

Supporting information

Cross-species molecular imaging of bile salts and lipids in liver: identification of molecular structural markers in health and disease.

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Special staining of frozen tissue sections

HE

- Fixation: 10 minutes in formalin 10% at 4°C
- Airdry, staining on room temperature
- Dip in demin water
- Haematoxyline Gill III (Prosan)(or Harris): 2 minutes
- Tap water: 8 dips
- Demin water: 8 dips
- Phosphotungstic acid 2 %: 4 dips
- Demin water: 8 dips
- Sodiumacetate 2%: 4 dips
- Demin water: 8 dips
- Eosin 1% (Prosan): 1 min
- Ethanol 96 %: 2 x 8 dips
- Ethanol absolute: 2 x 8 dips
- Xylene: 2 x 1 minute
- Mount with synthetic mounting medium

Masson's Trichrome

Preparation of reagents

- Ponceau de xyloidine 1 %: solve 3 g of Ponceau de Xylidine in 300 ml cooking demin water and let it cook for 1 or 2 minutes, then add 3 ml acetic acid and filter the solution.
- Acid anilin blue 0.5 %: solve 1 g of anilin blue in 200 ml demin water. Add 4 ml acetic acid.
- Phosphotungstic acid 3 %: solve 6 g of phosphotungstic acid in 200 ml demin water.
- Acetic acid 1 %: dilute 2 ml acetic acid in 200 ml demin water
- Mayer's haematoxylin: bought solution (Sigma Aldrich)

Staining:

- Fixation: 10 minutes in formalin 10 % at 4°C
- Airdry, staining on room temperature
- Mayer's haematoxylin: 10 minutes
- Running tap water: 10 minutes
- Rinse in demin water
- Phosphotungstic acid 3 %: 7 minutes
- Rinse in demin water
- Acid anilin blue 0.5 %: 4 minutes
- Acetic acid solution 1 %: dip a few times
- Ethanol 96 %: 2 x 2 minutes
- Ethanol absolute: 2 x 2 minutes
- Xylene: 2 x 2 minutes
- Mount with synthetic mounting medium

Immunohistochemistry

- Dry slides: 15-30 minutes
- Fix section: 10 minutes in acetone -20°C
- Rinse with PBS
- Encircle tissue section with DAKO-Cytomation pen
- Rinse with PBS
- Block endogenous peroxidases with 0.3% H₂O₂ in methanol: 30 minutes
1 ml 30% H₂O₂ + 100 ml methanol
- Rinse with PBS: 3 x 3 minutes
- Block with 5% BSA-PBS: 30 minutes
2 ml PBS + 2 ml 10% BSA
- Incubate with 1:2000 α -Cytokeratin (DAKO Z0622) in DB: 1 hour
- Rinse with PBS: 3 x 3 minutes
- Incubate with 1:500 Goat- α -Rabbit-HRP in DB: 1 hour
- Rinse with PBS: 3 x 3 minutes
- Incubate with AEC
19 ml 0.5M NaAc
171 ml with MQ
9 ml DMSO
Before use; add 69 μ l H₂O₂ + 1 ml 4% DMSO-AEC
- Rinse with demi water: \pm 5 minutes
- Stain with Haematoxylin GIII staining: 1-2 min (liver)
- Rinse with tap water: \pm 5 minutes
- Dry slides
- Mount with DAKO Faramount aqueous mounting medium ready-to-use S3025

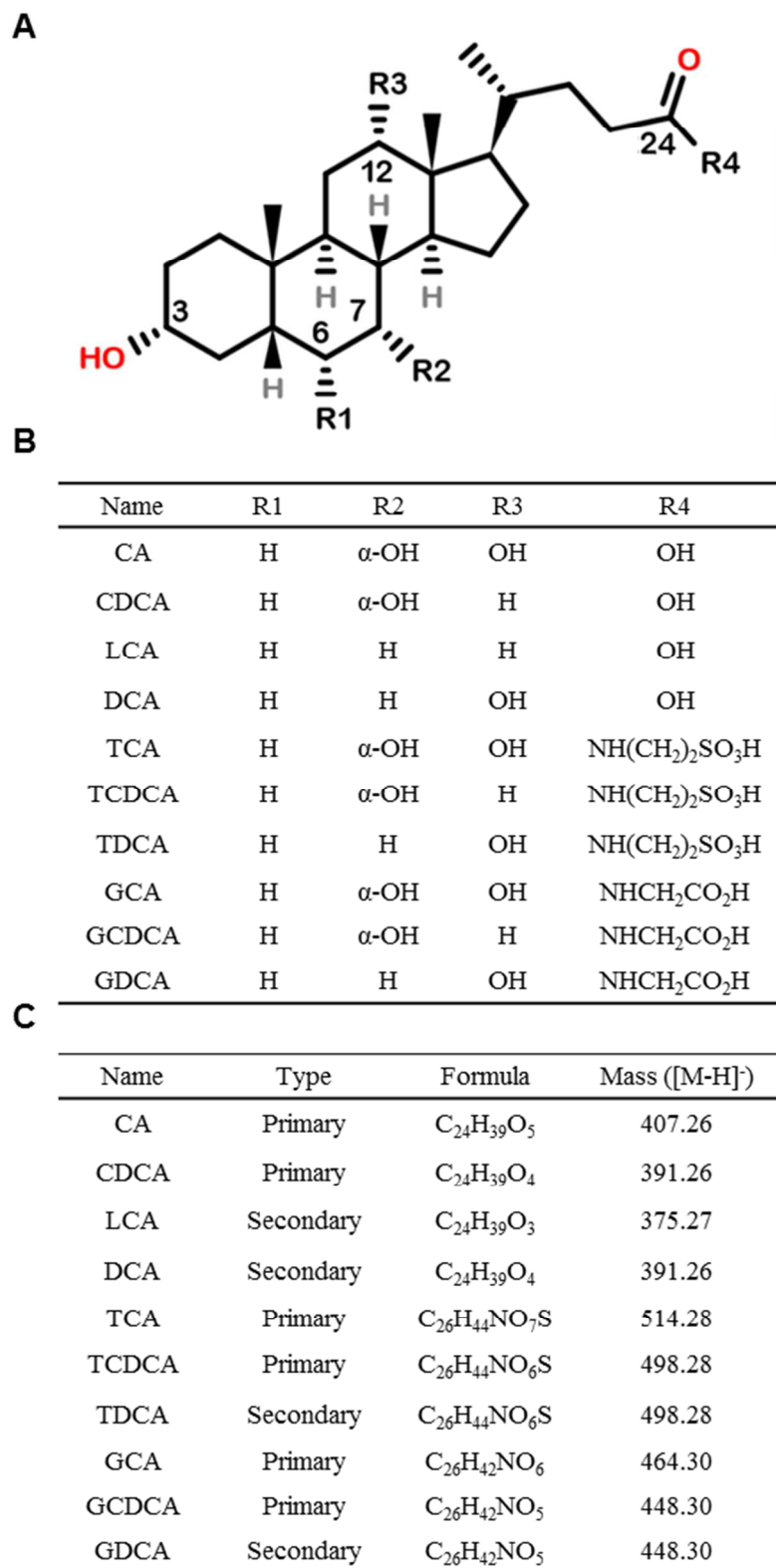


Figure S1. Information on bile acids. (A) Base structure of bile acids and (B) positions of the hydroxyl groups and site of conjugation. (C) Observed masses of bile acid standards in negative ion mode.

