Diogo et al.

SUPPLEMENTARY INFORMATION

Supplementary Methods

Gene selection

A short description of the selected candidate target genes and level of genetic and/or biological support for a link to common human disease is provided. All SNPs selected for PheWAS in this study (bold) had reported genome-wide significant GWAS associations (P<5e-8) in the European population.

- *GPR65 GPR65* encodes the G protein coupled receptor 65, also known as T-cell death associated gene 8 (TDAG8). GWAS have identified SNPs at the *GPR65* locus as associated with CD and UC^3 . The lead SNP, rs8005161, is in prefect LD with the *GPR65* missense SNP **rs3742704** (p.I231L). In functional studies, I231L was shown to alter GPR65 signaling and to affect lysosomal pH, leading to lysosomal dysfunction and impaired bacterial restriction¹⁹.
- *IFIH1 IFIH1* encodes the Interferon-induced helicase C domain-containing protein 1, also known as Melanoma differentiation-associated protein 5 (MDA5), a cytoplasmic sensor of viral nucleic acids that plays a major role in the activation of a cascade of antiviral responses including the induction of type I interferons and pro-inflammatory cytokines²⁰. Rare gain-of-function (GOF) mutations in *IFIH1* have been reported to cause Aicardi-Goutières syndrome and Singleton-Merten syndrome 1, two syndromes characterized by high interferon signaling²¹⁻ 23. Conversely, rare loss-of-function (LOF) variants that lead to severe disruption of *IFIH1* signaling function have been recently shown to cause a primary immunodeficiency manifested in extreme susceptibility to common respiratory RNA viruses ^{24, 25}. In addition, an allelic series of rare, low-frequency and common LOF alleles (rs35744605, **rs35667974**, **rs1990760**) has been shown to protect against multiple autoimmune diseases, including type 1 diabetes (T1D), vitiligo, lupus and psoriasis, while rs1990760 and rs35667974 were found associated with increased risk of ulcerative colitis 26-36. The rare T1D protective rs35744605 (p.E627X) and rs35667974 (p.I923V) alleles confer a significant inhibition of IFNβ production, and the common T1D protective allele rs1990760 (p.T946A) has been shown to reduce MDA5 expression following IFNβ treatment^{27, 32}. Furthermore, mice heterozygote or homozygote for the rs1990760 T1D risk allele exhibit higher interferon signature, enhanced resistance to viral challenge, and higher incidence of streptozocin-induced T1D 37 . Based on the genetic findings, it is hypothesized that MDA5 inhibition might be of therapeutic benefit for patients diagnosed with several autoimmune diseases.
- *IRF5* The interferon regulatory factor 5 (IRF5) is involved in the activation of type I interferon signaling in response to viral infection. Common GWAS SNPs are associated with multiple autoimmune diseases. Two independent signals at the locus, driven by **rs10488631** and **rs2004640**, are associated with rheumatoid arthritis (RA), primary biliary cirrhosis (PBC), systemic lupus erythematosus (SLE), and Sjogren's syndrome35, 38-40. In addition, a SNP in LD with rs10488631 (rs62478615) was reported associated with systemic sclerosis41, and **rs3823536** (in LD with rs2004640) has been reported associated with ulcerative colitis3 . Multiple studies evaluating the function of rs2004640 have shown the autoimmune risk allele to increase IRF5 mRNA and protein levels in immune cells, upregulate IFN α -induced genes, and increase pro-inflammatory cytokine production by monocyte-derived dendritic cells and monocyte-derived macrophages⁴²⁻⁴⁴. Based on the genetic findings, it is hypothesized that IRF5 inhibition might be of therapeutic benefit for patients diagnosed with autoimmune diseases.

TYK2 The Tyrosine kinase 2 (TYK2) is a member of the Janus kinase (JAK) family of proteins that mediates signaling downstream of several cytokine receptors. TYK2 signaling deficient patients carrying rare *TYK2* LOF mutations and characterized by immunodeficiency have been reported^{45, 46}. In GWAS, independent lowfrequency (**rs34536443**) and common (**rs12720356**) *TYK2* missense SNPs have been associated with a variety of autoimmune diseases, including RA, SLE, IBD, Psoriasis, and multiple sclerosis $(MS)^{3, 35, 38, 47, 49}$. The protective alleles have been reported to decrease TYK2 kinase activity and downstream signaling in immune $cells^{50, 51}$. It is hypothesized that TYK2 inhibition might be of therapeutic benefit for patients diagnosed with autoimmune diseases.

B. Immunity

CDHR3 The cadherin-related family, member 3 (CHDR3) transmembrane protein is involved in cell adhesion, epithelial polarity, cell-cell interaction and differentiation, and is predominantly expressed in respiratory epithelium. A GWAS in Danish asthma patients identified the coding SNP **rs6967330** (p.C529Y) in *CDHR3* as associated with recurrent childhood asthma exacerbation⁵². The disease-associated SNP locates to the protein's extracellular domain and is associated with increased CDHR3 cell surface expression. This SNP was shown to stimulate cell internalization of rhinovirus C, a known stimulus for asthma exacerbation, in cultured cells⁵³. It is hypothesized that inhibition of CDHR3 might reduce rhinovirus C internalization and virus-induced asthmarelated symptoms.

C. Cardiovascular & metabolic diseases

- *F11* Factor XI is a serine protease within the contact pathway of the coagulation cascade. In humans, Factor XI is encoded by *F11*. Deficiency of factor XI causes the rare condition hemophilia C which affects up to 8% of the Ashkenazi Jewish population54. Homozygotes and compound-heterozygotes for *F11* LOF mutations show little spontaneous bleeding, but surgical procedures may cause excessive blood loss, requiring prophylaxis. High levels of factor XI have been proposed to increase the risk of thrombosis. Common SNPs near *F11*, including rs4253417, rs1593, and **rs4253399**, have been associated with altered Factor XI plasma levels, activated partial thromboplastin time (aPTT) and venous thromboembolism risk $55-57$. FXI-reducing drugs are under development for the prevention of venous thrombosis⁵⁸.
- *F12* Coagulation factor XII, (Hageman factor) is a plasma serine protease and a zymogen form of factor XIIa, which is encoded by the *F12* gene. Factor XII is part of the coagulation cascade and activates factor XI and prekallikrein *in vitro*. Factor XII itself is activated by negatively charged surfaces and is the starting point of the intrinsic pathway. Factor XII deficiency is rare and inherited in an autosomal recessive manner^{59,60}. Typically, Factor XII deficiency is asymptomatic and, unlike FXI deficiency, does not result in excess bleeding. A suggested GOF mutation in $F12$ has been reported in patients with hereditary angioedema 61 . Knockout mice are less susceptible to thrombosis⁶². Factor XII appears to be involved also in the later stages of clot formation. Common variants in *F12*

(rs2545801, **rs2731672**) have been associated with Factor XII levels, aPTT and plasma levels of various vasoactive substances including endothelin, bradykinin, adrenomedullin, chromogranin, and phenylalanine^{63, 64}. Inhibition of Factor XII has been considered a therapeutic strategy for the prevention of thromboembolism.

