

Carm1^{F1/F1};Vav1-Cre *Carm1^{F1/F1};Vav1-Cre* *Carm1^{Δ/Δ};Vav1-Cre⁺* *Carm1^{Δ/Δ};Vav1-Cre⁺*

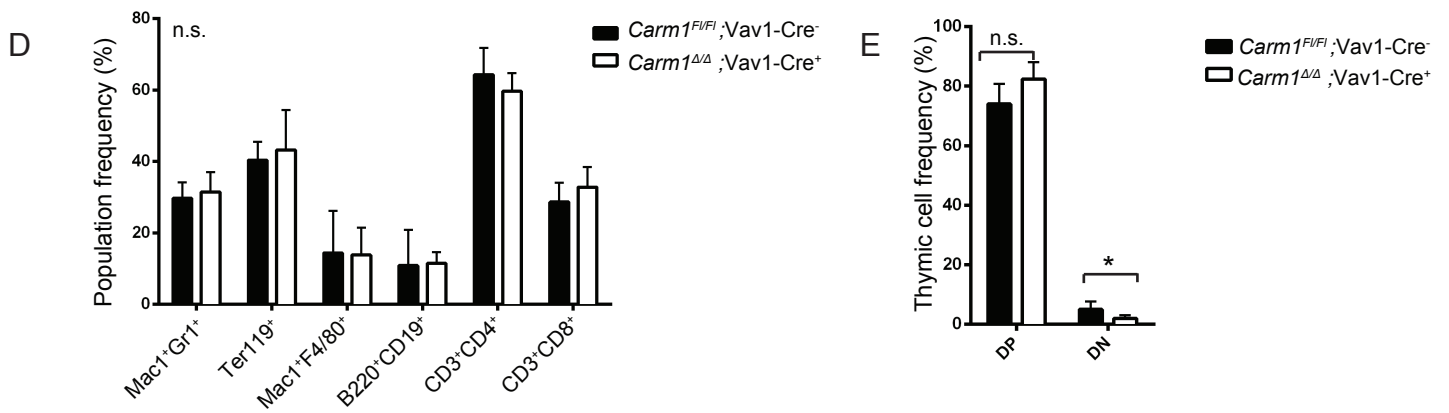
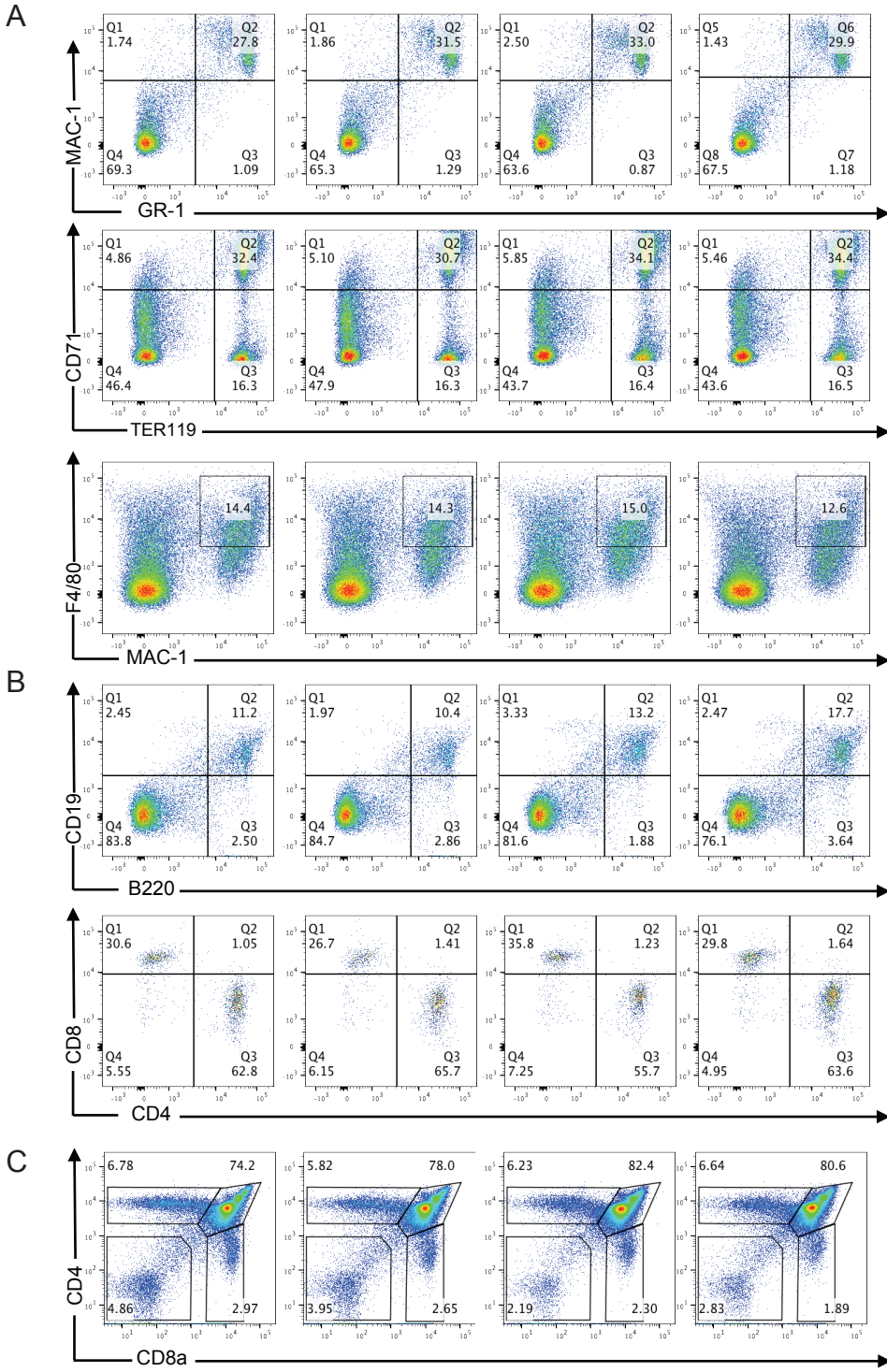
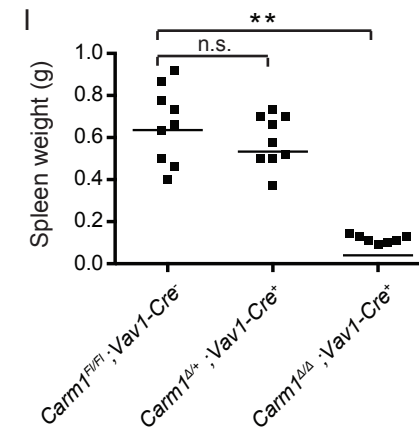
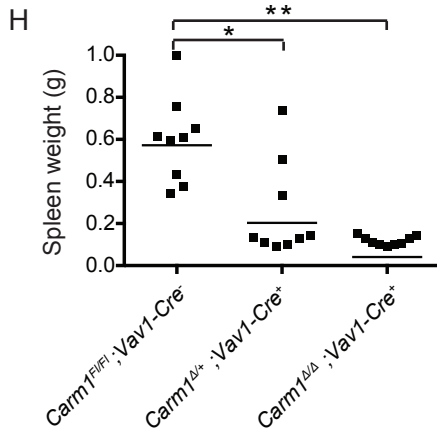
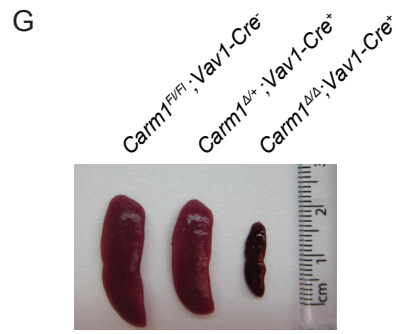
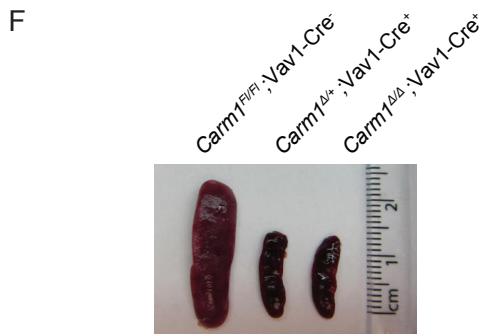
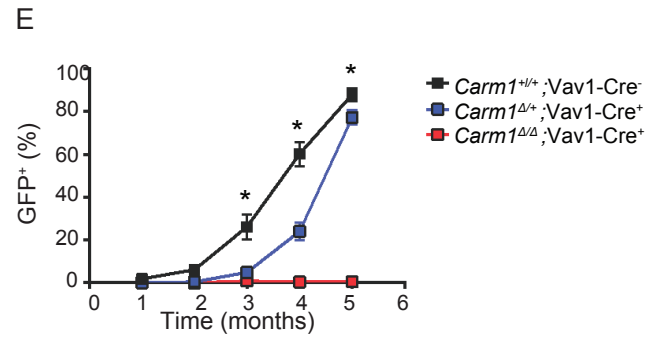
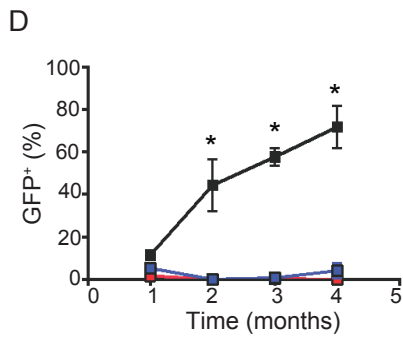
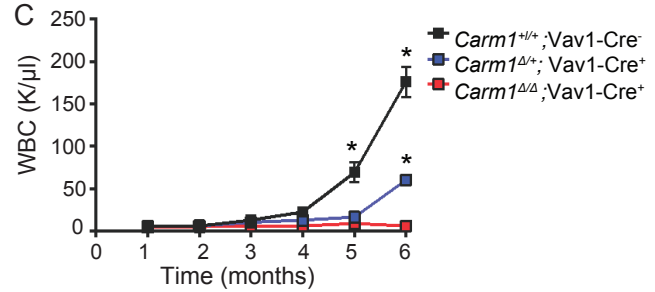
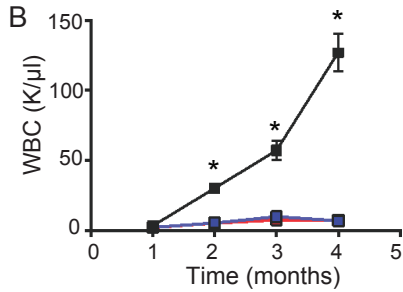
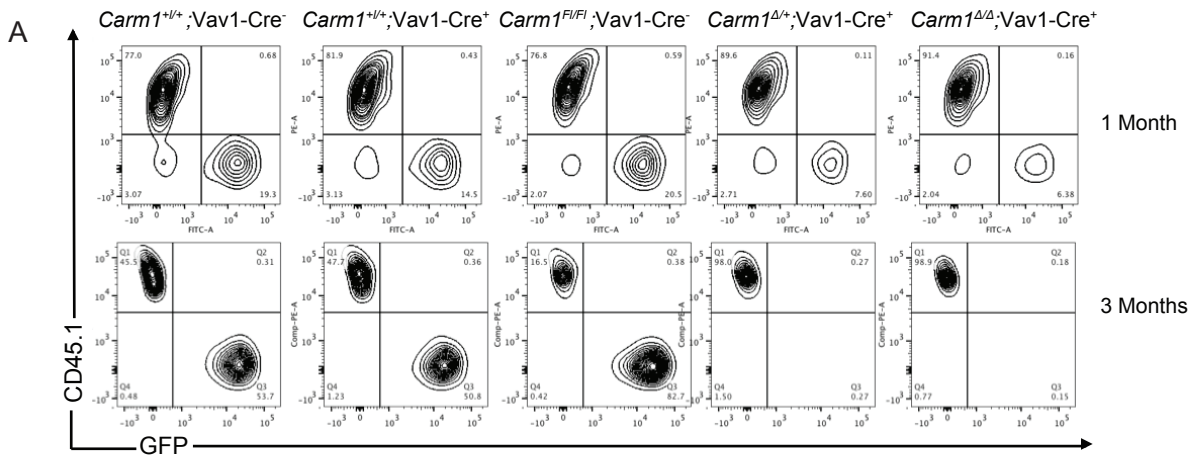
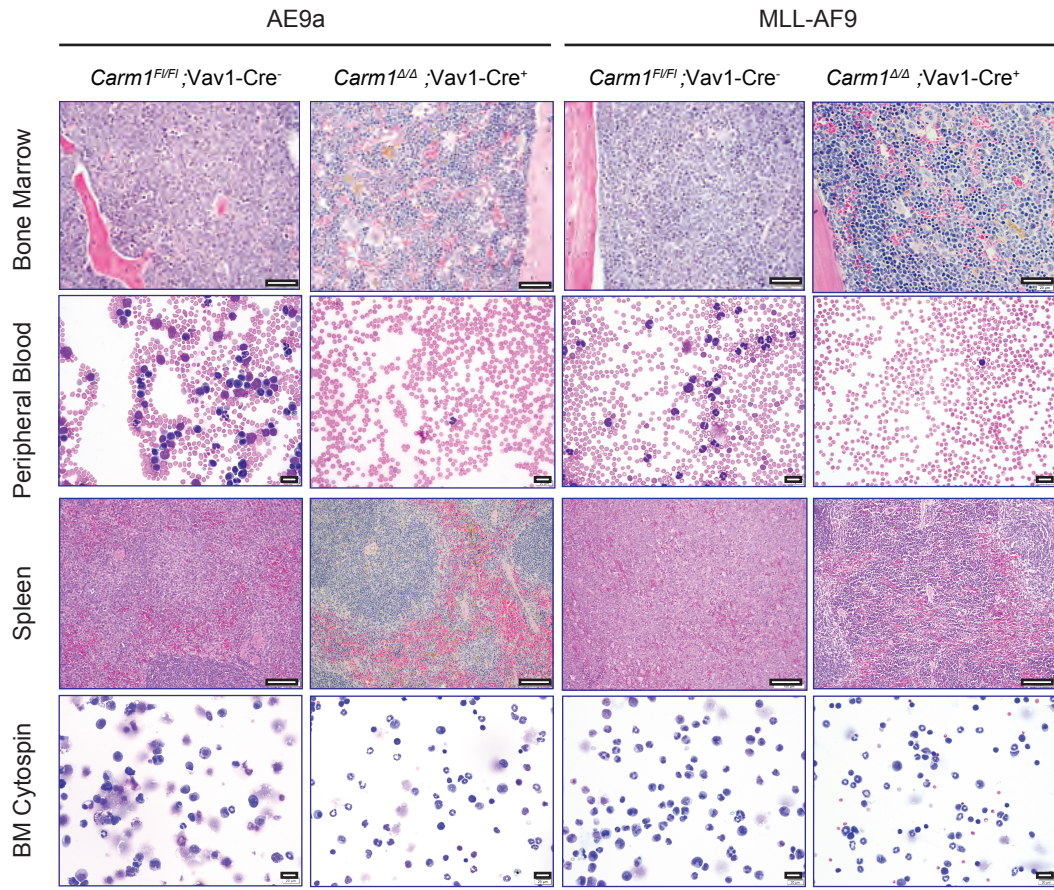