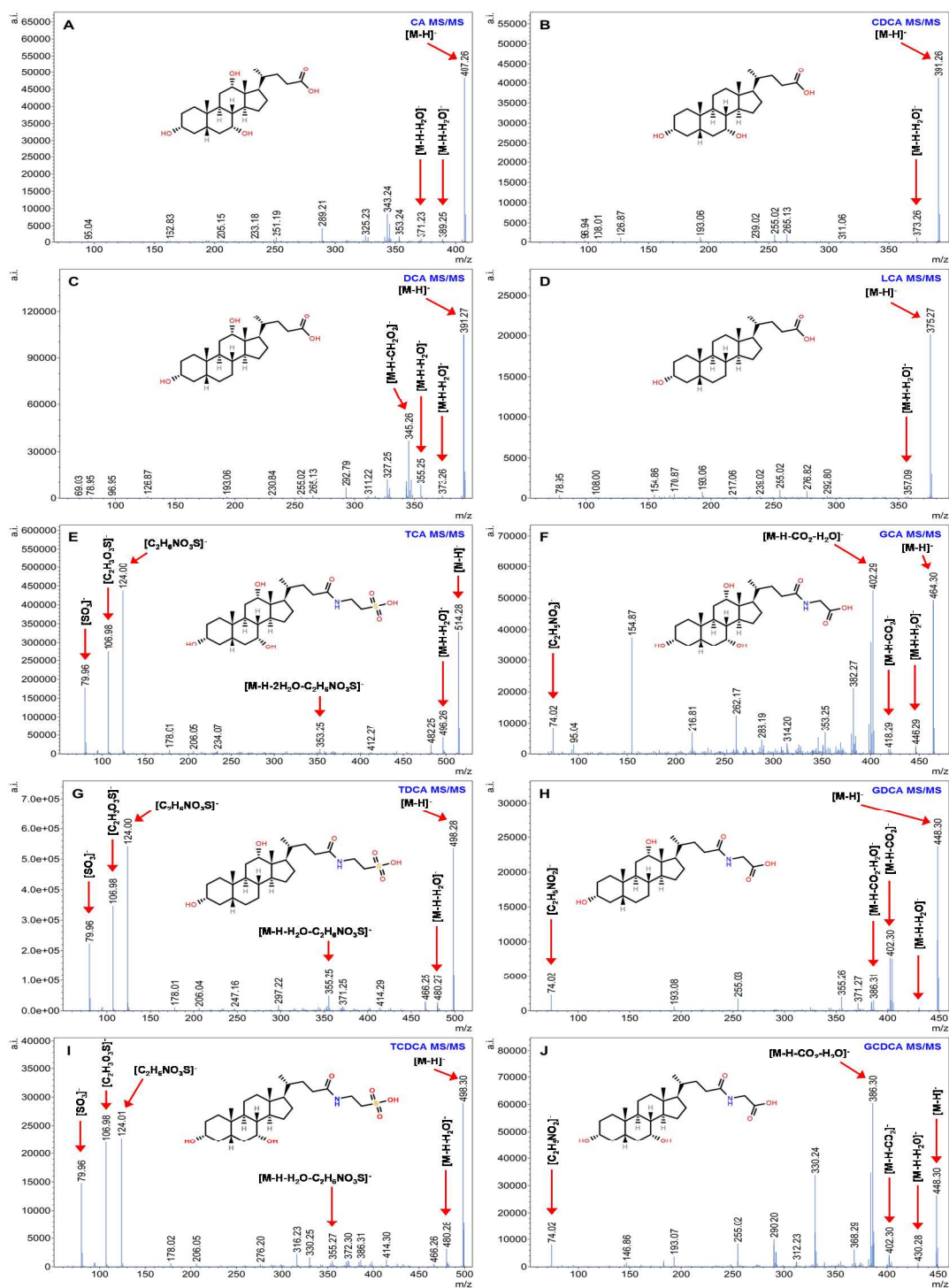


Figure S2. MALDI-MS/MS spectra of deprotonated ($[M-H]^-$) bile acid standards in negative ion mode. (A) cholic acid (m/z 407.26), (B) chenodeoxycholic acid (m/z 391.27), (C) deoxycholic acid (m/z 391.27), (D) lithocholic acid (m/z 375.27), (E) taurocholic acid (m/z 514.28), (F) glycocholic acid (m/z 464.30), (G) taurodeoxycholic acid (m/z 498.30), (H) glycodeoxycholic acid (m/z 448.30), (I) taurochenodeoxycholic acid (m/z 498.30), (J) glycochenodeoxycholic acid (m/z 448.30).

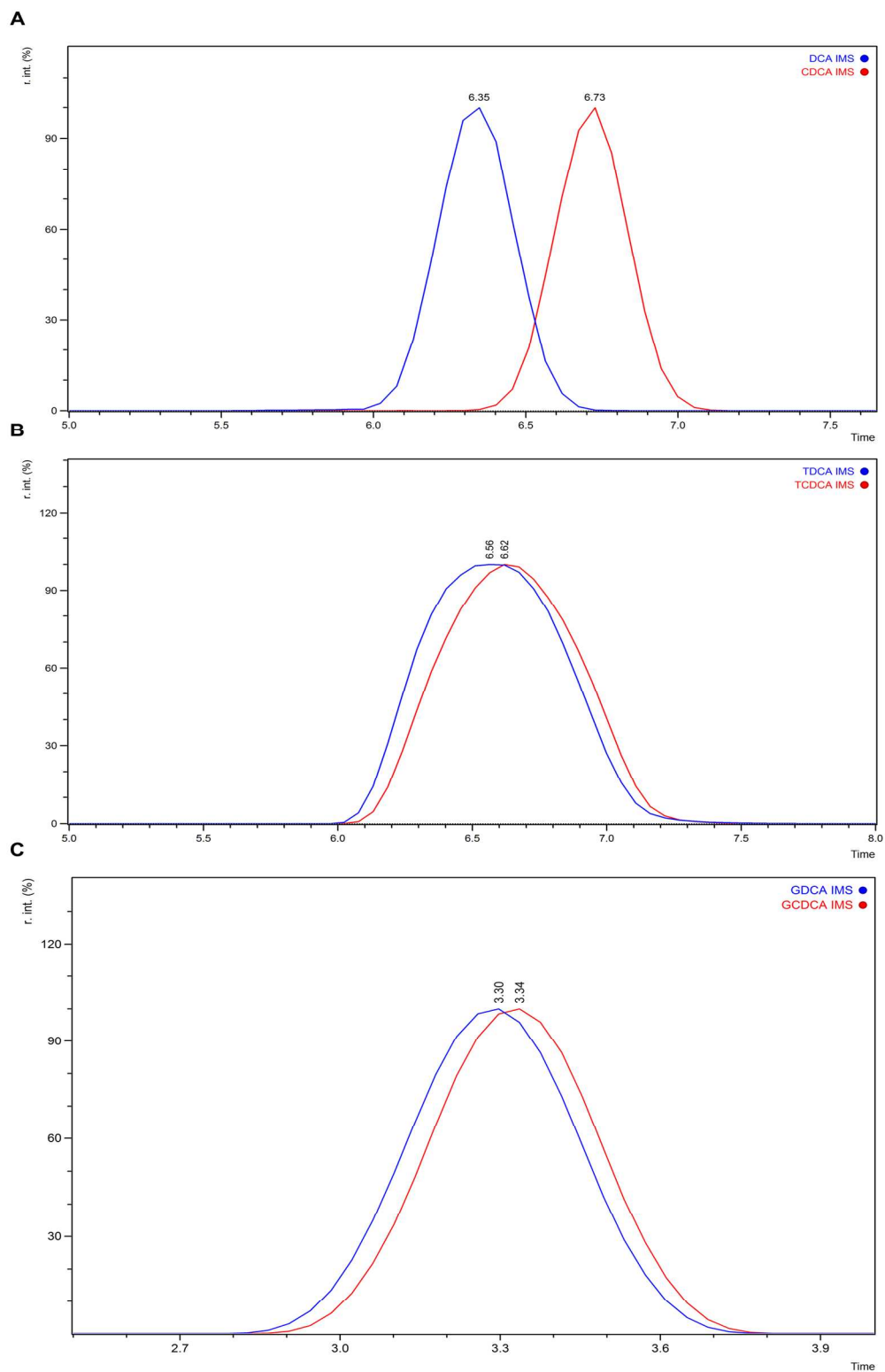


Figure S3. MALDI-IMS-MS of isomeric bile acids in negative ion mode. Mobilograms of (A) DCA/CDCA (m/z 391.27), (B) TDCA/TCDCA (m/z 498.28) and (C) GDCA/GCDCA (m/z 448.30).

