE (1 μ g) combined with different dose formulation of GM-CSF and IFN- α in a 2-week interval. **(A)** 7 days after the third immunization, DTH were tested at 24 hours after HBsAg challenge at footpads of immunized mice. **(B)** 7 days after the third immunization, serum of immunized mice were collected for anti-HBsAg test by ELISA. Bars represent mean \pm SEM from 3 independent experiments, * P < 0.05; ** P < 0.01; ns, not significant.



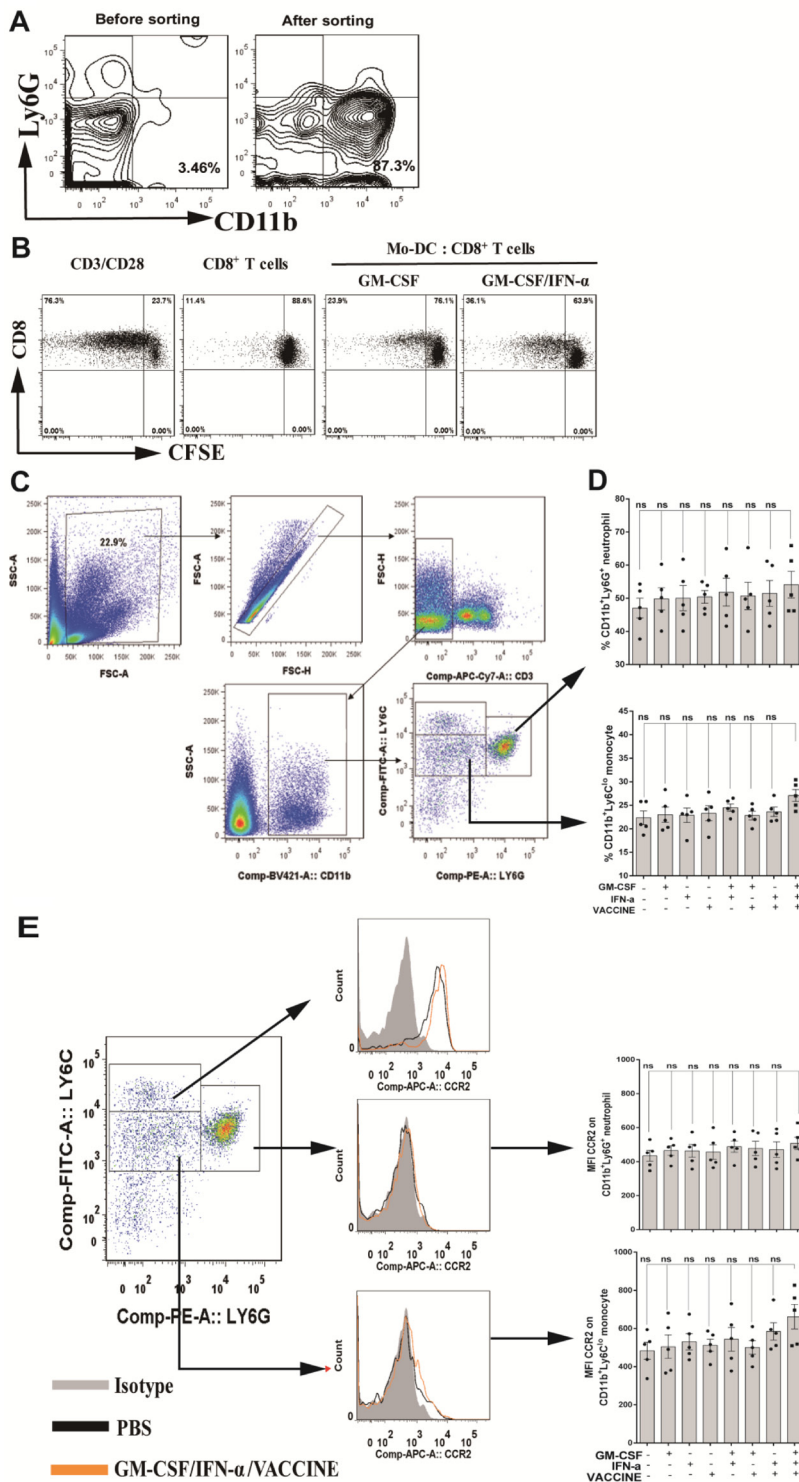
Supplementary Figure 2: Responses of AAV/HBV infected mice to the GM-CSF/IFN- α /VACCINE regimen. **(A)** At intervals up to 24 weeks, serum anti-HBeAg was determined by ELISA. Dotted lines represent the threshold of detection. >1 means anti-HBeAg-negative, and <1 means anti-HBeAg-positive. **(B)** One day before the first vaccination, the blood samples were collected from the AAV/HBV infected mice. Serum ALT was determined as a background level. Then the model mice were bled one day after the rest of three immunizations. After the end of vaccination, the mice were bled at 2-week, 3-week and 4-week follow up.