Figure S1: Flow cytometry analysis of mature bone marrow, spleen, and thymic hematopoietic populations, Related to Figure 2.

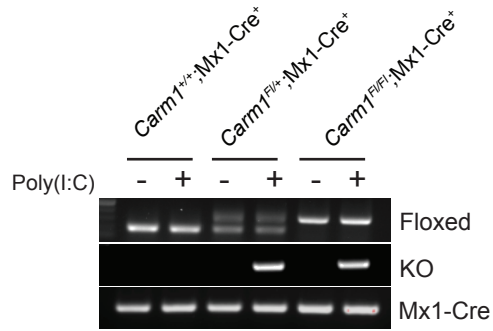
- A) Representative flow cytometry analysis of mature myeloid, erythroid, and macrophage populations in the bone marrow of 1-year-old mice.
- B) Representative flow cytometry analysis of mature B cell and T cell populations in the spleen of 1-year-old mice.
- C) Representative flow cytometry analysis of T cell populations in the thymus of 1-year-old mice.
- D) Percentages of *Carm1*^{Δ/Δ};Vav1-Cre⁺ and *Carm1*^{F1/F1};Vav1-Cre⁻ age-matched mice for mature bone marrow and spleen populations. n=5, n.s.= no significant differences.
- E) Average percentages of *Carm1*^{Δ/Δ};Vav1-Cre⁺ and *Carm1*^{F1/F1};Vav1-Cre⁻ age-matched mice for thymic T-cell populations. n=5, n.s.= no significant difference. *p< 0.05. Bar graphs represent the mean ± SD. Statistics represent a Student's t-test for samples of unequal variance.



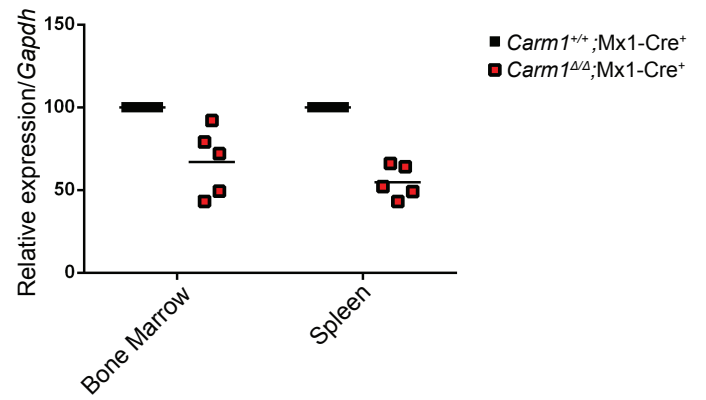
J



K



L



M

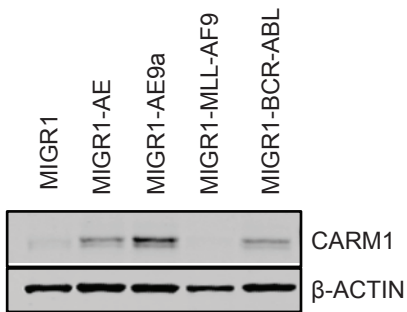


Figure S2: Characterization of AE9a and MLL-AF9 mice by flow cytometry, organ histology, and blood counts, Related to Figure 3.