- *GDF15* Growth differentiation factor-(GDF-)15 is a stress-responsive cytokine emerging as a biomarker of cardiac and vascular dysfunction and disease. Elevated circulating levels of GDF-15 identify high-risk individuals across the cardiovascular continuum, from stable coronary artery disease to acute coronary syndrome and heart failure. The association of GDF-15 with outcome in these conditions is independent of clinical risk factors and established biomarkers, including NT-proBNP (N-terminal pro-B-type natriuretic peptide) and troponin. Common SNPs associated with BMI by the GIANT consortium have been identified in the proximal region of the *GDF15* gene⁶⁵. The lead SNP **rs17724992** is an intronic SNP in *PGPEP1* where the minor allele associated with decreased BMI. *GDF15* was annotated as a gene with biological relevance to this locus in the original publication, but eQTL data does not support a direct functional effect on *GDF15* mRNA levels 66. Increased serum levels of GDF-15 have been reported associated with type 2 diabetes (T2D) and impaired fasting glucose 67 .
- *GUCY1A3* Soluble guanylate cyclase (sGC) is an intracellular enzyme that mediates the nitric oxide (NO) signaling pathway. It catalyzes synthesis of the second messenger cyclic guanosine monophosphate (cGMP), which leads to vasorelaxation and inhibits smooth muscle proliferation, leukocyte recruitment and platelet aggregation through a number of downstream mechanisms. In humans, sGC subunits are encoded by the genes *GUCY1A2*, *GUCY1A3*, *GUCY1B2* and *GUCY1B3*. An sGC activator, Ricoguat, is a marketed therapeutic in cardiopulmonary disease. Common SNPs near *GUCY1A3* (**rs13139571**) have been significantly associated with diastolic blood pressure, and suggestively associated with systolic blood pressure and hypertension⁶⁸. The same SNPs are also associated with coronary artery disease and myocardial infarction⁶⁸. rs13139571 is an intronic SNP in *GUCY1A3* and an eQTL for *GUCY1A3* in multiple tissues. Knockout of *GUCY1A3* or *GUCY1B3* in murine models results in hypertension⁶⁹. Truncating mutations have been observed in patients with moyamoya disease with achalasia⁷⁰.
- *KNG1* Kininogen-1, encoded by the KNG1 gene, is the precursor protein to highmolecular-weight kininogen (HMWK), low-molecular-weight kininogen (LMWK), and bradykinin. Alternative splicing of the *KNG1* gene leads to either low-molecular-weight kininogen (LMWK) or high-molecular-weight kininogen (HMWK) which can be cleaved to produce bradykinin. HMWK is essential for blood coagulation and assembly of the kallikrein-kinin system. HMWK interacts with Factor XII, Factor XI and prekallikrein where it acts as a cofactor to initiate the contact activation (intrinsic) pathway of coagulation. HMWK is not enzymatically active, but functions as a cofactor for the activation of kallikrein, factor XII and activation of factor XI by factor XIIa. GWAS have identified SNPs at the *KNG1* locus (**rs5030062** and rs710446, both in strong LD) that associate with des-arg(9) bradykinnin levels, Factor XI plasma levels and a $PTT₅₅₋₅₇$.
- *LGALS3* Galectin-3 (Gal-3) is a multifunctional protein of an expanding family of βgalactoside–binding animal lectins, mainly produced by macrophages and monocytes. It is implicated in a variety of biologic events, such as inflammation, angiogenesis and fibrotic disorders. Gal-3 is broadly expressed in various tissues and secreted into the extracellular matrix and circulation. Increased circulating levels of galectin-3 have been associated with various diseases, including cancer, immunological disorders, and heart failure. A GWAS of plasma galectin-3 levels identified **rs2274273** (a non-coding variant in LD with missense variants in *LGALS3*) as associated with reduced levels of circulating galectin-371. No specific disease associations have been reported for this variant.
- *PNPLA3* Patatin-like phospholipase domain-containing protein 3 (PNPLA3) also known as adiponutrin, acylglycerol O-acyltransferase or calcium-independent phospholipase A2-epsilon (iPLA2-epsilon), is an enzyme primarily expressed in the liver and adipose which partitions between membranes and lipid droplets. PNPLA3 hydrolyzes acylglycerols with maximal activity against triacylglycerol, diacylglycerol, and monoacylglycerol. A polymorphism (**rs738409**, p.I148M) in *PNPLA3* has been associated with Alcoholic and Non-alcoholic Fatty Liver Disease (NAFLD), Non-alcoholic Steatohepatitis (NASH), Liver Cirrhosis, Hepatic Cell Carcinoma, and Type 2 diabetes, although the mechanistic basis for these associations is not entirely resolved72-76. *Pnlpa3* knockout mice failed to demonstrate significant increases in liver fat. Overexpressing human PNPLA3 in mouse liver did not increase steatosis, but overexpression of the human p.I148M variant did recapitulate the fatty liver phenotype in mice, and metabolic studies suggested a complex mechanism⁷⁵. Knock-in of the human p.I148M variant at endogenous levels showed that accumulation of triacylglycerides in the liver is associated with an enrichment of catalytically inactive PNPLA3 on lipid droplets⁷⁶.
- *SLC30A8* Solute carrier family 30, member 8 (SLC30A8, or Znt8) is a zinc transporter highly expressed in the pancreas and implicated in insulin secretion in humans. Early GWAS studies by Sladek et al. ⁷⁷ associated the missense allele Arg325Trp (**rs13266634**) with susceptibility to T2D. The T2D risk allele was shown to increase fasting proinsulin, fasting insulin and fasting glucose, and was found associated with lower insulinogenic index and HOMA derived β-cell function, suggesting a defect in β-cell insulin processing, secretion or insulin clearance. Humans homozygous for the diabetes risk variant exhibited a post–oral glucose increase in the C-peptide:insulin ratio. Functional studies of the p.Arg325Trp risk variant have reported conflicting effects on zinc transporter activity. However, in 2014, large scale sequencing of *SLC30A8* in multiple populations demonstrated that heterozygosity for truncating LOF mutations in *SLC30A8* results in substantial protection from the risk of T2D78.