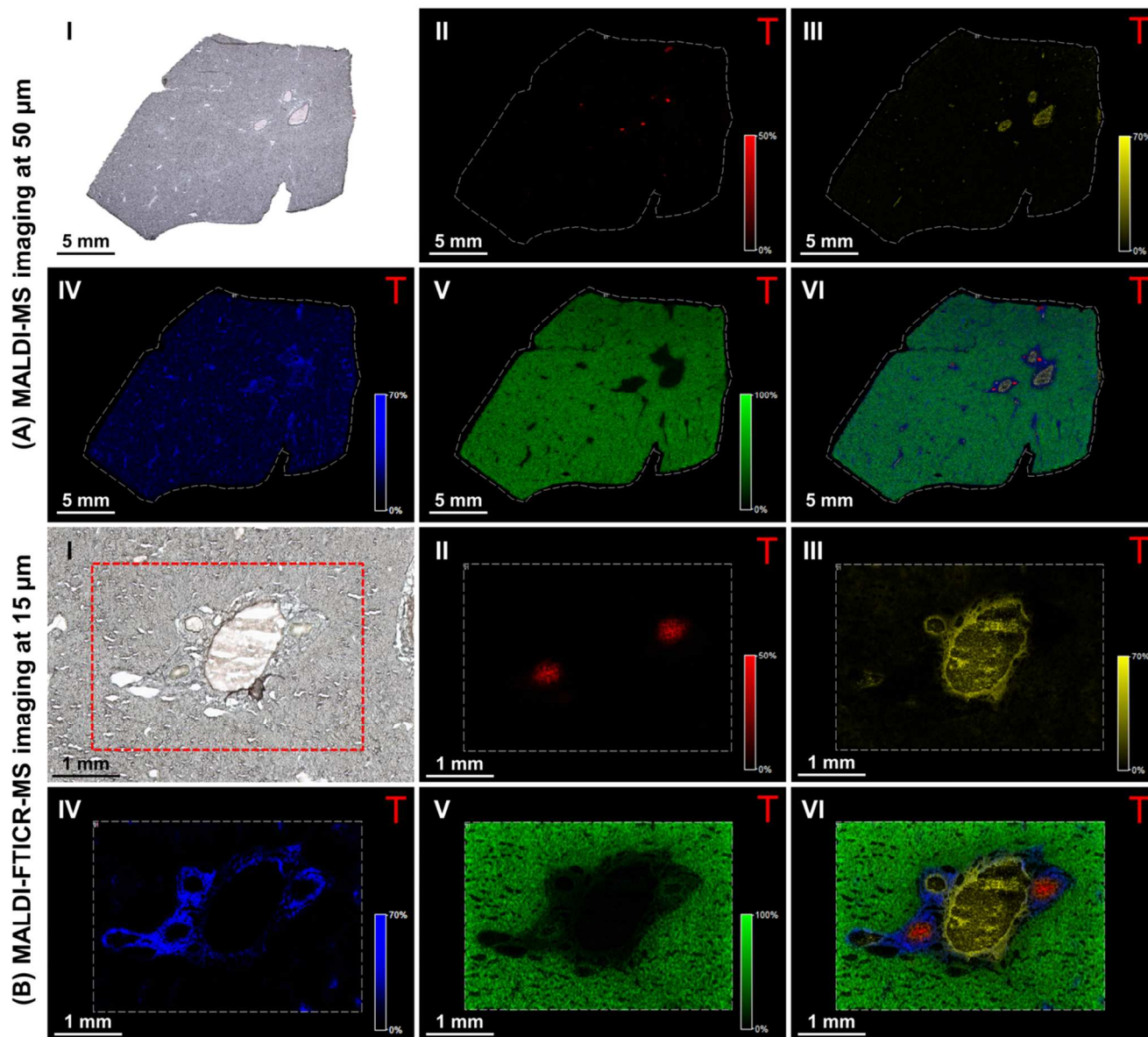
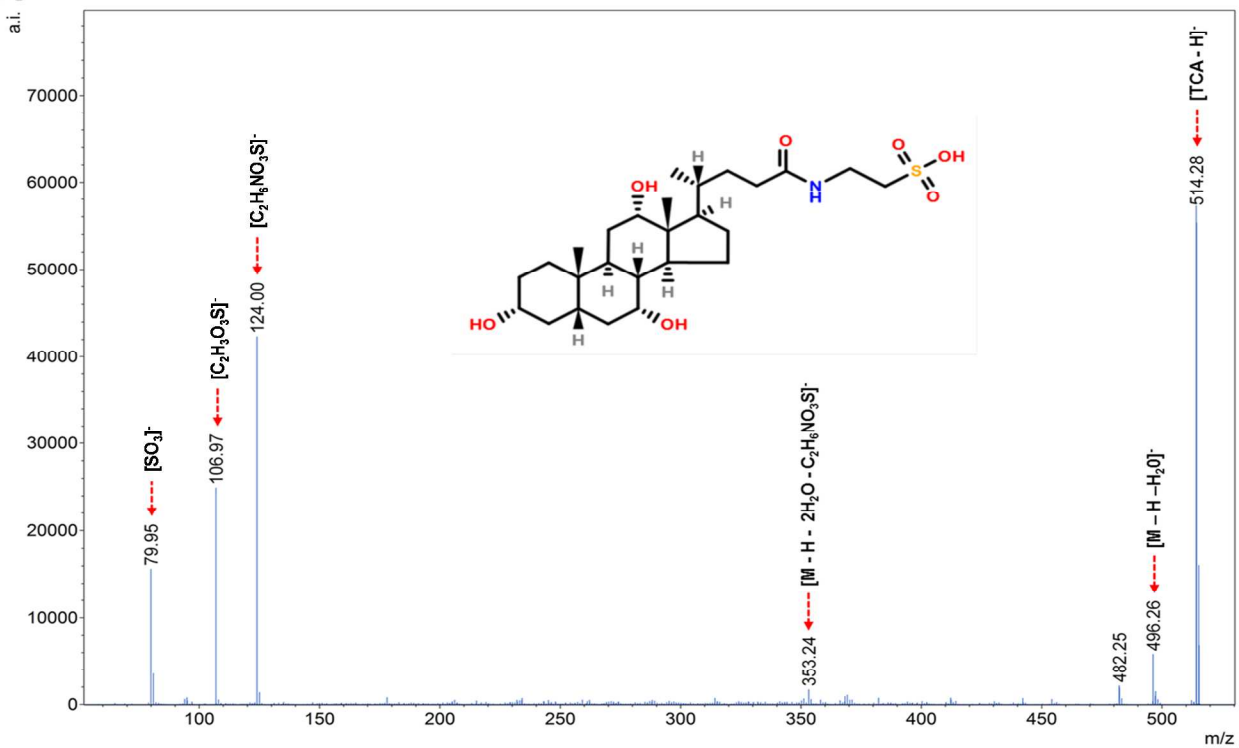
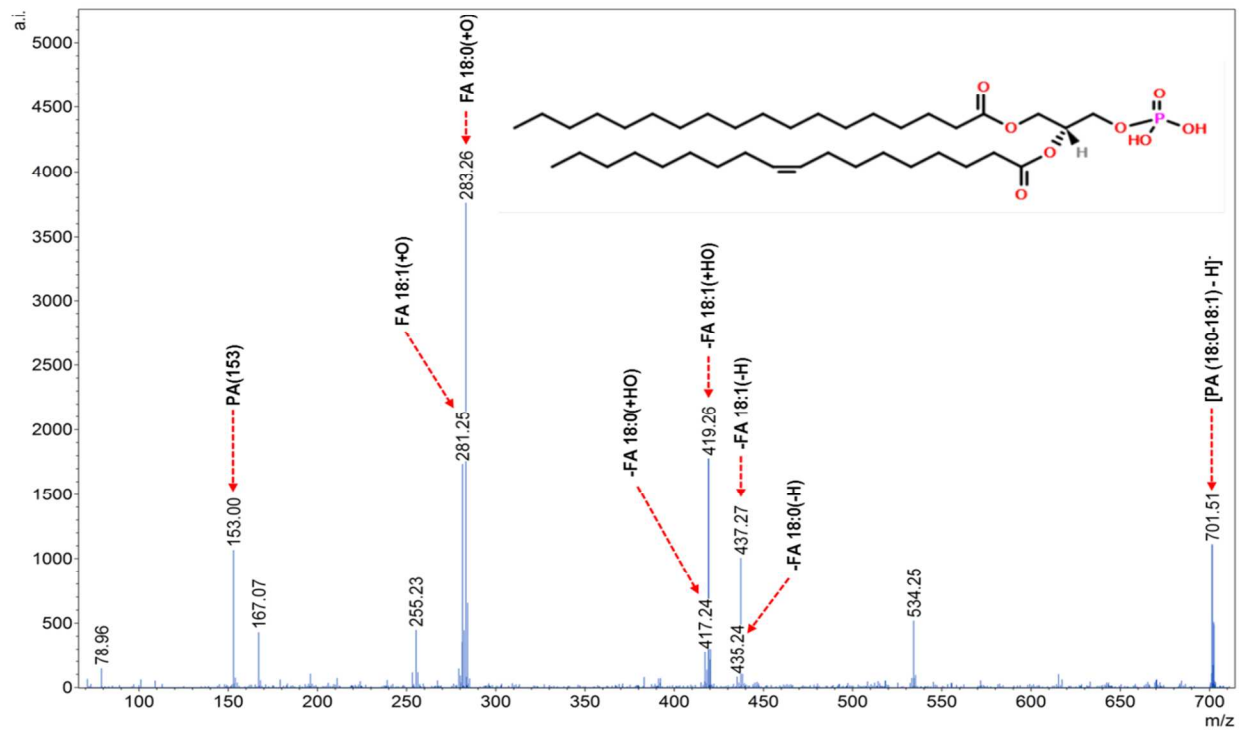


Figure S4. Multimodal imaging of healthy dog liver tissue. (I) Optical image of the tissue section prior to matrix application. Positive-ion mode (A) MALDI-MS and (B) MALDI-FTICR-MS images showing the distribution of selected molecular species in the (II) bile duct lumen (unknown at m/z 576.2418), (III) blood vessels (heme $[M]^+$ at m/z 616.1804), (IV) connective tissue ($[PC(32:0)+K]^+$ at m/z 772.5254), (V) parenchyma ($[PC(38:4)+K]^+$ at m/z 848.5564) and (VI) overlay of the selected species. (Designated masses are from MALDI-FTICR-MS imaging measurements, MALDI-MS image field of view 141828 pixels, area 124 mm² and spatial resolution 50 μm , MALDI-FTICR-MS image field of view 41700 pixels, 9.3 mm² area and spatial resolution 15 μm . Both images normalized with TIC).