- A) Representative flow cytometry plot of GFP⁺ and CD45.1⁺ peripheral blood engraftment over time in the AE9a recipient initiation experiments.
- B) Average AE9a *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient white blood cell count (WBC) over time. n = 10, *p<0.01
- C) Average MLL-AF9 *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient white blood cell count (WBC) over time. n = 10, *p<0.01
- D) Average AE9a *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient peripheral blood GFP⁺ percentage over time. n = 10, *p<0.01
- E) Average AE9a *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient peripheral blood GFP⁺ percentage over time. n = 10, *p<0.01
- F) Representative spleen size for AE9a *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient mice, three months post-transplant.
- G) Representative spleen size for MLL-AF9 *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient mice, three months post-transplant.
- H) Average spleen weight for AE9a *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient mice, three months post-transplant. n = 10, *p<0.01, **p<0.001
- I) Average spleen weight for MLL-AF9 *Carm1*^{F1/F1};Vav1-Cre⁻, *Carm1*^{Δ/+};Vav1-Cre⁺, and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient mice, three months post-transplant. n =10, **p<0.001, n.s. = no significant differences
- J) Histological analysis of *Carm1*^{F1/F1};Vav1-Cre⁻ and *Carm1*^{Δ/Δ};Vav1-Cre⁺ recipient mice. Scale bars represent 20 μm for the bone marrow, peripheral blood smears and bone marrow cytopspins. Scale bars for the spleen represents 100 μm (spleen and bone marrow) or 20 μm (peripheral blood smears and bone marrow cytopspins)
- K) PCR for the knockout of *Carm1* post poly(I:C) induction in *Carm1*^{+/+};Mx1-Cre⁺, *Carm1*^{Δ/+};Mx1-Cre⁺, and *Carm1*^{Δ/Δ};Mx1-Cre⁺ maintenance recipient mice pre and post induction of Cre by poly(I:C).
- L) Expression of *Carm1* in *Carm1*^{Δ/Δ};Mx1-Cre⁺ maintenance recipient mice normalized to *Gapdh* and compared to *Carm1*^{+/+};Mx1-Cre⁺ recipient mice.
- M) Comparison of CARM1 protein expression in fetal liver cells transduced with the MIGR1 vector alone, or MIGR1 AML1-ETO (AE), AML1-ETO9a (AE9a), MLL-AF9, or BCR-ABL. β-ACTIN is shown as a loading control.
- All error bars represent the mean ± SD. Statistics represent a Student's t-test for samples of unequal variance.

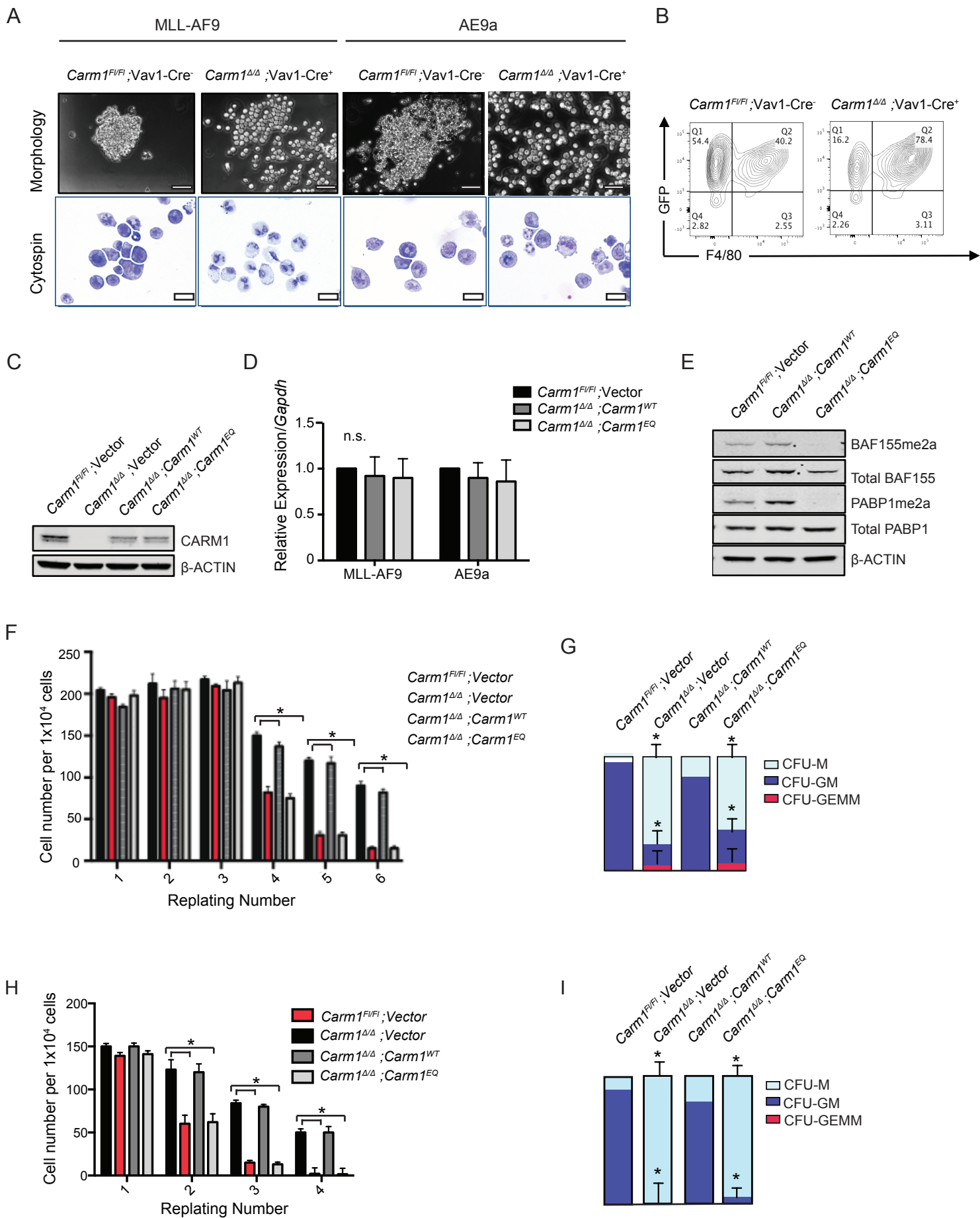
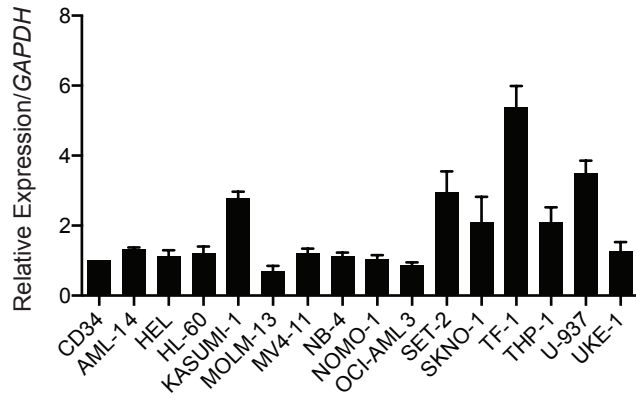


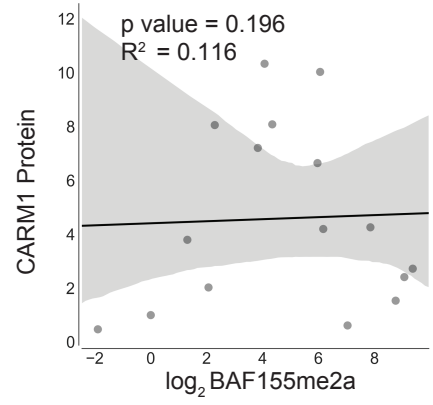
Figure S3: In vitro rescue of CARM1, Related to Figure 3.