D. Neurodegeneration

LRRK2 Mutations in *LRRK2* are the most common genetic cause of both familial and sporadic Parkinson's disease (PD). The most common PD mutation, p.G2019S (rs34637584), located within the LRRK2 kinase domain, causes autosomal dominant PD. G2019S increases kinase activity, auto-phosphorylation and

phosphorylation of generic substrates, and is enriched in Ashkenazi Jews and North African Arab Berbers⁷⁹. Non-coding variants at the *LRRK2* locus (e.g., **rs1491942**) are associated with increased *LRRK2* expression in multiple cell types and increased risk of sporadic PD through GWAS⁸⁰. Together, the genetic and functional data suggest that inhibition of LRRK2's kinase function is a potential therapeutic mechanism for the treatment of PD patients with the G2019S mutation, and perhaps more broadly patients with PD due to other etiologies. Both, p.G2019S and another coding variant, **rs11564187** (p.N2081D), are also unequivocally associated with risk of CD and $UC^{3,9}$.

TMEM175 GWAS and conditional analyses identified **rs34311866** (p.M393T) and **rs34884217** (p.Q65P) within *TMEM175* at the *GAK/TMEM175/DGKQ* locus as associated with the risk of PD^{80} . TMEM175 is a lysosomal potassium channel that regulates lysosomal $K(+)$ permeability⁸¹. Deficiency for TMEM175 in neuronal cells impairs lysosomal function and increases α -synuclein aggregation⁸². Modification of TMEM175's $K(+)$ channel activity is considered a potential therapeutic mechanism for the treatment of PD patients.

Supplementary Notes

Supplementary Note 1. List of published studies in the Genomics plc GWAS database used in this study

Berndt, SI. et al. Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. *Nat Genet.* **45**, 501-12. doi: 10.1038/ng.2606. (2013). Bradfield, JP. et al. A genome-wide association meta-analysis identifies new childhood obesity loci. *Nat Genet.* **44**, 526-31. doi: 10.1038/ng.2247. (2012). (www.egg-consortium.org)

Buch, S. et al. A genome-wide association study confirms PNPLA3 and identifies TM6SF2 and MBOAT7 as risk loci for alcohol-related cirrhosis. *Nat Genet.* **47**, 1443-8. doi: 10.1038/ng.3417. (2015).

CONVERGE consortium et al. Sparse whole-genome sequencing identifies two loci for major depressive disorder. *Nature.* **523**, 588-91. doi: 10.1038/nature14659. (2015).

Cordell, HJ. et al. International genome-wide meta-analysis identifies new primary biliary cirrhosis risk loci and targetable pathogenic pathways. *Nat Commun.* **6**, 8019. doi: 10.1038/ncomms9019. (2015).

Coronary Artery Disease (C4D) Genetics Consortium et al. A genome-wide association study in Europeans and South Asians identifies five new loci for coronary artery disease. *Nat Genet.* **43**, 339- 44. doi: 10.1038/ng.782. (2011).

Evans, DM. et al. Interaction between ERAP1 and HLA-B27 in ankylosing spondylitis implicates peptide handling in the mechanism for HLA-B27 in disease susceptibility. *Nat Genet.* **43**, 761-7. doi: 10.1038/ng.873. (2011).

Fogh, I. et al. A genome-wide association meta-analysis identifies a novel locus at 17q112 associated with sporadic amyotrophic lateral sclerosis. *Hum Mol Genet.* **23**, 2220-31. doi: 10.1093/hmg/ddt587. (2014).

Gaulton, KJ. et al. Genetic fine mapping and genomic annotation defines causal mechanisms at type 2 diabetes susceptibility loci. *Nat Genet.* **47**, 1415-25. doi: 10.1038/ng.3437. (2015).

Genetic Analysis of Psoriasis Consortium & the Wellcome Trust Case Control Consortium 2 et al. A genome-wide association study identifies new psoriasis susceptibility loci and an interaction between HLA-C and ERAP1. *Nat Genet.* **42**, 985-90. doi: 10.1038/ng.694. (2010).

International Consortium for Systemic Lupus Erythematosus Genetics (SLEGEN) et al. Genome-wide association scan in women with systemic lupus erythematosus identifies susceptibility variants in ITGAM, PXK, KIAA1542 and other loci. *Nat Genet.* **40**, 204-10. doi: 10.1038/ng.81. (2008).

International Multiple Sclerosis Genetics Consortium et al. Genetic risk and a primary role for cellmediated immune mechanisms in multiple sclerosis. *Nature.* **476**, 214-9. doi: 10.1038/nature10251. (2011).

International Stroke Genetics Consortium (ISGC) et al. Genome-wide association study identifies a variant in HDAC9 associated with large vessel ischemic stroke. *Nat Genet.* **44**, 328-33. doi: 10.1038/ng.1081. (2012).

Lambert, JC. et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet.* **45**, 1452-8. doi: 10.1038/ng.2802. (2013).

Liu, JZ. et al. Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations. *Nat Genet.* **47**, 979-86. doi: 10.1038/ng.3359. (2015). Mells, GF. et al. Genome-wide association study identifies 12 new susceptibility loci for primary biliary cirrhosis. *Nat Genet.* **43**, 329-32. doi: 10.1038/ng.789. (2011).

Moffatt, MF. et al. A large-scale, consortium-based genomewide association study of asthma. N Engl J Med. 363, 1211-21. doi: 10.1056/NEJMoa0906312. (2010).

Morris, AP. et al. Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet.* **44**, 981-90. doi: 10.1038/ng.2383. (2012).

Nair, RP. et al. Genome-wide scan reveals association of psoriasis with IL-23 and NF-kappaB pathways. *Nat Genet.* **41**, 200-204. doi: 10.1038/ng.311. (2009). (dbGaP accession number phs000019) Okada, Y. et al. Genetics of rheumatoid arthritis contributes to biology and drug discovery. *Nature.* **506**, 376-81. doi: 10.1038/nature12873. (2014).

Otowa, T. et al. Meta-analysis of genome-wide association studies of anxiety disorders. *Mol Psychiatry.* **21**, 1391-9. doi: 10.1038/mp.2015.197. (2016).

Patsopoulos, NA. et al. Genome-wide meta-analysis identifies novel multiple sclerosis susceptibility loci. *Ann Neurol.* **70**, 897-912. doi: 10.1002/ana.22609. (2011).