A**B**

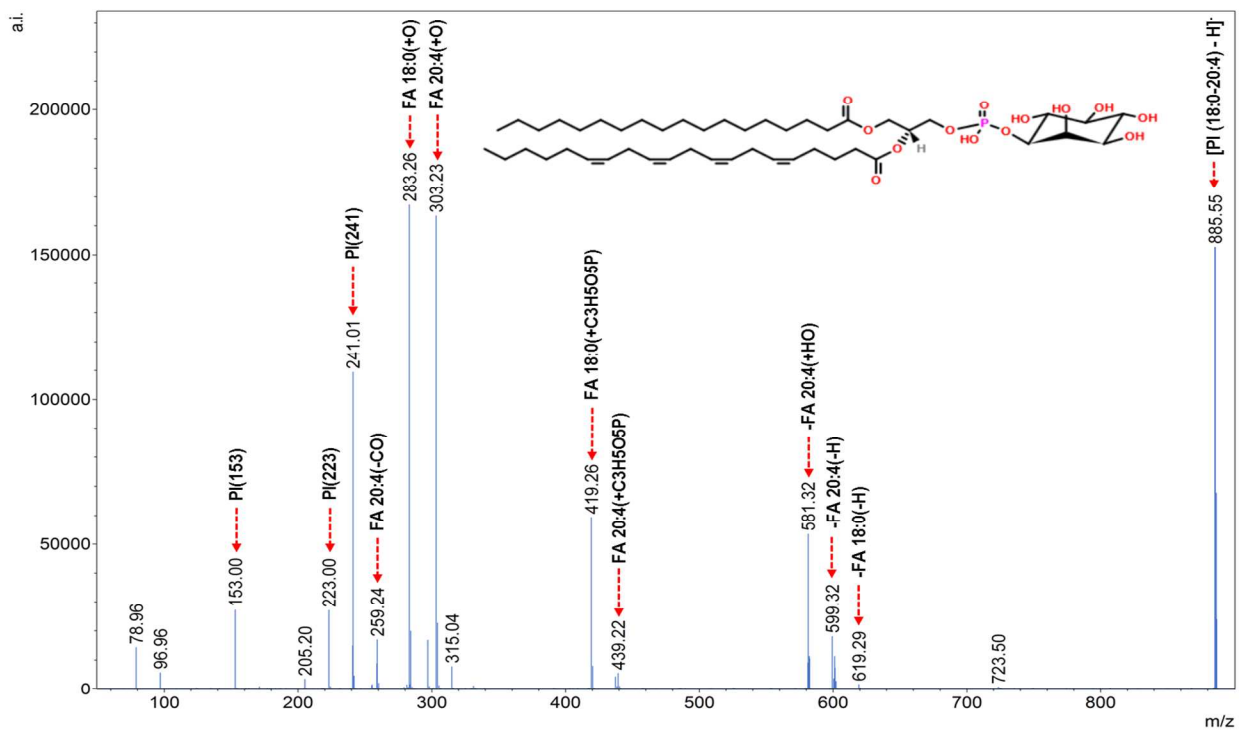
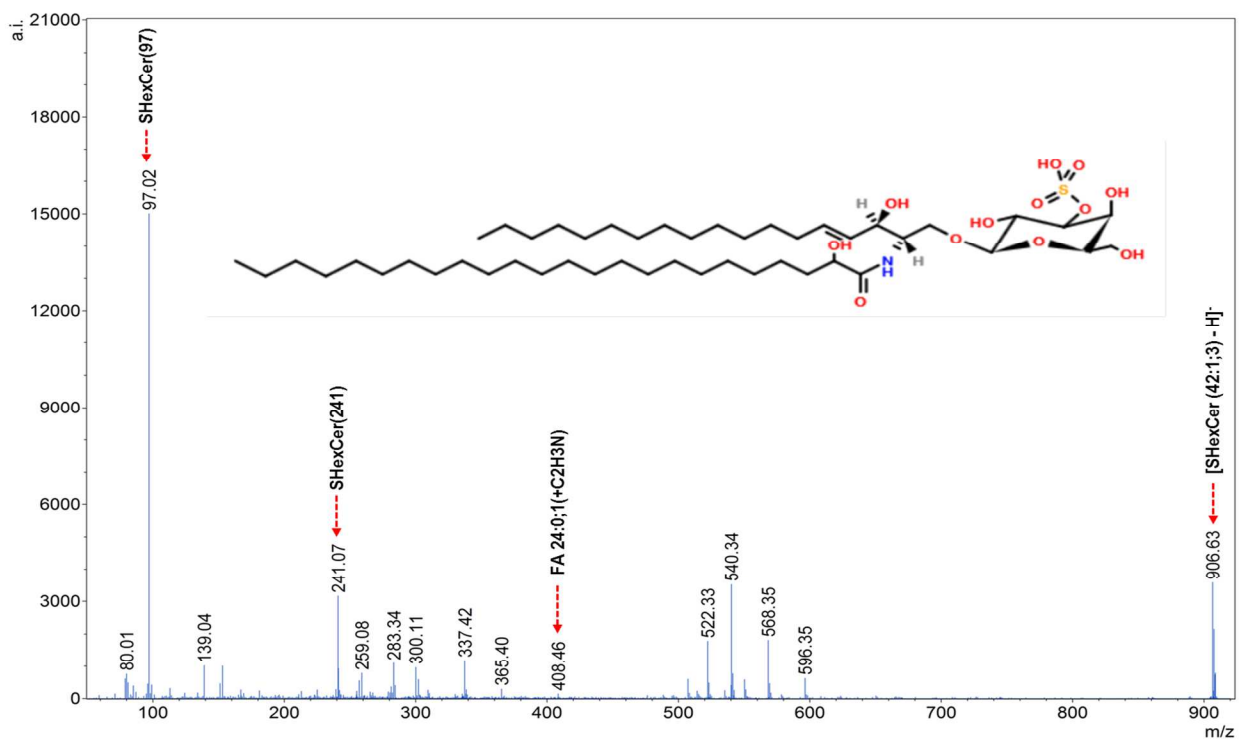
C**D**

Figure S5. MALDI-MS/MS spectra of structural molecular markers for (A) bile duct lumen, (B) connective tissue, (C) parenchyma and (D) bile duct. Spectra obtained from healthy dog liver tissue.