- A) Morphological analysis of colony formation (top) and colony cytopspins (bottom) from MLL-AF9 and AE9a $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells cultured in M3434 methylcellulose. Scale bar (top) = 50 μm , Scale bar (bottom) = 20 μm
- B) Representative flow cytometry plot for GFP and the macrophage cell differentiation marker F4/80 in the AE9a $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells after the second replating.
- C) CARM1 protein levels following transduction of $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$. β -ACTIN is shown as a loading control.
- D) $Carm1$ mRNA levels following transduction of $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$, normalized to *Gapdh*. n.s.= no significant differences
- E) Western blot analysis of CARM1 enzymatic activity following transduction of $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$. β -ACTIN is shown as a loading control.
- F) CFU replating assay of MLL-AF9 expressing $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells transduced with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$. * $p < 0.01$
- G) Average colony percentage of MLL-AF9 expressing $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells transduced with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$, scoring CFU-M, CFU-GM, and CFU-GEMM from triplicate CFU experiments after replating 4. * $p < 0.01$
- H) CFU replating assay of AE9a expressing $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells transduced with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$. Error bars represent the mean \pm SD for three independent experiments. * $p < 0.01$
- I) Average colony percentage of AE9a expressing $Carm1^{F1/F1};Vav1-Cre^{-}$ and $Carm1^{\Delta/\Delta};Vav1-Cre^{+}$ fetal liver cells transduced with either vector alone, $Carm1^{WT}$ or $Carm1^{EQ}$ scoring CFU-M, CFU-GM, and CFU-GEMM from triplicate CFU experiments after replating 4. * $p < 0.01$
- All error bars represent the mean \pm SD. Statistics represent a Student's t-test for samples of unequal variance.

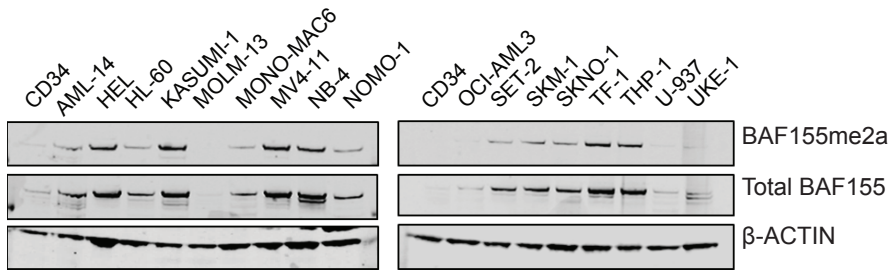
A



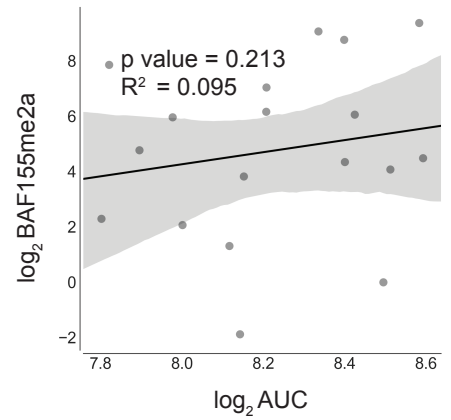
B



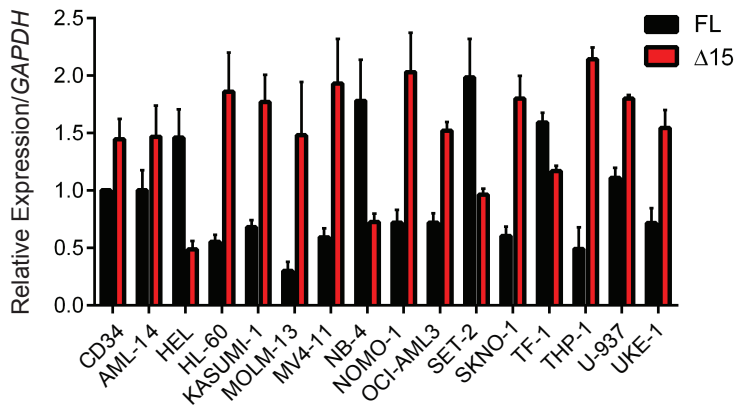
C



D



E



F

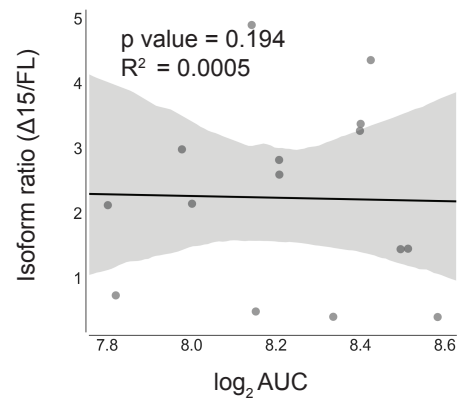


Figure S4. Comparison of CARM1 mRNA, isoform abundance, and target methylation in AML cell lines, Related to Figure 5.

A) Quantitative RT-PCR for *CARM1* mRNA expression in 16 AML cell lines and CD34⁺ cells. Error bars represent the mean \pm SD of three independent experiments. Statistics represent a Student's t-test for samples of unequal variance.

B) Quantification of log₂ BAF155me2a compared to *CARM1* protein expression in 16 AML cell lines and CD34⁺ cells. Spearman correlation and linear regression: p value = 0.196, R² = 0.116

C) Representative western blot showing levels of methylated BAF155 and total BAF155 in 18 AML cells lines and CD34⁺ cells. β -ACTIN is shown as a loading control.

D) Comparison of the log₂ AUC for 18 cell lines and CD34⁺ cells treated with EPZ025654 for 10 days vs. log₂ BAF155me2a. Spearman correlation and linear regression: p value = 0.213, R² = 0.1095

E) Quantification of *CARM1* full length (FL) and *CARM1* lacking exon 15 (Δ 15) by semi quantitative RT-PCR. Error bars represent the mean \pm SD from three independent samples from each cell line. Statistics represent a Student's t-test for samples of unequal variance.

F) Comparison of the log₂ AUC for 18 cell lines and CD34⁺ cells treated with EPZ025654 for 10 days vs. isoform ratios. Spearman correlation and linear regression: p value = 0.194, R² = 0.0005

Table S2. Top 100 differentially regulated genes in AML cell lines, Related to Figure 5.

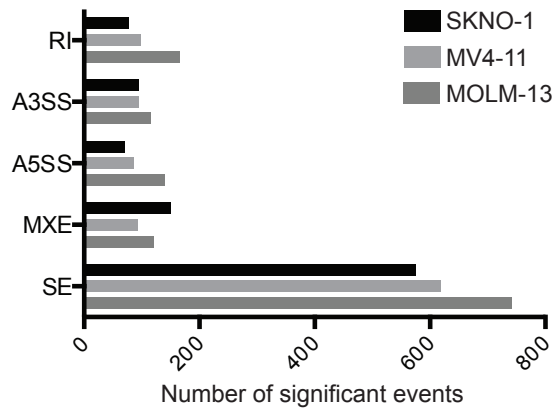
Gene ID	HGNC symbol	p value	Adjusted p value	log ₂ FoldChange
ENSG00000166803_KIAA0101	<i>KIAA0101</i>	2.84E-09	9.99E-07	-2.34
ENSG00000124635_HIST1H2BJ	<i>HIST1H2BJ</i>	6.63E-10	2.88E-07	-2.21
ENSG00000142453_CARM1	<i>CARM1</i>	2.23E-25	5.09E-21	-2.21
ENSG00000196584_XRCC2	<i>XRCC2</i>	3.26E-18	1.86E-14	-2.07
ENSG00000065328_MCM10	<i>MCM10</i>	2.87E-08	6.78E-06	-2.01
ENSG00000159259_CHAF1B	<i>CHAF1B</i>	1.00E-08	2.85E-06	-1.95
ENSG00000128408_RIBC2	<i>RIBC2</i>	2.45E-17	1.12E-13	-1.89
ENSG00000276043_UHRF1	<i>UHRF1</i>	2.54E-12	2.23E-09	-1.88
ENSG00000127564_PKMYT1	<i>PKMYT1</i>	1.06E-12	1.05E-09	-1.87
ENSG00000256663_RP11-424C20.2	<i>RP11-424C20.2</i>	4.49E-10	2.14E-07	-1.84
ENSG00000171320_ESCO2	<i>ESCO2</i>	6.83E-10	2.89E-07	-1.83
ENSG00000121211_MND1	<i>MND1</i>	2.35E-13	2.99E-10	-1.83
ENSG00000215784_FAM72D	<i>FAM72D</i>	1.68E-13	2.37E-10	-1.81
ENSG00000092470_WDR76	<i>WDR76</i>	1.87E-08	4.70E-06	-1.80
ENSG00000159055_MIS18A	<i>MIS18A</i>	3.61E-14	6.35E-11	-1.77
ENSG00000165490_DDIAS	<i>DDIAS</i>	3.38E-08	7.74E-06	-1.73
ENSG00000168078_PBK	<i>PBK</i>	8.20E-09	2.44E-06	-1.71
ENSG00000104738_MCM4	<i>MCM4</i>	8.81E-22	1.01E-17	-1.71
ENSG00000119969_HELLS	<i>HELLS</i>	2.41E-15	6.13E-12	-1.71
ENSG00000188610_FAM72B	<i>FAM72B</i>	5.32E-10	2.48E-07	-1.63
ENSG00000105011_ASF1B	<i>ASF1B</i>	3.70E-09	1.26E-06	-1.59
ENSG00000076003_MCM6	<i>MCM6</i>	4.81E-16	1.83E-12	-1.57
ENSG00000137812_KNL1	<i>KNL1</i>	9.85E-10	3.69E-07	-1.56
ENSG00000164109_MAD2L1	<i>MAD2L1</i>	5.34E-11	3.13E-08	-1.56
ENSG00000060982_BCAT1	<i>BCAT1</i>	2.57E-11	1.73E-08	-1.56
ENSG00000123219_CENPK	<i>CENPK</i>	1.86E-08	4.70E-06	-1.54
ENSG00000132436_FIGNL1	<i>FIGNL1</i>	4.86E-15	1.01E-11	-1.51
ENSG00000277224_HIST1H2BF	<i>HIST1H2BF</i>	1.05E-08	2.93E-06	-1.50
ENSG00000175279_CENPS	<i>CENPS</i>	1.21E-08	3.29E-06	-1.47
ENSG00000091651_ORC6	<i>ORC6</i>	3.87E-09	1.30E-06	-1.45
ENSG00000196550_FAM72A	<i>FAM72A</i>	6.14E-13	6.39E-10	-1.44
ENSG00000012048_BRCA1	<i>BRCA1</i>	6.42E-15	1.22E-11	-1.44
ENSG00000111445_RFC5	<i>RFC5</i>	1.76E-13	2.37E-10	-1.44
ENSG00000198056_PRIM1	<i>PRIM1</i>	3.06E-10	1.52E-07	-1.43
ENSG00000146918_NCAPG2	<i>NCAPG2</i>	2.80E-12	2.37E-09	-1.43
ENSG00000162062_C16orf59	<i>C16orf59</i>	1.61E-08	4.24E-06	-1.39
ENSG00000122952_ZWINT	<i>ZWINT</i>	5.36E-09	1.78E-06	-1.35
ENSG00000164087_POC1A	<i>POC1A</i>	2.93E-13	3.53E-10	-1.33
ENSG00000167325_RRM1	<i>RRM1</i>	9.71E-16	2.77E-12	-1.31
ENSG00000058804_NDC1	<i>NDC1</i>	1.11E-11	8.74E-09	-1.31
ENSG00000109881_CCDC34	<i>CCDC34</i>	1.03E-11	8.38E-09	-1.30
ENSG00000112118_MCM3	<i>MCM3</i>	6.06E-09	1.90E-06	-1.30
ENSG00000177602_GSG2	<i>GSG2</i>	9.96E-09	2.85E-06	-1.29
ENSG00000097046_CDC7	<i>CDC7</i>	1.83E-11	1.27E-08	-1.28
ENSG00000153044_CENPH	<i>CENPH</i>	2.47E-10	1.25E-07	-1.26