Pattaro, C. et al. Genetic associations at 53 loci highlight cell types and biological pathways relevant for kidney function. *Nat Commun.* **7**, 10023. doi: 10.1038/ncomms10023. (2016).

Schunkert, H. et al. Large-scale association analysis identifies 13 new susceptibility loci for coronary artery disease. *Nat Genet.* **43**, 333-8. doi: 10.1038/ng.784. (2011).

(www.CARDIOGRAMPLUSC4D.ORG)

Tobacco and Genetics Consortium et al. Genome-wide meta-analyses identify multiple loci associated with smoking behavior. *Nat Genet.* **42**, 441-7. doi: 10.1038/ng.571. (2010).

UK IBD Genetics Consortium et al. Genome-wide association study of ulcerative colitis identifies three new susceptibility loci, including the HNF4A region. *Nat Genet.* **41**, 1330-4. doi: 10.1038/ng.483. (2009).

UK Parkinson's Disease Consortium et al. Dissection of the genetics of Parkinson's disease identifies an additional association 5' of SNCA and multiple associated haplotypes at 17q21. *Hum Mol Genet.* **20**, 345-53. doi: 10.1093/hmg/ddq469. (2011).

Wellcome Trust Case Control Consortium et al. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature.* **447**, 661-78. doi: 10.1038/nature05911. (2007).

Supplementary Figures

B

Supplementary Fig. 1. Replication of known GWAS associations through meta-analysis of PheWAS results from four disease-agnostic cohorts with extensive health information. (A) Distribution of replication rate based on statistical power to detect an association. Power to replicate at *P*<0.05 is shown. Power is calculated based on the SNP frequency, the direction and point estimate of effect size of the SNP in published studies, the number of cases and controls in the meta-analysis, and a disease prevalence of 1%. **(B)** Number of known GWAS associations that replicated or failed to replicate in the PheWAS meta-analysis, segregated by disease. The published Parkinson's disease (PD) GWAS included 23andMe results.

Supplementary Fig. 2. Effect sizes **of known GWAS associations in the meta-PheWAS**. Odds ratios (ORs) and 95% confidence intervals in the published GWAS are shown on the x axis. ORs and 95% confidence intervals in the meta-PheWAS are shown on the y axis. Powered associations (beta<0.8, alpha = 0.05) reaching *P*<1.8x10- ⁶ (Bonferroni corrected pvalue), FDR<0.1 or *P*<0.05 in meta-PheWAS are shown in dark red, red and light red respectively. Powered associations not reaching *P*<0.05 in meta-PheWAS are shown in white. Associations not powered in the meta-PheWAS are shown in grey. The regression coefficient R^2 between published ORs and ORs in meta-PheWAS is indicated.

Supplementary Fig. 3. Effect sizes of known GWAS associations in the disease-agnostic cohorts PheWAS. Odds ratios (ORs) and 95% confidence intervals in the published GWAS are shown on the x axis. ORs and 95% confidence intervals in 23andMe (A), Gplc/UK Biobank (B), FINRISK (C) and CHOP (D) are shown on the y axis. Associations reaching $P \le 1.8x10^{-6}$ (Bonferroni corrected pvalue), FDR ≤ 0.1 or $P \le 0.05$ in the PheWAS are shown in dark red, red and light red respectively. Associations not reaching *P*<0.05 are shown in white. The regression coefficients R^2 between published ORs and ORs in meta-PheWAS are indicated.

Supplementary Fig. 4. Effects of replicated and putative novel associations (FDR<0.1). Sixteen SNPs showed novel associations at FDR<0.1 (*P*<7e-4) in the meta-analysis of PheWAS and GWAS results. Effects (Odds ratio, OR) of the replicated known (black) and novel (grey) associations reaching FDR<0.1 are shown. The number (N) of disease-agnostic and GWAS cohorts contributing to a respective phenotype is indicated. *, Result derived from GWAS cohort only since the corresponding phenotype is not captured or not powered in the disease-agnostic cohorts tested.

A *I* **2<40%**

OR

B ²>40%

Supplementary Fig. 5. Effect sizes of putative novel associations observed in the individual cohorts and in the meta-analysis. 31 putative novel associations were obtained by meta-analyzing associations results from the disease-agnostic cohorts with GWAS results. Odds ratios (OR), 95% confidence intervals and *P* values are shown. **A.** Heterogeneity I^2 statistic <40%. **B.** $I^2 > 40\%$.

Supplementary Fig. 6. Independent associations for galectin-3 plasma levels and Parkinson's disease risk at the *GCH1-LGALS3* **locus.** Regional association results at the *GCH1-LGALS3* demonstrate that the associations with circulating plasma galectin-3 levels (top panel) and Parkinson's disease (PD, bottom panel) risk are driven by different causal SNPs. Co-localization test using *coloc* returns a posterior probability (PP) for a shared variant of 0.0008%.

Supplementary Fig. 7. Phenotype-phenotype correlations in 23andMe. Pairwise phenotype correlation was calculated in participants for whom data on both phenotypes was available through *r*=cov/sqrt(var1*var2), with cov=covariance of phenotype 1 and phenotype 2, var1=variance in phenotype 1, var2=variance of phenotype 2. Only phenotypes associated at FDR<0.1 with indicated SNPs are shown. Phenotypes related to known associations are highlighted in bold.

Supplementary Fig. 8. Association of *PNPLA3* **rs738409-G with multiple disease-relevant endpoints.** Associations reaching FDR<0.1 in joint meta-analysis of PheWAS results with available GWAS results are shown. Odds ratios, 95% confidence intervals, *P* values, and numbers (*N*) of cases and controls are represented. Known GWAS associations are indicated with an asterix (*).

Supplementary Tables

Supplementary Table 1. SNP-phenotype associations reported earlier in published GWAS for the 25 SNPs interrogated through PheWAS in this study

 $$ Significant associations (P < 5e-8) in the European population$

^ Effect of minor allele

* Quantitative traits

** Quantitative traits related to binary traits tested in PheWAS: blood pressure and high blood pressure, ALT and elevated liver test

IBD, inflammatory bowel disease; T1D, type 1 diabetes; aPTT, activated partial thromboplastin time; VTE, venous thromboembolism; BMI, body mass index; CAD, Coronary Artery Disease; MI, myocardial Infarction; IgAD, immunoglobulin A Deficiency; SLE, Systemic Lupus Erythematosus; RA, rheumatoid arthritis; NAFLD, Non-alcoholic fatty liver disease; ALT, alanine aminotransferase

Full list of individual phenotypic endpoints that are mapped for meta-analysis is provided in Supplementary Data file 1.