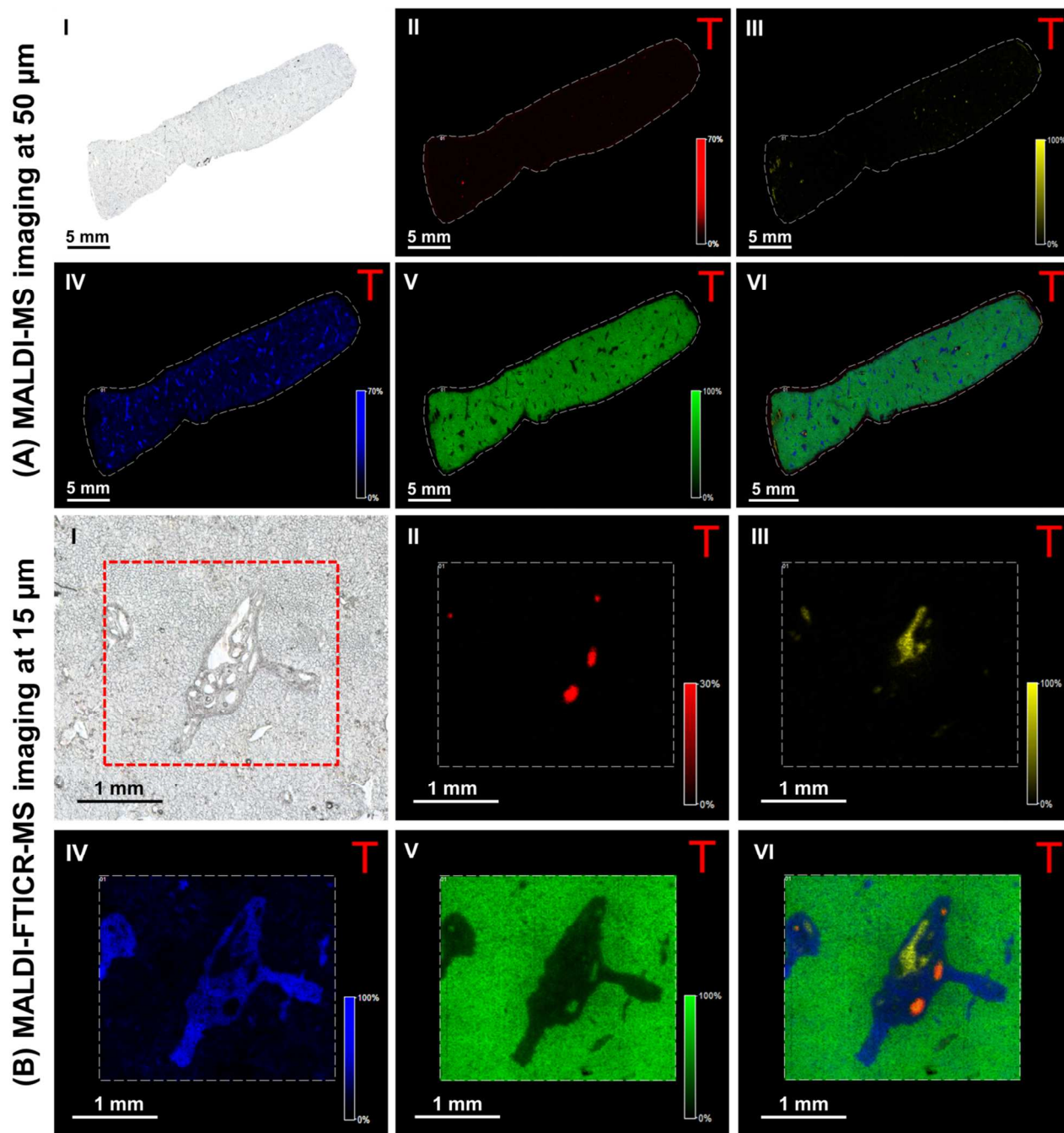


Figure S6. Multimodal imaging of healthy human liver tissue. (I) Optical image of tissue section prior to matrix application. Positive-ion mode (A) MALDI-MS and (B) MALDI-FTICR-MS images showing the distribution of selected molecular species in the (II) bile duct lumen (unknown at m/z 560.2427), (III) blood vessels (heme $[M]^+$ at m/z 616.1782), (IV) connective tissue ($[PC(32:0)+K]^+$ at m/z 772.5254), (V) parenchyma ($[PC(34:2)+K]^+$ at m/z 796.5252) and (VI) overlay of the selected species. (Designated masses are from MALDI-FTICR-MS imaging measurements, MALDI-MS image field of view 72058 pixels, area 110 mm^2 and spatial resolution 50 μm , MALDI-FTICR-MS image field of view 22338 pixels, 5 mm^2 area and spatial resolution 15 μm . Images normalized with TIC).

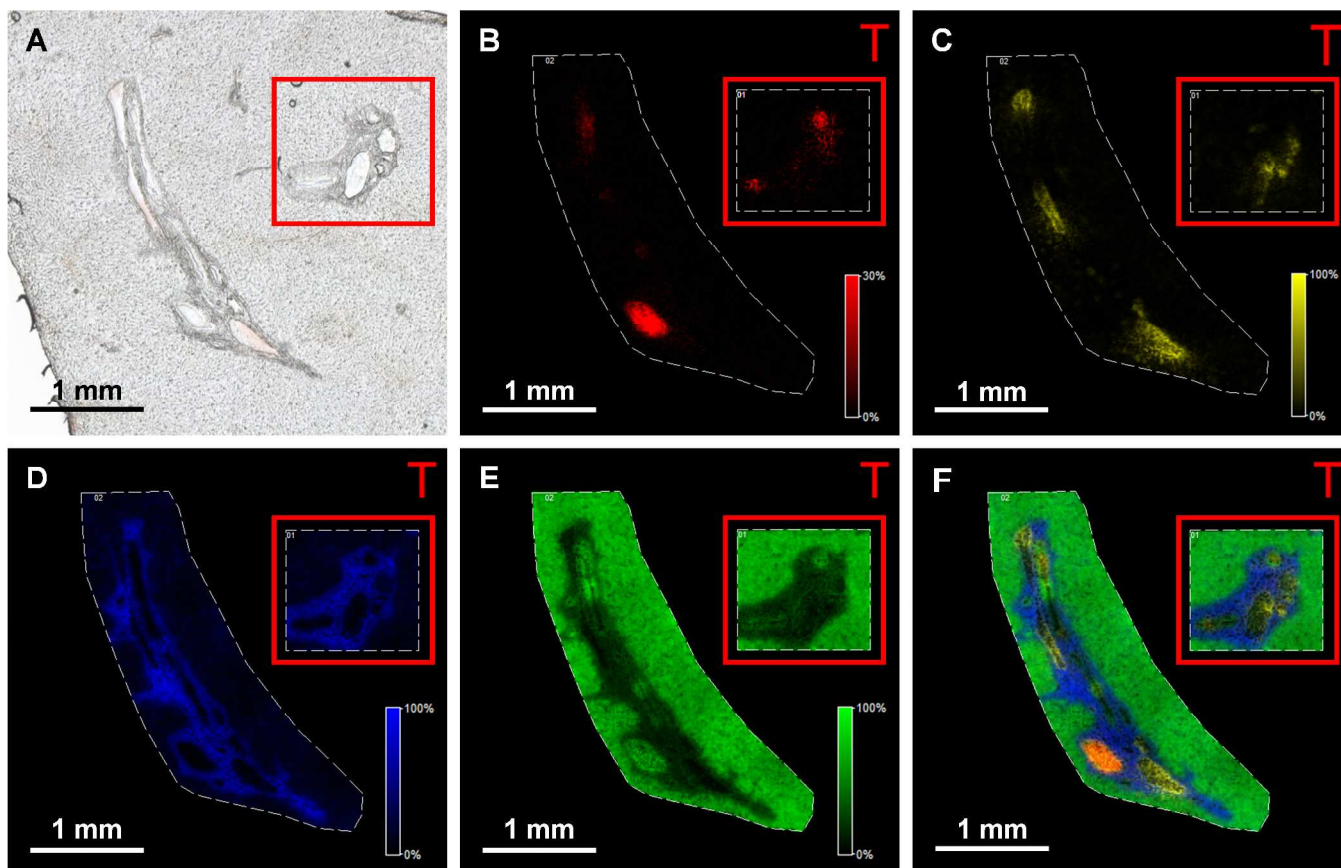


Figure S7. Multimodal imaging of liver from a patient with mild PSC (case 1). (A) Optical image of tissue section prior to matrix application. Positive-ion mode MALDI-FTICR-MS images showing the distribution of selected molecular species in the (B) bile duct lumen (unknown at m/z 560.2434), (C) heme ($[M]^+$) at m/z 616.1804, (D) connective tissue ($[PC (32:0)+K]^+$ at m/z 772.5251), (E) parenchyma ($[PC (34:2)+K]^+$ at m/z 796.5254) and (F) overlay of the selected species. (Designated masses are from MALDI-FTICR-MS imaging measurements, field of view 20238 pixels, area 10 mm^2 and spatial resolution $15 \mu\text{m}$. Images normalized with TIC).