Gene ID	HGNC symbol	p value	Adjusted p value	log ₂ FoldChange
ENSG00000156802_ATAD2	<i>ATAD2</i>	1.64E-10	8.52E-08	-1.25
ENSG00000149636_DSN1	<i>DSN1</i>	3.05E-15	6.98E-12	-1.24
ENSG00000112029_FBXO5	<i>FBXO5</i>	1.33E-08	3.54E-06	-1.22
ENSG00000213585_VDAC1	<i>VDAC1</i>	1.13E-09	4.09E-07	-1.21
ENSG00000197299_BLM	<i>BLM</i>	1.51E-11	1.12E-08	-1.21
ENSG00000104889_RNASEH2A	<i>RNASEH2A</i>	9.14E-10	3.52E-07	-1.21
ENSG00000112312_GMNN	<i>GMNN</i>	2.07E-12	1.97E-09	-1.20
ENSG00000158169_FANCC	<i>FANCC</i>	5.23E-14	8.54E-11	-1.19
ENSG00000040275_SPDL1	<i>SPDL1</i>	9.24E-10	3.52E-07	-1.18
ENSG00000146263_MMS22L	<i>MMS22L</i>	1.01E-08	2.85E-06	-1.18
ENSG00000144554_FANCD2	<i>FANCD2</i>	1.84E-08	4.70E-06	-1.13
ENSG00000136824_SMC2	<i>SMC2</i>	1.73E-11	1.24E-08	-1.13
ENSG00000176974_SHMT1	<i>SHMT1</i>	4.30E-11	2.66E-08	-1.10
ENSG00000077514_POLD3	<i>POLD3</i>	3.82E-19	2.91E-15	-1.09
ENSG00000178966_RMI1	<i>RMI1</i>	7.55E-16	2.47E-12	-1.08
ENSG00000166881_NEMP1	<i>NEMP1</i>	5.87E-09	1.88E-06	-1.06
ENSG00000120802_TMPO	<i>TMPO</i>	6.06E-11	3.47E-08	-1.05
ENSG00000156876_SASS6	<i>SASS6</i>	8.69E-10	3.52E-07	-1.05
ENSG00000106399_RPA3	<i>RPA3</i>	1.20E-10	6.36E-08	-1.04
ENSG00000075131_TIPIN	<i>TIPIN</i>	2.67E-11	1.75E-08	-1.01
ENSG00000080839_RBL1	<i>RBL1</i>	1.22E-13	1.86E-10	-0.99
ENSG00000172009_THOP1	<i>THOP1</i>	1.26E-08	3.40E-06	-0.98
ENSG00000101868_POLA1	<i>POLA1</i>	6.16E-10	2.76E-07	-0.97
ENSG00000198826_ARHGAP11A	<i>ARHGAP11A</i>	8.96E-10	3.52E-07	-0.96
ENSG00000184445_KNTC1	<i>KNTC1</i>	3.60E-10	1.75E-07	-0.96
ENSG00000106462_EZH2	<i>EZH2</i>	4.53E-11	2.73E-08	-0.95
ENSG00000204899_MZT1	<i>MZT1</i>	2.10E-08	5.16E-06	-0.93
ENSG0000014138_POLA2	<i>POLA2</i>	3.55E-11	2.26E-08	-0.91
ENSG00000213390_ARHGAP19	<i>ARHGAP19</i>	6.19E-09	1.91E-06	-0.88
ENSG00000166483_WEE1	<i>WEE1</i>	2.97E-08	6.93E-06	-0.85
ENSG00000138780_GSTCD	<i>GSTCD</i>	2.36E-12	2.16E-09	-0.84
ENSG00000105486_LIG1	<i>LIG1</i>	1.19E-10	6.36E-08	-0.83
ENSG00000171208_NETO2	<i>NETO2</i>	1.92E-08	4.77E-06	-0.81
ENSG00000079616_KIF22	<i>KIF22</i>	3.45E-09	1.19E-06	-0.79
ENSG00000166147_FBN1	<i>FBN1</i>	1.82E-09	6.50E-07	-0.79
ENSG00000163781_TOPBP1	<i>TOPBP1</i>	3.89E-13	4.45E-10	-0.77
ENSG00000111775_COX6A1	<i>COX6A1</i>	5.51E-10	2.52E-07	-0.71
ENSG00000120539_MASTL	<i>MASTL</i>	2.24E-08	5.38E-06	-0.69
ENSG00000198924_DCLRE1A	<i>DCLRE1A</i>	1.14E-08	3.14E-06	-0.64
ENSG00000129484_PARP2	<i>PARP2</i>	5.93E-09	1.88E-06	-0.64
ENSG00000232653_GOLGA8N	<i>GOLGA8N</i>	6.92E-09	2.11E-06	-0.57
ENSG00000143315_PIGM	<i>PIGM</i>	2.16E-08	5.25E-06	-0.44
ENSG00000142687_KIAA0319L	<i>KIAA0319L</i>	3.22E-08	7.43E-06	0.50
ENSG00000060971_ACAA1	<i>ACAA1</i>	2.30E-08	5.48E-06	0.57
ENSG00000112679_DUSP22	<i>DUSP22</i>	5.16E-13	5.62E-10	0.70

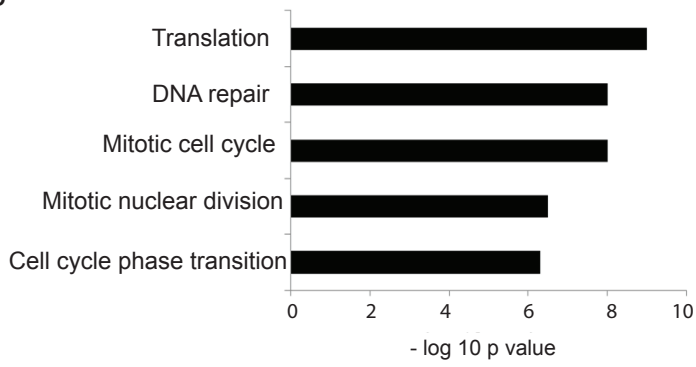
Gene ID	HGNC symbol	p value	Adjusted p value	log ₂ FoldChange
ENSG00000243749_TM35B	<i>TM35B</i>	8.82E-10	3.52E-07	0.87
ENSG00000163956_LRPAP1	<i>LRPAP1</i>	8.43E-09	2.47E-06	0.88
ENSG00000161011_SQSTM1	<i>SQSTM1</i>	1.11E-09	4.09E-07	0.89
ENSG00000167996_FTH1	<i>FTH1</i>	8.49E-10	3.52E-07	1.41
ENSG00000177606_JUN	<i>JUN</i>	6.68E-10	2.88E-07	1.58
ENSG00000185909_KLHDC8B	<i>KLHDC8B</i>	7.14E-09	2.15E-06	2.04
ENSG00000100292_HMOX1	<i>HMOX1</i>	5.92E-09	1.88E-06	2.33
ENSG00000244734_HBB	<i>HBB</i>	1.26E-11	9.57E-09	6.09