Supplementary Table 3. Mapped phenotypes found in at least two disease-agnostic cohorts and included in the meta-analysis

7. Diseases of the eye

Full list of individual phenotypic endpoints that are mapped for meta-analysis is provided in Supplementary Data file 1. NOS, not otherwise specified

Supplementary Table 4. Replication of known GWAS associations in the PheWAS meta-analysis

^ Effect of minor allele. Refer to Supplementary Data file 1 for detailed information.

\$ Direction of effect in 23andMe, Genomics plc UK Biobank, FINRISK, and CHOP, respectively.

Statistical power to detect an association in the meta-PheWAS was calculated based on the effect allele frequency in the 1000Genomes European population, the published effect size in GWAS, the number of cases and controls in the meta-PheWAS and the corresponding disease prevalence reported by the Centers for Disease Control and Prevention (see Methods). Three significance cutoffs were tested: *P*<0.05, FDR<0.1 (*P*<3.8e-4), and Bonferroni-corrected significance $(P<1.8e-6)$.

CAD, coronary artery disease; IBD, inflammatory bowel disease; SLE, systemic lupus erythematosus; PBC, primary biliary cirrhosis.

Supplementary Table 5. Putative novel associations identified in the meta-PheWAS

*Putative novel associations reaching FDR<0.1 are shown. Associations reaching Bonferroni-corrected significance (*P*<1.8e-6) are highlighted in bold. Full meta-PheWAS results are provided in Supplementary file 1.

^ Effect of minor allele. Refer to Supplementary Data file 1 for detailed information.

\$ Direction of effect in 23andMe, Genomics plc UK Biobank, FINRISK, CHOP and GWAS, respectively.

Numbers in parentheses after the Phenotype name identify related/non-independent phenotypic endpoints

Supplementary Table 6. Replication of potential novel associations in the latest release of UK Biobank

*Associations reaching Bonferroni-corrected significance (*P*<1.8e-6) in the discovery PheWAS meta-analysis are highlighted in bold.

#Directions of effect in 23andMe, Gplc UK Biobank, FINRISK, CHOP and GWAS

^ Statistical power to detect an association at *P*<0.05 in UK Biobank v2 based on the SNP MAF, the number of cases and controls in UK Biobank v2 and the OR observed in the PheWAS meta-analysis

\$Weighted *Z* score meta-analysis using the association results from the 23andMe, FINRISK and CHOP PheWAS cohorts (when available), GWAS (when available) and UK Biobank v2. Weights are calculated based on the number of cases and controls. Details are provided in Methods. The Gplc UK Biobank association results are excluded in this meta-analysis.

Supplementary Table 7. Novel association of the *PNPLA3* **SNP rs738409 in the 23andMe cohort, before and after adjusting for elevated liver test**

*In the elevated liver test-adjusted analysis, self-report of elevated liver test was included as covariate in the logistic regression, and only participants who answered both liver test questions and questions related to the primary phenotype tested were included.

^ Odds ratio of the minor allele, which is associated with elevated alanine aminotransferase (ALT) levels

Supplementary Table 8. Association of the *IFIH1* **SNP rs2111485 (in complete LD with the** *IFIH1* **missense SNP rs1990760) with asthma in 23andMe cohort, before and after adjusting for autoimmune diseases**

*In the autoimmune disease-adjusted analysis, self-report of autoimmune disorder was included as covariate in the logistic regression, and only participants who answered both autoimmune and asthma-related questions were included. ^ Odds ratio of the minor allele, which is known to protect against multiple autoimmune diseases.