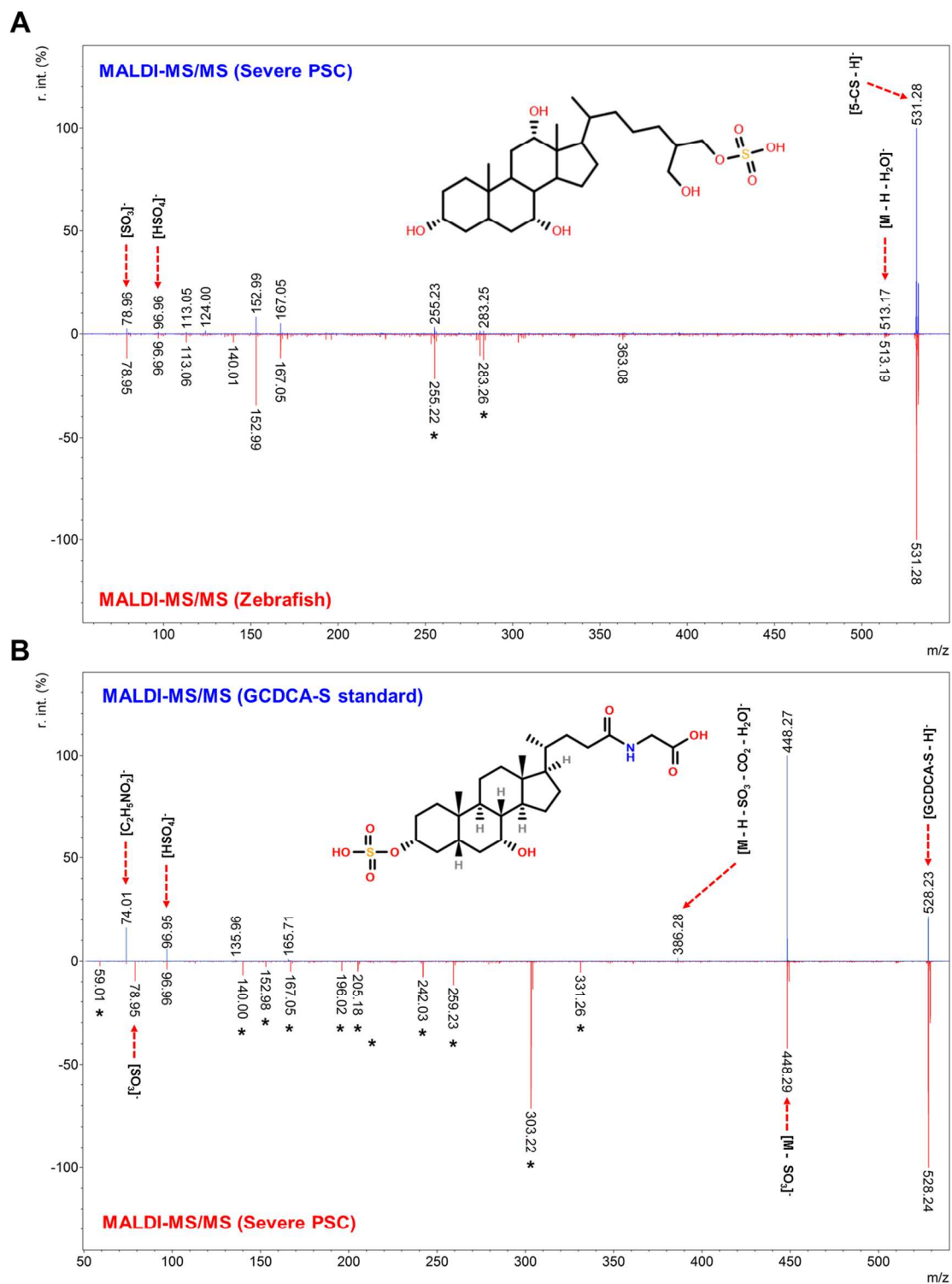


Figure S8. MALDI-MS/MS spectra of 5-CS and GCDCA-S in negative ion mode. A) MALDI-MS/MS spectra of 5-CS at *m/z* 531.3 obtained from severe PSC liver (blue) and whole-body zebrafish (red) tissue sections. B) MALDI-MS/MS spectra of G(C)DCA-S at *m/z* 528.2 obtained from pure standard (blue) and severe PSC liver tissue section (red). * indicated peaks come from unidentified endogenous species tentatively identified as fatty acids.

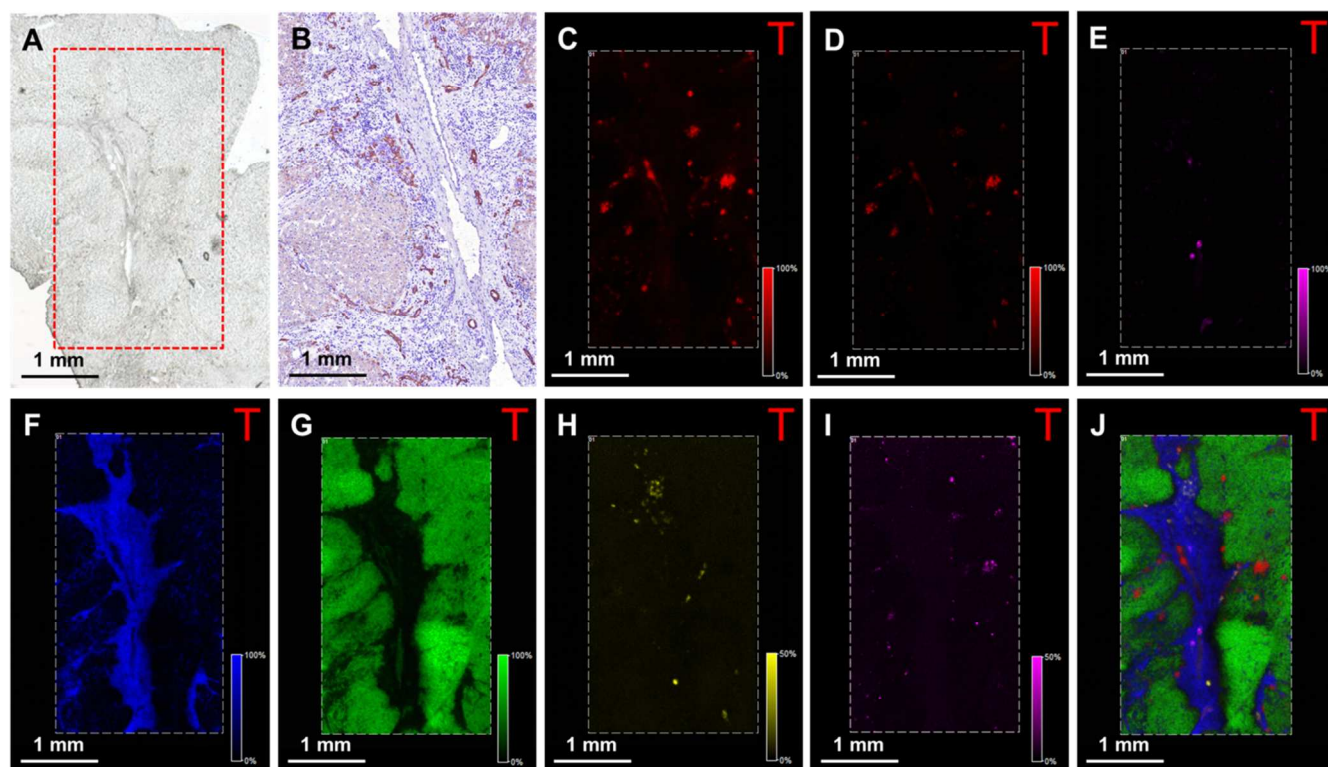


Figure S9. Comparison of immunohistochemical stain and MALDI-MS images from a patient with severe PSC (case 2). (A) Optical image of tissue section prior to matrix application and (B) cytokeratin staining of a consecutive tissue section. Negative-ion mode MALDI-MS images showing the distribution of (C) [TCA-H]⁻ at m/z 514.28, (D) 5-cyprinolsulfate ([M-H]⁻) at m/z 531.30, (E) heme ([M-H]⁻) at m/z 615.17, (F) [PA (18:0_18:1)-H]⁻ at m/z 701.51, (G) [PI (18:0_20:4)-H]⁻ at m/z 885.55, (H) [ST-OH (18:1_24:0)-H]⁻ at m/z 906.63 and (I) bilirubin diglucuronide ([M-H]⁻) at m/z 935.32. (J) Overlay of the selected species. (Field of view 81186 pixels, area 8 mm² and spatial resolution 10 μm. Images normalized with TIC).