* Red font represents gene associated with Gene Ontology Term 'Cell Cycle Progression'*

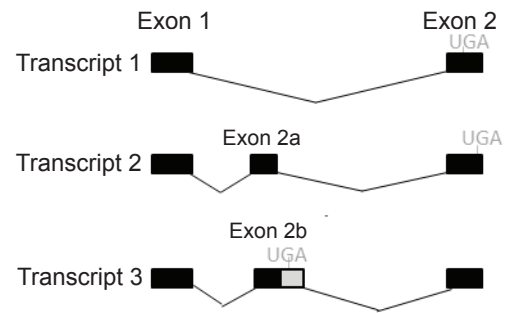
A



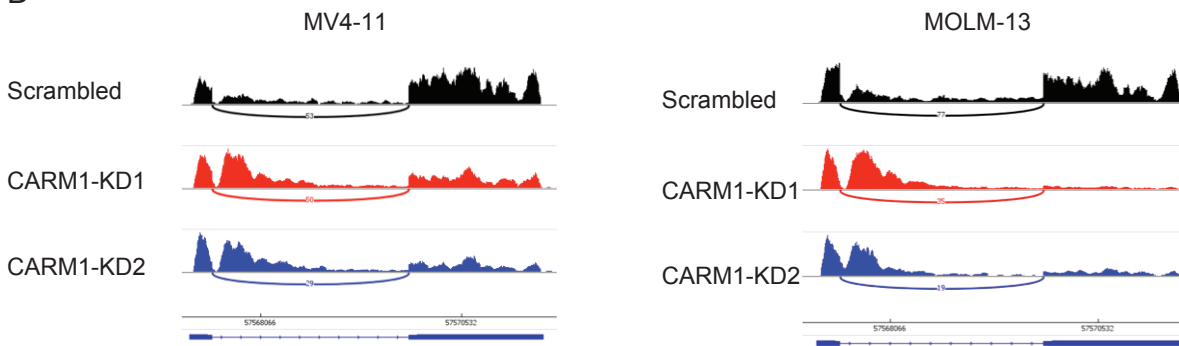
B



C



D



E

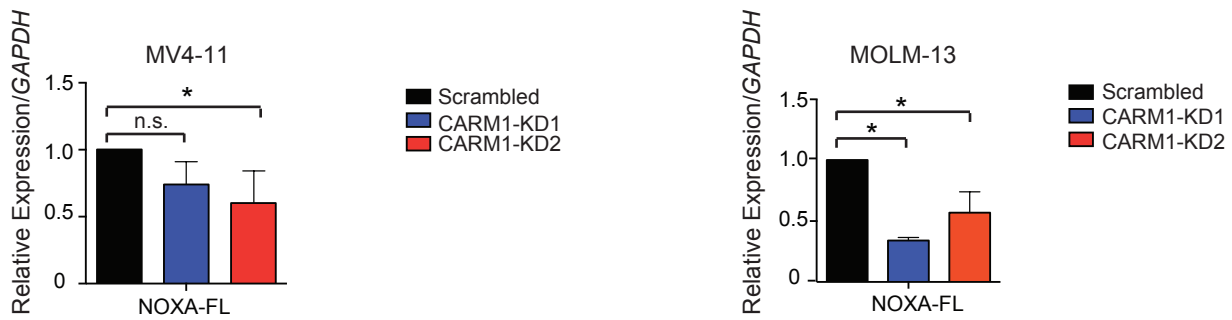
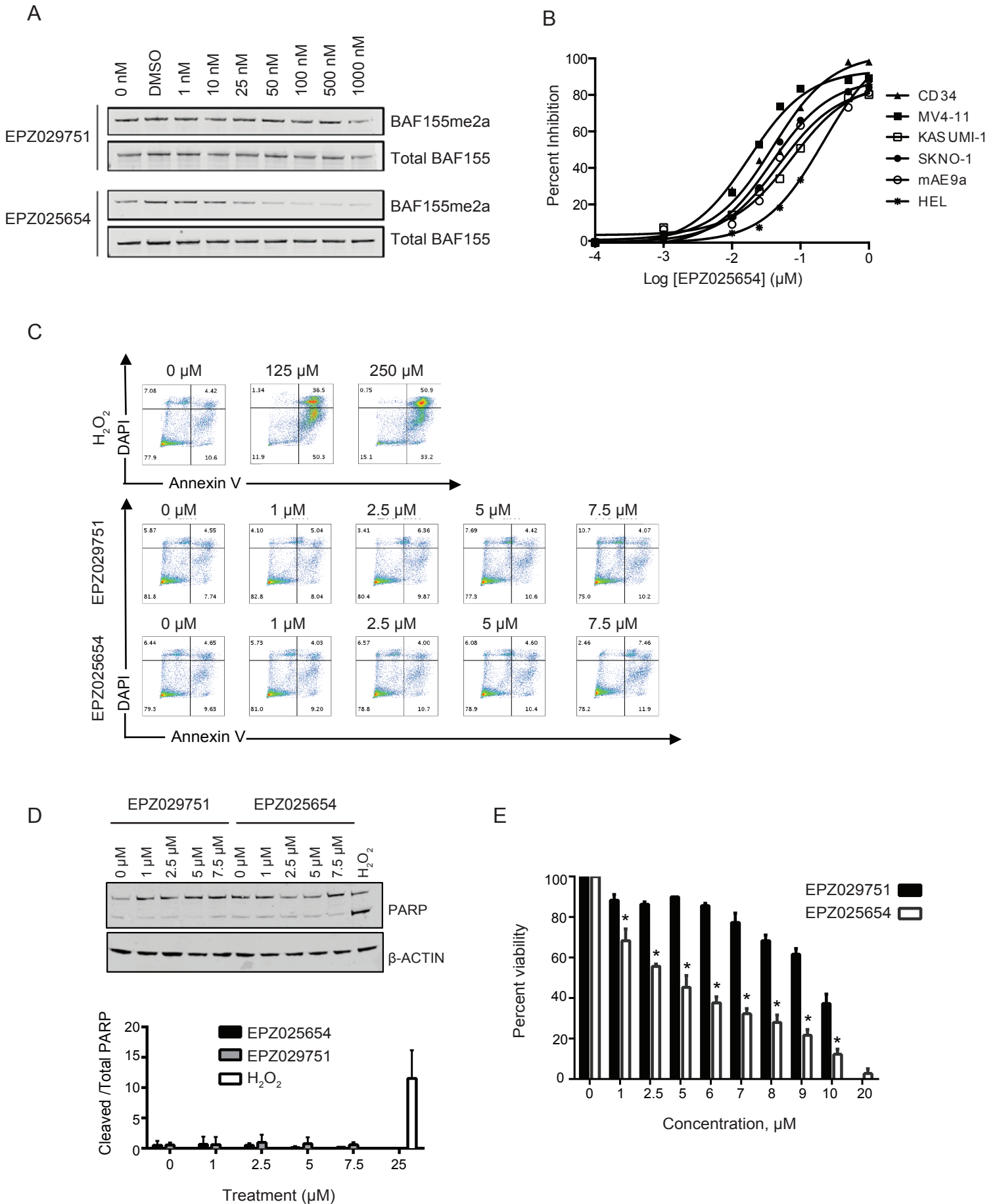
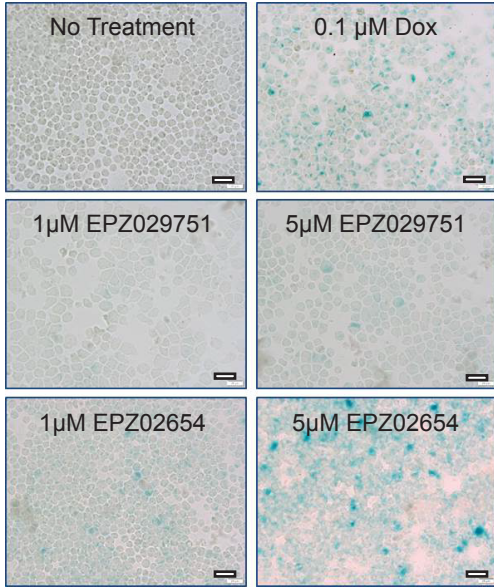


Figure S5: Characterization of alternative splicing in leukemia cell lines in response to CARM1 knockdown, Related to Figure 5.