Supplementary References

- 1. Hampe, J. et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. *Nat Genet* **39**, 207-211 (2007).
- 2. Rioux, J.D. et al. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. *Nat Genet.* **39**, 596-604. Epub 2007 Apr 2015. (2007).
- 3. Jostins, L. et al. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* **491**, 119-124 (2012).
- 4. Lassen, K.G. et al. Atg16L1 T300A variant decreases selective autophagy resulting in altered cytokine signaling and decreased antibacterial defense. *Proc Natl Acad Sci U S A.* **111**, 7741- 7746. doi: 7710.1073/pnas.1407001111. Epub 1407002014 May 1407001112. (2014).
- 5. Murthy, A. et al. A Crohn's disease variant in Atg16l1 enhances its degradation by caspase 3. *Nature.* **506**, 456-462. doi: 410.1038/nature13044. Epub 12014 Feb 13019. (2014).
- 6. Franke, A. et al. Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat Genet.* **42**, 1118-1125. doi: 1110.1038/ng.1717. (2010).
- 7. Fairfax, B.P. et al. Innate immune activity conditions the effect of regulatory variants upon monocyte gene expression. *Science.* **343**, 1246949. doi: 1246910.1241126/science.1246949. (2014).
- 8. Janse, M. et al. Three ulcerative colitis susceptibility loci are associated with primary sclerosing cholangitis and indicate a role for IL2, REL, and CARD9. *Hepatology.* **53**, 1977-1985. doi: 1910.1002/hep.24307. Epub 22011 May 24302. (2011).
- 9. Rivas, M.A. et al. Deep resequencing of GWAS loci identifies independent rare variants associated with inflammatory bowel disease. *Nat Genet.* **43**, 1066-1073. doi: 1010.1038/ng.1952. (2011).
- 10. Lanternier, F. et al. Deep dermatophytosis and inherited CARD9 deficiency. *N Engl J Med.* **369**, 1704-1714. doi: 1710.1056/NEJMoa1208487. Epub 1202013 Oct 1208416. (2013).
- 11. Ruland, J. CARD9 signaling in the innate immune response. *Ann N Y Acad Sci.* **1143:35-44.**, 10.1196/annals.1443.1024. (2008).
- 12. Elishmereni, M., Bachelet, I. & Levi-Schaffer, F. DNAM-1: an amplifier of immune responses as a therapeutic target in various disorders. *Curr Opin Investig Drugs.* **9**, 491-496. (2008).
- 13. Xiong, P., Sang, H.W. & Zhu, M. Critical roles of co-activation receptor DNAX accessory molecule-1 in natural killer cell immunity. *Immunology.* **146**, 369-378. doi: 310.1111/imm.12516. Epub 12015 Sep 12528. (2015).
- 14. Onengut-Gumuscu, S. et al. Fine mapping of type 1 diabetes susceptibility loci and evidence for colocalization of causal variants with lymphoid gene enhancers. *Nat Genet.* **47**, 381-386. doi: 310.1038/ng.3245. Epub 2015 Mar 1039. (2015).
- 15. Gieger, C. et al. New gene functions in megakaryopoiesis and platelet formation. *Nature.* **480**, 201-208. doi: 210.1038/nature10659. (2011).
- 16. Gross, C.C. et al. Impaired NK-mediated regulation of T-cell activity in multiple sclerosis is reconstituted by IL-2 receptor modulation. *Proc Natl Acad Sci U S A.* **113**, E2973-2982. doi: 2910.1073/pnas.1524924113. Epub 1524922016 May 1524924119. (2016).
- 17. Lofgren, S.E. et al. A 3'-untranslated region variant is associated with impaired expression of CD226 in T and natural killer T cells and is associated with susceptibility to systemic lupus erythematosus. *Arthritis Rheum.* **62**, 3404-3414. doi: 3410.1002/art.27677. (2010).
- 18. Divorty, N., Mackenzie, A.E., Nicklin, S.A. & Milligan, G. G protein-coupled receptor 35: an emerging target in inflammatory and cardiovascular disease. *Front Pharmacol.* **6:41.**, 10.3389/fphar.2015.00041. eCollection 02015. (2015).
- 19. Lassen, K.G. et al. Genetic Coding Variant in GPR65 Alters Lysosomal pH and Links Lysosomal Dysfunction with Colitis Risk. *Immunity.* **44**, 1392-1405. doi: 1310.1016/j.immuni.2016.1305.1007. Epub 2016 Jun 1397. (2016).
- 20. Looney, B.M., Xia, C.Q., Concannon, P., Ostrov, D.A. & Clare-Salzler, M.J. Effects of type 1 diabetes-associated IFIH1 polymorphisms on MDA5 function and expression. *Curr Diab Rep.* **15**, 96. doi: 10.1007/s11892-11015-10656-11898. (2015).
- 21. Oda, H. et al. Aicardi-Goutieres syndrome is caused by IFIH1 mutations. *Am J Hum Genet.* **95**, 121-125. doi: 110.1016/j.ajhg.2014.1006.1007. (2014).
- 22. Rice, G.I. et al. Gain-of-function mutations in IFIH1 cause a spectrum of human disease phenotypes associated with upregulated type I interferon signaling. *Nat Genet.* **46**, 503-509. doi: 510.1038/ng.2933. Epub 2014 Mar 1030. (2014).
- 23. Rutsch, F. et al. A specific IFIH1 gain-of-function mutation causes Singleton-Merten syndrome. *Am J Hum Genet.* **96**, 275-282. doi: 210.1016/j.ajhg.2014.1012.1014. Epub 2015 Jan 1022. (2015).
- 24. Asgari, S. et al. Severe viral respiratory infections in children with IFIH1 loss-of-function mutations. *Proc Natl Acad Sci U S A* **17**, 1704259114 (2017).
- 25. Lamborn, I.T. et al. Recurrent rhinovirus infections in a child with inherited MDA5 deficiency. *J Exp Med.* **214**, 1949-1972. doi: 1910.1084/jem.20161759. Epub 20162017 Jun 20161712. (2017).
- 26. Budu-Aggrey, A. et al. A rare coding allele in IFIH1 is protective for psoriatic arthritis. *Ann Rheum Dis.* **76**, 1321-1324. doi: 1310.1136/annrheumdis-2016-210592. Epub 212017 May 210513. (2017).
- 27. Downes, K. et al. Reduced expression of IFIH1 is protective for type 1 diabetes. *PLoS One.* **5(9).** e12646. doi: 12610.11371/journal.pone.0012646. (2010).
- 28. Jin, Y. et al. Genome-wide association analyses identify 13 new susceptibility loci for generalized vitiligo. *Nat Genet.* **44**, 676-680. doi: 610.1038/ng.2272. (2012).
- 29. Jin, Y., Andersen, G.H., Santorico, S.A. & Spritz, R.A. Multiple Functional Variants of IFIH1, a Gene Involved in Triggering Innate Immune Responses, Protect against Vitiligo. *J Invest Dermatol.* **137**, 522-524. doi: 510.1016/j.jid.2016.1009.1021. Epub 2016 Oct 1015. (2017).
- 30. Li, Y. et al. Carriers of rare missense variants in IFIH1 are protected from psoriasis. *J Invest Dermatol.* **130**, 2768-2772. doi: 2710.1038/jid.2010.2214. Epub 2010 Jul 2729. (2010).