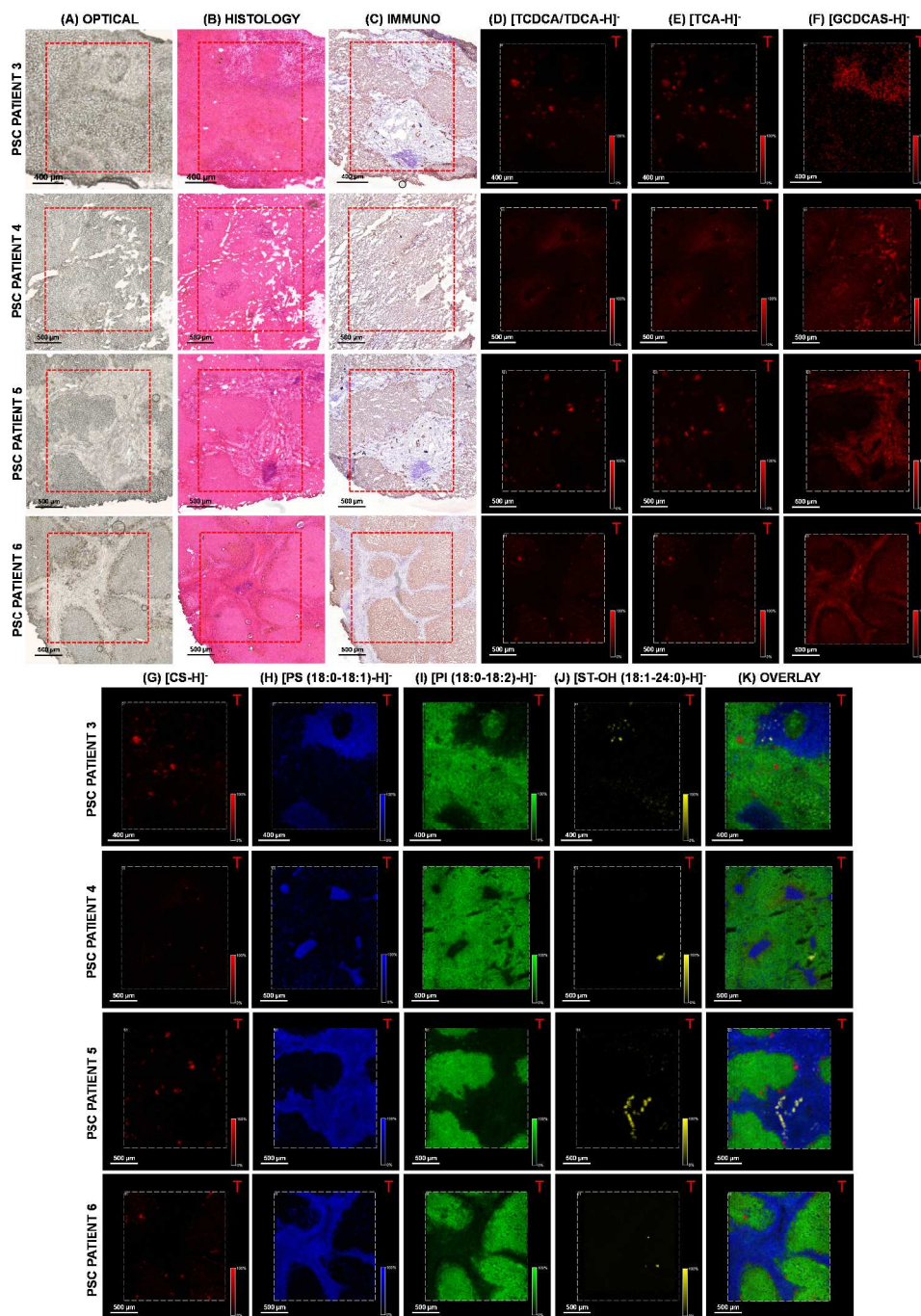


Figure S10. Multimodal imaging of liver tissue from patients with severe PSC (cases 3-6). (A) Optical image of tissue section prior to matrix application, (B) H&E staining of tissue sections post imaging and (C) cytokeratin staining of a consecutive tissue section. Negative-ion mode MALDI-FTICR-MS images showing the distribution of (D) [TCDCA/TDCA-H]⁻ at m/z 498.2896, (E) [TCA-H]⁻ at m/z 514.2844, (F) [G(C)DCAS-H]⁻ at m/z 528.2639, (G) [5-CS-H]⁻ at m/z 531.2998, (H) [PS (18:0_18:1)-H]⁻ at m/z 788.5449, (I) [PI (18:0_18:2)-H]⁻ at m/z 861.5502, (J) [ST-OH (18:1_24:0)-H]⁻ at m/z 906.6352 and (K) overlay of the selected species. (PSC 1-4 fields of view 10356, 25482, 18987 and 21872 pixels and areas 2.3, 5.7, 4.2 and 4.8 mm² respectively. Spatial resolution of all images 15 μ m, normalized with TIC).

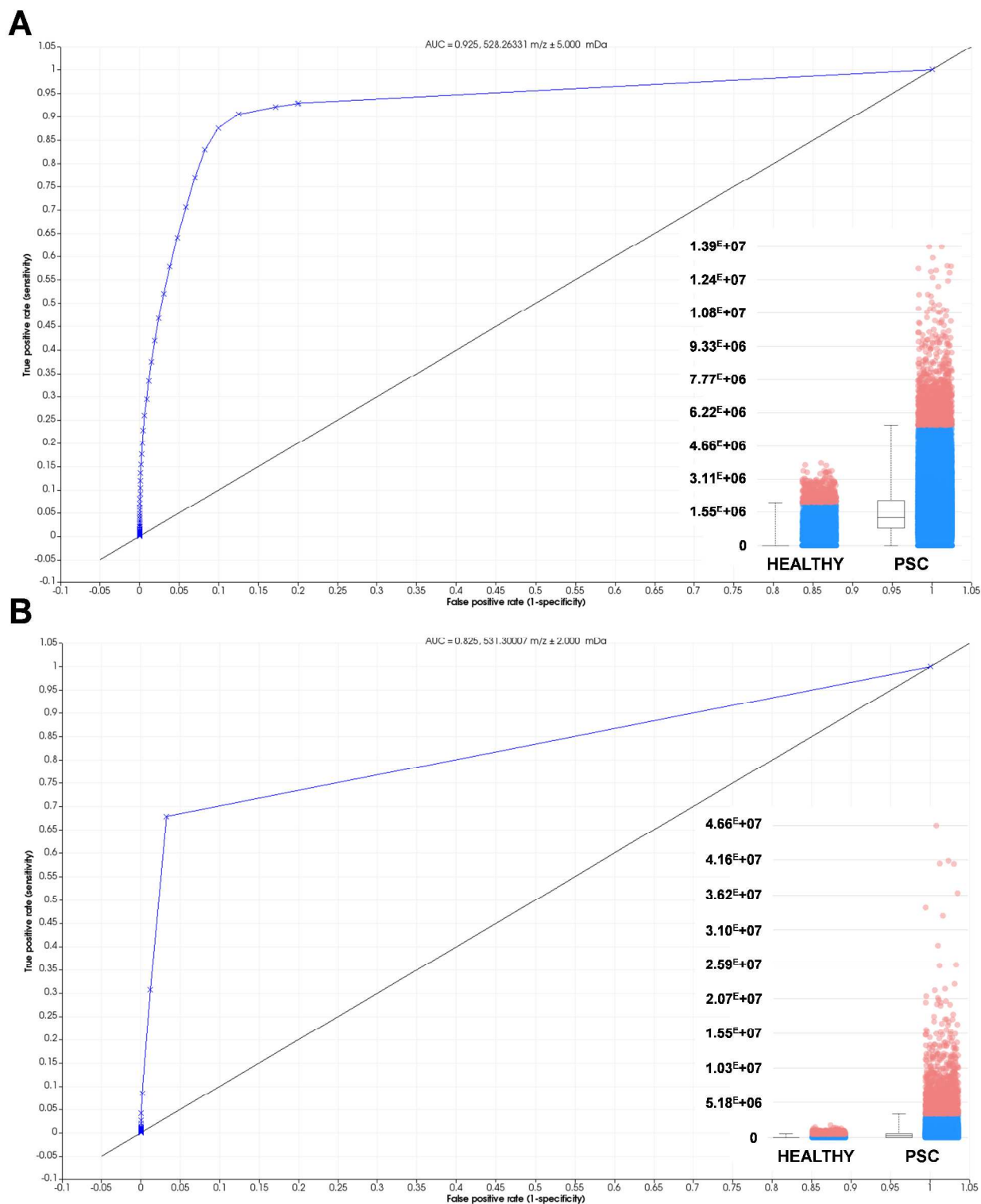


Figure S11. Receiver operating characteristic (ROC) analysis of A) G(C)DCA-S at m/z 528.2642 and B) 5-CS at m/z 531.2999 from healthy liver tissue and tissue from severe PSC cases. Inserts show the relative ion intensity of the displayed ions.