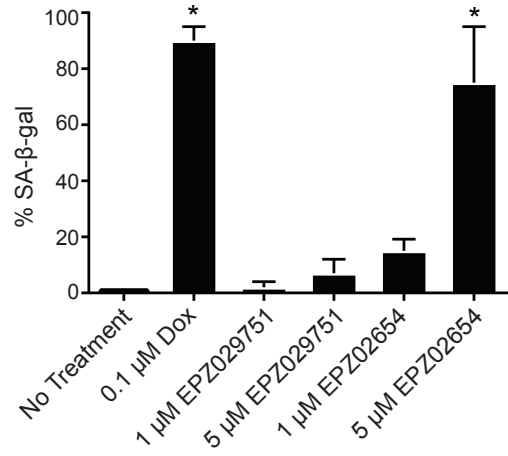
- A) Number of significant alternative splicing events for the SKNO-1, MV4-11, and MOLM-13 cells lines. Events are annotated as Retained Intron (RI), Alternative 3' Splice Site (A3SS), Alternative 5' Splice Site (A5SS), Mutually Exclusive Exons (MXE), or Skipped Exon (SE). Events were considered significant if present in both CARM1 KD samples and not the scrambled control for each cell line.
- B) Gene ontology (GO) analysis of significant skipped exon events common to all three cell lines. Bars represent the $-\log_{10}$ p value.
- C) Schematic of PMAIP transcripts.
- D) Sashimi plot for PMAIP1/NOXA in Scrambled, CARM1-KD1, or CARM1-KD2 MV4-11 (left) and MOLM-13 (right) cells.
- E) Full length *PMAIP/NOXA* mRNA expression in Scrambled, CARM1-KD1, or CARM1-KD2 sorted MV4-11 (left) and MOLM-13 (right) cells, normalized to *GAPDH*. Results represent the mean \pm SD of three independent experiments. n=3, *p<0.05
Statistics represent a Student's t-test for samples of unequal variance.



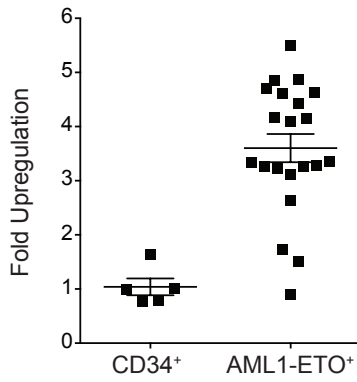
F



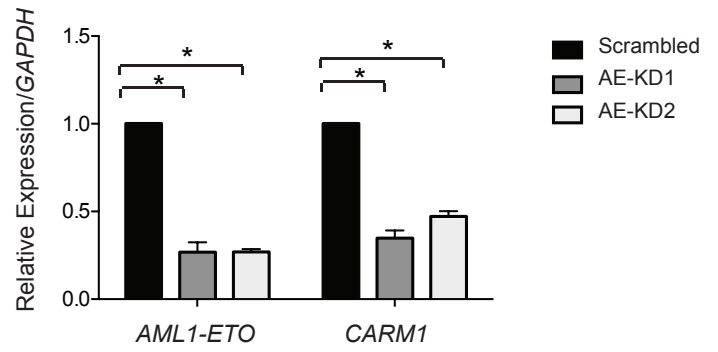
G



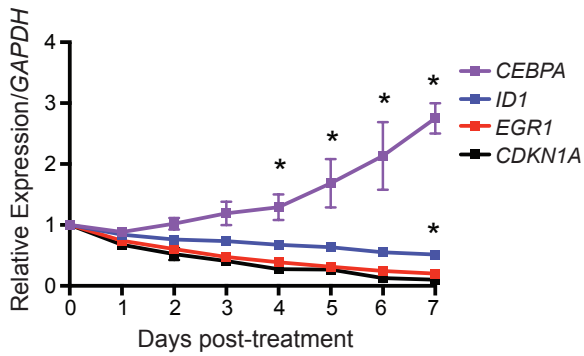
H



I



J



K

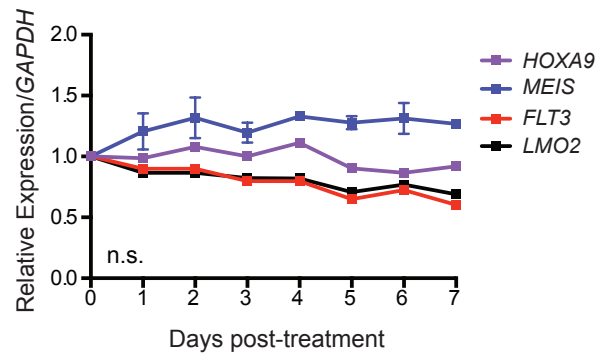


Figure S6. Biological comparison of pharmacological inhibition of EPZ025654 and EPZ029751, Related to Figure 6.

- A) Quantification of immunoblots showing the percent inhibition of BAF155me2a vs. BAF155 total protein expression in SKNO-1 cells after 48 hours of treatment with increasing doses of the active CARM1 inhibitor (EPZ025654) and control compound(EPZ029751).
- B) Log[EPZ05654] compared to the percent inhibition of the asymmetric methylation of BAF155 in CD34⁺, MV4-11, KASUMI-1, SKNO-1, HEL, or mouse AE9a cells treated for 48 hours. Data points represent the quantification of BAF155me2a and total BAF155 by western blot. Lines represent the non-linear regression fit with variable slope.
- C) Comparison of the effects of EPZ025654 or EZP029751 on Annexin V/DAPI positivity after 7 days of treatment. Cells treated with 125 μ M or 250 μ M of H₂O₂ are shown as a positive control for apoptosis.
- D) Western blot comparing the effects of EPZ025654 or EZP029751 on PARP cleavage in SKNO-1 cells after 7 days of treatment (top). Cells treated with 125 μ M of H₂O₂ are shown as a positive control. β -ACTIN is shown as a loading control. Quantification of Cleaved/Total PARP for three independent experiments (bottom).
- E) Comparison of the effects of EPZ025654 or EZP029751 on cell viability after 7 days of treatment as measured by Cell Titer Glo. n = 3, *p < 0.01
- F) IHC for senescence associated β -galactosidase in SKNO-1 cells treated with 1 μ M or 5 μ M of EPZ025654 for 6 days. Cells with no treatment, identical doses of the control compound EPZ02975, or Doxorubicin (Dox) are shown for comparison. Scale bar = 20 μ m
- G) Quantification of percentage senescence associated β -galactosidase positive SKNO-1 cells treated with EPZ025654 or EPZ029751 for 6 days. Doxorubicin (Dox) is shown as a positive control for senescence. Scale bar = 20 μ m, n=2, *p < 0.01.
- H) Comparison of *CARM1* expression in AML1-ETO⁺ AML patients in the Eastern Cooperative Oncology Group (ECOG) cohort compared to normal CD34⁺ controls.
- I) Comparison of *CARM1* expression following the KD of AML1-ETO in the t(8;21) positive cell line, Kasumi-1. n=3, *p < 0.01
- J) Evaluation of AML1-ETO specific gene regulation (*CEBPA*, *ID1*, *EGR1*, and *CDKN1A*) over time in SKNO-1 cells treated with 5 μ M EPZ025654 for 7 days. Expression is relative to *GAPDH*. n=3, *p < 0.01
- K) Evaluation of MLL-AF9 specific gene regulation (*HOXA9*, *MEIS1*, *FLT3*, and *LMO2*) over time in MV4-11 cells treated with 5 μ M EPZ025654 for 7 days. Expression is relative to *GAPDH*. n=3, n.s. = no significant differences
- For all bar graphs, error bars represent the mean \pm SD. Statistics represent a Student's t-test for samples of unequal variance.