- 31. Nejentsev, S., Walker, N., Riches, D., Egholm, M. & Todd, J.A. Rare variants of IFIH1, a gene implicated in antiviral responses, protect against type 1 diabetes. *Science.* **324**, 387-389. doi: 310.1126/science.1167728. Epub 1162009 Mar 1167725. (2009).
- 32. Shigemoto, T. et al. Identification of loss of function mutations in human genes encoding RIG-I and MDA5: implications for resistance to type I diabetes. *J Biol Chem.* **284**, 13348-13354. doi: 13310.11074/jbc.M809449200. Epub 809442009 Mar 809449226. (2009).
- 33. Smyth, D.J. et al. A genome-wide association study of nonsynonymous SNPs identifies a type 1 diabetes locus in the interferon-induced helicase (IFIH1) region. *Nat Genet.* **38**, 617-619. Epub 2006 May 2014. (2006).
- 34. Zurawek, M. et al. Cumulative effect of IFIH1 variants and increased gene expression associated with type 1 diabetes. *Diabetes Res Clin Pract.* **107**, 259-266. doi: 210.1016/j.diabres.2014.1011.1008. Epub 2014 Dec 1014. (2015).
- 35. Bentham, J. et al. Genetic association analyses implicate aberrant regulation of innate and adaptive immunity genes in the pathogenesis of systemic lupus erythematosus. *Nat Genet* **47**, 1457-1464 (2015).
- 36. Liu, J.Z. et al. Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations. *Nat Genet.* **47**, 979-986. doi: 910.1038/ng.3359. Epub 2015 Jul 1020. (2015).
- 37. Gorman, J.A. et al. The A946T variant of the RNA sensor IFIH1 mediates an interferon program that limits viral infection but increases the risk for autoimmunity. *Nat Immunol.* **18**, 744-752. doi: 710.1038/ni.3766. Epub 2017 May 1029. (2017).
- 38. Okada, Y. et al. Genetics of rheumatoid arthritis contributes to biology and drug discovery. *Nature* **506**, 376-381 (2014).
- 39. Taylor, K.E. et al. Genome-Wide Association Analysis Reveals Genetic Heterogeneity of Sjogren's Syndrome According to Ancestry. *Arthritis Rheumatol.* **69**, 1294-1305. doi: 1210.1002/art.40040. Epub 42017 May 40049. (2017).
- 40. Liu, X. et al. Genome-wide meta-analyses identify three loci associated with primary biliary cirrhosis. *Nat Genet.* **42**, 658-660. doi: 610.1038/ng.1627. Epub 2010 Jul 1018. (2010).
- 41. Radstake, T.R. et al. Genome-wide association study of systemic sclerosis identifies CD247 as a new susceptibility locus. *Nat Genet.* **42**, 426-429. doi: 410.1038/ng.1565. Epub 2010 Apr 1011. (2010).
- 42. Hedl, M. & Abraham, C. IRF5 risk polymorphisms contribute to interindividual variance in pattern recognition receptor-mediated cytokine secretion in human monocyte-derived cells. *J Immunol.* **188**, 5348-5356. doi: 5310.4049/jimmunol.1103319. Epub 1102012 Apr 1103327. (2012).
- 43. Hedl, M., Yan, J. & Abraham, C. IRF5 and IRF5 Disease-Risk Variants Increase Glycolysis and Human M1 Macrophage Polarization by Regulating Proximal Signaling and Akt2 Activation. *Cell Rep.* **16**, 2442-2455. doi: 2410.1016/j.celrep.2016.2407.2060. Epub 2016 Aug 2418. (2016).
- 44. Niewold, T.B. et al. IRF5 haplotypes demonstrate diverse serological associations which predict serum interferon alpha activity and explain the majority of the genetic association with systemic lupus erythematosus. *Ann Rheum Dis.* **71**, 463-468. doi: 410.1136/annrheumdis-2011- 200463. Epub 202011 Nov 200416. (2012).
- 45. Kreins, A.Y. et al. Human TYK2 deficiency: Mycobacterial and viral infections without hyper-IgE syndrome. *J Exp Med.* **212**, 1641-1662. doi: 1610.1084/jem.20140280. Epub 20142015 Aug 20140224. (2015).
- 46. Minegishi, Y. et al. Human tyrosine kinase 2 deficiency reveals its requisite roles in multiple cytokine signals involved in innate and acquired immunity. *Immunity.* **25**, 745-755. (2006).
- 47. Tsoi, L.C. et al. Identification of 15 new psoriasis susceptibility loci highlights the role of innate immunity. *Nat Genet.* **44**, 1341-1348. doi: 1310.1038/ng.2467. Epub 2012 Nov 1311. (2012).
- 48. Beecham, A.H. et al. Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis. *Nat Genet.* **45**, 1353-1360. doi: 1310.1038/ng.2770. Epub 2013 Sep 1329. (2013).
- 49. Diogo, D. et al. TYK2 protein-coding variants protect against rheumatoid arthritis and autoimmunity, with no evidence of major pleiotropic effects on non-autoimmune complex traits. *PLoS One.* **10**, e0122271. doi: 0122210.0121371/journal.pone.0122271. eCollection 0122015. (2015).
- 50. Couturier, N. et al. Tyrosine kinase 2 variant influences T lymphocyte polarization and multiple sclerosis susceptibility. *Brain.* **134**, 693-703. doi: 610.1093/brain/awr1010. (2011).
- 51. Dendrou, C.A. et al. Resolving TYK2 locus genotype-to-phenotype differences in autoimmunity. *Sci Transl Med.* **8**, 363ra149. (2016).
- 52. Bonnelykke, K. et al. A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. *Nat Genet.* **46**, 51-55. doi: 10.1038/ng.2830. Epub 2013 Nov 1017. (2014).
- 53. Bochkov, Y.A. et al. Cadherin-related family member 3, a childhood asthma susceptibility gene product, mediates rhinovirus C binding and replication. *Proc Natl Acad Sci U S A.* **112**, 5485- 5490. doi: 5410.1073/pnas.1421178112. Epub 1421172015 Apr 1421178116. (2015).
- 54. Asakai, R., Chung, D.W., Ratnoff, O.D. & Davie, E.W. Factor XI (plasma thromboplastin antecedent) deficiency in Ashkenazi Jews is a bleeding disorder that can result from three types of point mutations. *Proc Natl Acad Sci U S A.* **86**, 7667-7671. (1989).
- 55. Sabater-Lleal, M. et al. A genome-wide association study identifies KNG1 as a genetic determinant of plasma factor XI Level and activated partial thromboplastin time. *Arterioscler Thromb Vasc Biol.* **32**, 2008-2016. doi: 2010.1161/ATVBAHA.2112.248492. Epub 242012 Jun 248414. (2012).
- 56. Tang, W. et al. Genetic associations for activated partial thromboplastin time and prothrombin time, their gene expression profiles, and risk of coronary artery disease. *Am J Hum Genet.* **91**, 152-162. doi: 110.1016/j.ajhg.2012.1005.1009. Epub 2012 Jun 1014. (2012).
- 57. Tang, W. et al. A genome-wide association study for venous thromboembolism: the extended cohorts for heart and aging research in genomic epidemiology (CHARGE) consortium. *Genet Epidemiol.* **37**, 512-521. doi: 510.1002/gepi.21731. Epub 22013 May 21735. (2013).