Supplementary Figure 3: GM-CSF/IFN- α /VACCINE elicit cellular immunity against AAV/HBV infection. (A) 14 days after the fourth immunization, the spleen cells were stimulated with 10 μ g/mL HBsAg for 18 hours *in vitro*. Cells were incubated for 6 hours with PMA/Ionomycin (100 ng/mL / 1 μ g/mL) as a positive control. Th2 was detected by intracellular cytokine. The percentage of IFN- γ ⁺CD4⁺ T cells is presented. (B) Representative flow cytometric profiles of IFN- γ ⁺CD8⁺ T cells. (C) Representative flow cytometric histograms of HBsAg-specific CTL activity *in vivo* in AAV/HBV infected mice.



Supplementary Figure 4: GM-CSF/IFN- α /VACCINE promoted both the production and the function of CD11b⁺CD11c⁺ DC. (A) Representative flow cytometric profiles of CD11b⁺CD11c⁺ DC in blood. (B–C) Percentage of CD11b⁺F4/80⁺ macrophage and CD11b⁺PDCA-1⁺ plasmacytoid dendritic cells in blood of AAV/HBV infected mice inoculated with different formulations of the vaccine. (D) Comparison of MHC-II on of CD11b⁺CD11c⁺ DC in blood. (E) Serum concentration of IL-4 of AAV/HBV infected mice.



Supplementary Figure 5: GM-CSF plus IFN- α promoted both the production and the function of CD11c⁺ monocyte derived DC (MoDC) *in vitro* and *in vivo*. (A) CD11b⁺Ly6G⁻ monocytes were sorted from PBMC of male wild-type C57BL/6 ($n = 50$). (B) Conventional CD8⁺ T cells were sorted from spleen cells of AAV8-1.3HBV mice and co-cultured with HBsAg (10 μ g/mL)-pulsed MoDC which generated from GM-CSF or GM-CSF/IFN- α at ratio 10:1 for 72 hours. CD3/CD28 (1 μ g/mL / 100 ng/mL) was used as a positive control. (C) Flow cytometry gating of inflammatory monocytes (CD3⁻CD11b⁺Ly6G^{hi}), circulating monocytes (CD3⁻CD11b⁺Ly6G^{lo}), and granulocytes (CD3⁻CD11b⁺Ly6G^{hi}) in blood of AAV8-1.3HBV-infected mice. (D) Percentage of blood Ly6G⁻Ly6C^{lo} monocytes, and Ly6G⁺Ly6C⁺ granulocytes of AAV8-1.3HBV-infected mice treated different formulation vaccine for 3 days. Numbers adjacent to outlined area indicate percent Ly6G⁻Ly6C^{hi} monocytes (top left), Ly6G⁻Ly6C^{lo} monocytes (bottom left), and Ly6G⁺Ly6C⁺ granulocytes (right). Bars are shown as mean \pm SEM. ns, not significant. (E) Comparison of MFI CCR2 on Ly6G⁻Ly6C^{lo} monocytes, and Ly6G⁺Ly6C⁺ granulocytes. Data are pooled from two independent experiments with 5 mice per group. Bars are shown as mean \pm SEM. ns, not significant.