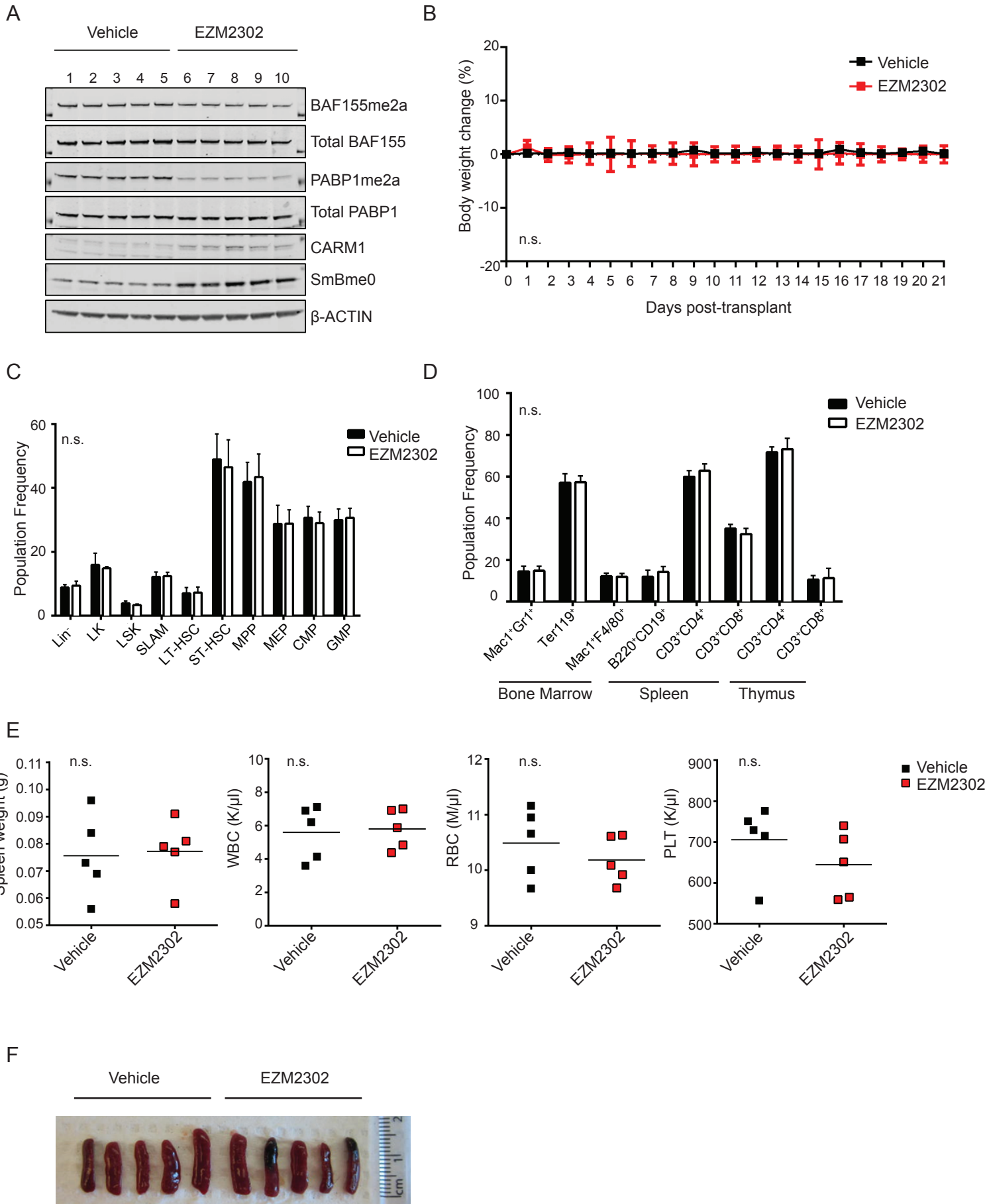


Figure S7. Evaluation of in vivo CARM1 inhibitor treatment, Related to Figure 7.

A) Western blot analysis for BAF155 and PABP1 asymmetric methylation in the spleen of mice treated with either vehicle or 100 mg/kg EZM2302 for three weeks. β -ACTIN is shown as a loading control. n = 5

B) Daily percentage body weight change for mice treated with either vehicle or 100 mg/kg EZM2302 for three weeks. n = 5, n.s.= not statistically significant

C) Frequency of hematopoietic stem and progenitor populations from mice treated with either vehicle or 100 mg/kg EZM2302 for three weeks. n = 5, n.s. = not statistically significant

D) Frequency of mature hematopoietic populations in the bone marrow, spleen, and thymus, from mice treated with either vehicle or 100 mg/kg EZM2302 for three weeks. n= 5, n.s. = not statistically significant

E) Comparison of spleen weights, peripheral blood white blood cell (WBC), red blood cell (RBC), and platelet (PLT) counts from mice treated with either vehicle or 100 mg/kg EZM2302 for three weeks. n = 5, n.s. = not statistically significant

F) Image showing spleen morphology of mice treated with either vehicle or 100 mg/kg EZM2302 for three weeks. n = 5

All error bars represent the mean \pm SD. Statistics represent a Student's t-test for samples of unequal variance.

Table S4. IC₅₀ of leukemia cell lines treated with EPZ025654 for 10 days, Related to Figure 7.

Cell line	Viability EPZ025654	Media	Age	Sex	Reported mutations (CCLE/COSMIC/Tru Myeloid)	FAB ^a	Fusion Oncoprotein
CD34 ⁺	>10 μ M	X-vivo +20% BIT	N/A	N/A	N/A	N/A	N/A
AML-14	>10 μ M	RPMI 10%	68	M	TP53, NRAS	M2	
HEL	>10 μ M	RPMI 10%	30	M	JAK2V617F, TET2, KMT2D, TP53, EP300	M6	
HL-60	6.3 μ M	IMDM 20%	36	F	CDKN2A, NRAS, SF3B1	M2	
KASUMI-1	1.4 μ M	RPMI 20%	7	M	C-KIT, TP53, ASXL1, CBP, RAD21	M2	AML1-ETO
MOLM-13	1.1 μ M	RPMI 10%	20	M	FLT3/ITD, CBL, KMT2A, KMT2C, NF1, SETD1A	M5	MLL-AF9
MONO-MAC6	>10 μ M	RPMI 10%	64	M	FLT3/ITD, KAT6B, RUNX1, ARID2A, ASXL1, IDH1, U2AF1	M5	MLL-AF9
MV4-11	3.0 μ M	IMDM 10%	10	M	FLT3/ITD, NPM1	M5	MLL-AF4
NB-4	2.3 μ M	RPMI 10%	23	F	KRAS, TP53, ETO, EP300, MOZ, ASXL2	M3	PML-RARA
NOMO-1	1.1 μ M	RPMI 10%	31	F	KRAS, ASXL1, EP300, SF3B1, TP53, CDKN1B, KMT2C	M5	MLL-AF9
OCI-AML3	0.1 μ M	alpha-MEM 20% FBS	57	M	NPM1, NRAS, DNMT3A	M4	
SET-2	7.8 μ M	RPMI 20%	71	F	JAK2V617F, TP53, UTX, DNMT3A	N.D.	
SKNO-1	0.4 μ M	RPMI 10% +GM-CSF	22	M	TP53, KDM6B, KRAS, TET2, ASXL1	M2	AML1-ETO
SKM-1	2.2 μ M	RPMI 20%	76	M	TP53, c-KIT	M5	
TF-1	0.2 μ M	RPMI 10%+ IL-3	35	M	TP53, NRAS	M6	
THP-1	6.3 μ M	RPMI 10%	1	M	CDKN2A, NRAS, TP53, ARID1A, UTX	M5	MLL-AF9
U-937	1.0 μ M	RPMI 10%	37	M	TP53, WT1, PTEN, ETV6, PTPN11	M5	
UKE-1	3.5 μ M	IMDM 10% +HS+HC	59	F	JAK2V617F, IKZF1, EZH2, ETV6, PTPN11, STAG2	N.D.	

^aFAB: French-American-British classification system

Table S6. Primers used in study, Related to STAR methods

Probe	Source	Identifier
<i>GAPDH</i> (human)	Thermo Fisher	Hs02758991_g1
<i>CARM1</i> (human)	Thermo Fisher	Hs00406354_m1
<i>MEIS1</i> (human)	Thermo Fisher	Hs01017441_m1
<i>HOXA9</i> (human)	Thermo Fisher	Hs00365956_m1
<i>FLT3</i> (human)	Thermo Fisher	Hs00174690_m1
<i>LMO2</i> (human)	Thermo Fisher	Hs00153473_m1
<i>E2F1</i> (human)	Thermo Fisher	Hs00153451_m1
<i>E2F2</i> (human)	Thermo Fisher	Hs00231667_m1
<i>CENPA</i> (human)	Thermo Fisher	Hs0015655_m1
<i>CDC25A</i> (human)	Thermo Fisher	Hs00947994_m1
<i>MYBL2</i> (human)	Thermo Fisher	Hs00942543_m1
<i>TOP2A</i> (human)	Thermo Fisher	Hs01032137_m1
<i>UHRF1</i> (human)	Thermo Fisher	Hs00380204_m1
<i>E2F1</i> (human)	Thermo Fisher	Hs00153451_m1
<i>E2F2</i> (human)	Thermo Fisher	Hs00231667_m1
<i>E2F3</i> (human)	Thermo Fisher	Hs00605457_m1
<i>E2F4</i> (human)	Thermo Fisher	Hs00608098_m1
<i>E2F5</i> (human)	Thermo Fisher	Hs00231092_m1
<i>E2F6</i> (human)	Thermo Fisher	Hs01034552_m1
<i>E2F7</i> (human)	Thermo Fisher	Hs00403170_m1
<i>E2F8</i> (human)	Thermo Fisher	Hs00226635_m1
<i>TP53</i> (human)	Thermo Fisher	Hs01034249_m1
<i>CDKN1A</i> (human)	Thermo Fisher	Hs00355782_m1
<i>Gapdh</i> (mouse)	Thermo Fisher	Mm99999915_g1
<i>Carm1</i> (mouse)	Thermo Fisher	Mm01171448_m1