- 58. Buller, H.R. et al. Factor XI antisense oligonucleotide for prevention of venous thrombosis. *N Engl J Med.* **372**, 232-240. doi: 210.1056/NEJMoa1405760. Epub 1402014 Dec 1405767. (2015).
- 59. Egeberg, O. Factor XII defect and hemorrhage. Evidence for a new type of hereditary hemostatic disorder. *Thromb Diath Haemorrh.* **23**, 432-440. (1970).
- 60. Ratnoff, O.D. & Steinberg, A.G. Further studies on the inheritance of Hageman trait. *J Lab Clin Med.* **59**, 980-985. (1962).
- 61. Dewald, G. & Bork, K. Missense mutations in the coagulation factor XII (Hageman factor) gene in hereditary angioedema with normal C1 inhibitor. *Biochem Biophys Res Commun.* **343**, 1286-1289. doi: 1210.1016/j.bbrc.2006.1203.1092 (2006).
- 62. Renne, T. et al. Defective thrombus formation in mice lacking coagulation factor XII. *J Exp Med.* **202**, 271-281. Epub 2005 Jul 2011. (2005).
- 63. Calafell, F. et al. Sequence variation and genetic evolution at the human F12 locus: mapping quantitative trait nucleotides that influence FXII plasma levels. *Hum Mol Genet.* **19**, 517-525. doi: 510.1093/hmg/ddp1517. Epub 2009 Nov 1023. (2010).
- 64. Houlihan, L.M. et al. Common variants of large effect in F12, KNG1, and HRG are associated with activated partial thromboplastin time. *Am J Hum Genet.* **86**, 626-631. doi: 610.1016/j.ajhg.2010.1002.1016. Epub 2010 Mar 1018. (2010).
- 65. Locke, A.E. et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature* **518**, 197-206 (2015).
- 66. Dobrin, R. et al. Dissecting cis regulation of gene expression in human metabolic tissues. *PLoS One* **6**, e23480. doi: 23410.21371/journal.pone.0023480. Epub 0022011 Aug 0023431. (2011).
- 67. Hong, J.H. et al. GDF15 Is a Novel Biomarker for Impaired Fasting Glucose. *Diabetes Metab J.* **38**, 472-479. doi: 410.4093/dmj.2014.4038.4096.4472. Epub 2014 Dec 4015. (2014).
- 68. Ehret, G.B. et al. Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. *Nature.* **478**, 103-109. doi: 110.1038/nature10405. (2011).
- 69. Rippe, C. et al. Hypertension reduces soluble guanylyl cyclase expression in the mouse aorta via the Notch signaling pathway. *Sci Rep.* **7**, 1334. doi: 1310.1038/s41598-41017-01392- 41591. (2017).
- 70. Herve, D. et al. Loss of alpha1beta1 soluble guanylate cyclase, the major nitric oxide receptor, leads to moyamoya and achalasia. *Am J Hum Genet.* **94**, 385-394. doi: 310.1016/j.ajhg.2014.1001.1018. Epub 2014 Feb 1027. (2014).
- 71. de Boer, R.A. et al. A genome-wide association study of circulating galectin-3. *PLoS One* **7**, e47385. doi: 47310.41371/journal.pone.0047385. Epub 0042012 Oct 0047389. (2012).
- 72. Chambers, J.C. et al. Genome-wide association study identifies loci influencing concentrations of liver enzymes in plasma. *Nat Genet.* **43**, 1131-1138. doi: 1110.1038/ng.1970. (2011).
- 73. Speliotes, E.K., Butler, J.L., Palmer, C.D., Voight, B.F. & Hirschhorn, J.N. PNPLA3 variants specifically confer increased risk for histologic nonalcoholic fatty liver disease but not metabolic disease. *Hepatology.* **52**, 904-912. doi: 910.1002/hep.23768. (2010).
- 74. Tian, C., Stokowski, R.P., Kershenobich, D., Ballinger, D.G. & Hinds, D.A. Variant in PNPLA3 is associated with alcoholic liver disease. *Nat Genet.* **42**, 21-23. doi: 10.1038/ng.1488. Epub 2009 Nov 1029. (2010).
- 75. Li, J.Z. et al. Chronic overexpression of PNPLA3I148M in mouse liver causes hepatic steatosis. *J Clin Invest.* **122**, 4130-4144. (2012).
- 76. Smagris, E. et al. Pnpla3I148M knockin mice accumulate PNPLA3 on lipid droplets and develop hepatic steatosis. *Hepatology.* **61**, 108-118. doi: 110.1002/hep.27242. Epub 22014 Oct 27241. (2015).
- 77. Sladek, R. et al. A genome-wide association study identifies novel risk loci for type 2 diabetes. *Nature.* **445**, 881-885. Epub 2007 Feb 2011. (2007).
- 78. Flannick, J. et al. Loss-of-function mutations in SLC30A8 protect against type 2 diabetes. *Nat Genet.* **46**, 357-363. doi: 310.1038/ng.2915. Epub 2014 Mar 1032. (2014).
- 79. Paisan-Ruiz, C., Lewis, P.A. & Singleton, A.B. LRRK2: cause, risk, and mechanism. *J Parkinsons Dis* **3**, 85-103. doi: 110.3233/JPD-130192. (2013).
- 80. Nalls, M.A. et al. Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. *Nat Genet* **46**, 989-993 (2014).
- 81. Cang, C., Aranda, K., Seo, Y.J., Gasnier, B. & Ren, D. TMEM175 Is an Organelle K(+) Channel Regulating Lysosomal Function. *Cell.* **162**, 1101-1112. doi: 1110.1016/j.cell.2015.1108.1002. (2015).
- 82. Jinn, S. et al. TMEM175 deficiency impairs lysosomal and mitochondrial function and increases alpha-synuclein aggregation. *Proc Natl Acad Sci U S A.* **114**, 2389-2394. doi: 2310.1073/pnas.1616332114. Epub 1616332017 Feb 1616332113. (2017).
- 83. Nikpay, M. et al. A comprehensive 1,000 Genomes-based genome-wide association metaanalysis of coronary artery disease. *Nat Genet.* **47**, 1121-1130. doi: 1110.1038/ng.3396. Epub 2015 Sep 1127. (2015).
- 84. Ferreira, R.C. et al. Association of IFIH1 and other autoimmunity risk alleles with selective IgA deficiency. *Nat Genet.* **42**, 777-780. doi: 710.1038/ng.1644. Epub 2010 Aug 1038. (2010).
- 85. Jin, Y. et al. Genome-wide association studies of autoimmune vitiligo identify 23 new risk loci and highlight key pathways and regulatory variants. *Nat Genet.* **48**, 1418-1424. doi: 1410.1038/ng.3680. Epub 2016 Oct 1410. (2016).
- 86. Buch, S. et al. A genome-wide association study confirms PNPLA3 and identifies TM6SF2 and MBOAT7 as risk loci for alcohol-related cirrhosis. *Nat Genet.* **47**, 1443-1448. doi: 1410.1038/ng.3417. Epub 2015 Oct 1419. (2015).
- 87. Liu, C.T. et al. Trans-ethnic Meta-analysis and Functional Annotation Illuminates the Genetic Architecture of Fasting Glucose and Insulin. *Am J Hum Genet.* **99**, 56-75. doi: 10.1016/j.ajhg.2016.1005.1006. Epub 2016 Jun 1016. (2016).
- 88. Mahajan, A. et al. Genome-wide trans-ancestry meta-analysis provides insight into the genetic architecture of type 2 diabetes susceptibility. *Nat Genet.* **46**, 234-244. doi: 210.1038/ng.2897. Epub 2014 Feb 1039. (2014).