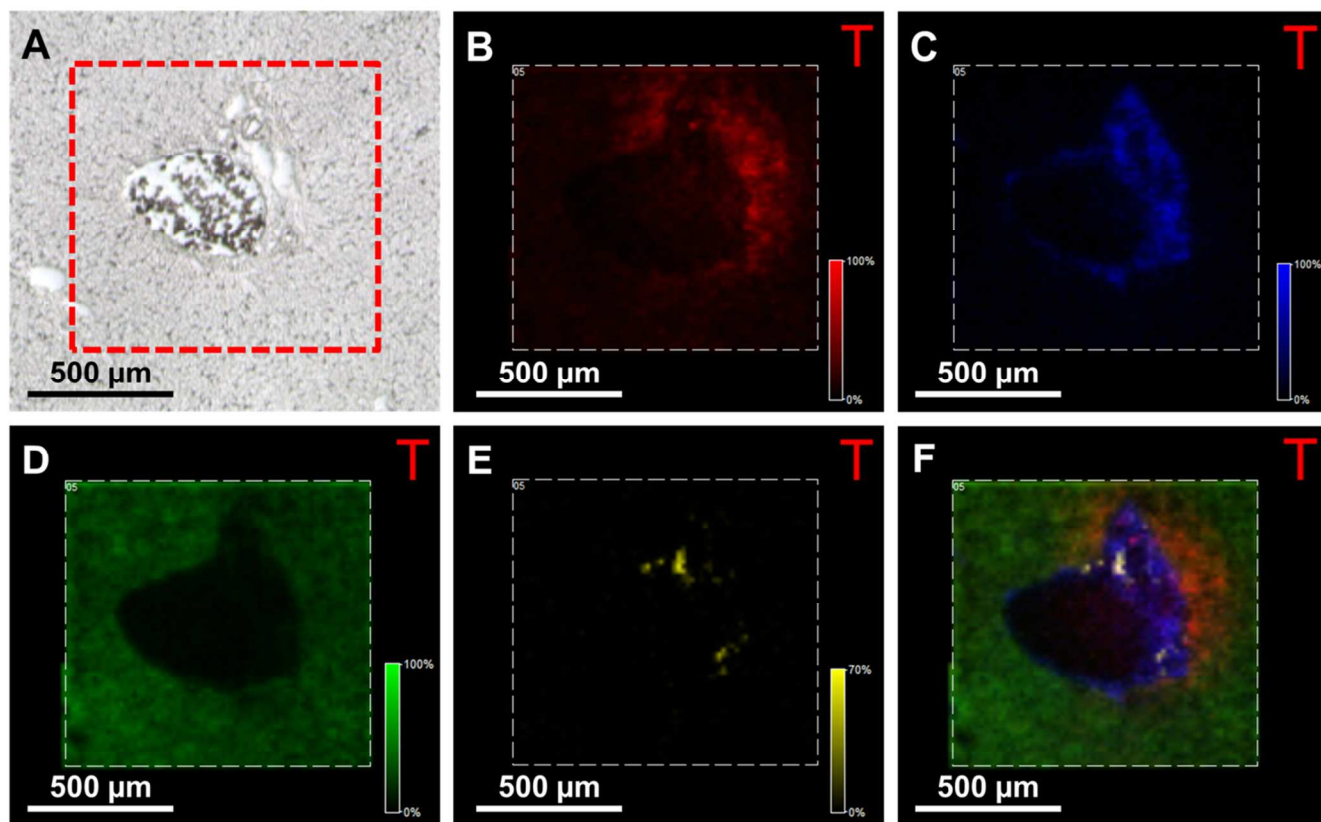


Figure S12. MALDI-FTICR-MS imaging of healthy rat liver tissue. (A) Optical image of tissue section prior to matrix application. Negative-ion mode MALDI-FTICR-MS images showing the distribution of selected molecular species in the (B) bile duct lumen ($[\text{TCA-H}]^-$ at m/z 514.2845), (C) connective tissue ($[\text{PA (18:0}_{18:1})\text{-H}]^-$ at m/z 701.5129), (D) parenchyma ($[\text{PI (18:0}_{20:4})\text{-H}]^-$ at m/z 885.5499), (E) bile duct ($[\text{ST-OH (18:1}_{24:0})\text{-H}]^-$ at m/z 906.6348) and (F) overlay of the selected species. (Designated masses are from MALDI-FTICR-MS imaging measurements, field of view 4352 pixels, area 1 mm² and spatial resolution 15 μm. Images normalized with TIC).

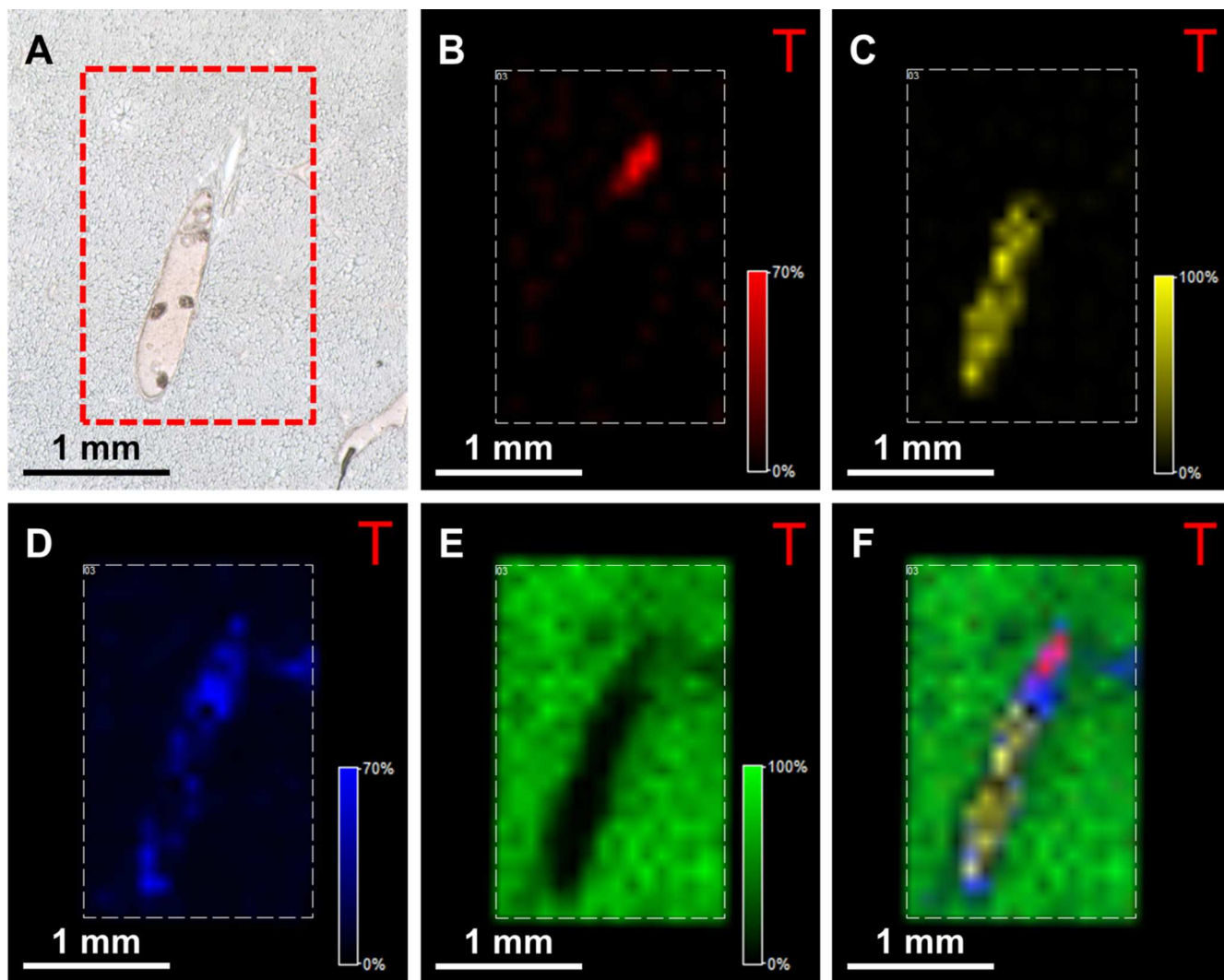


Figure S13. MALDI-FTICR-MS imaging of healthy rat liver tissue. (A) Optical image of tissue section prior to matrix application. Positive-ion mode MALDI-FTICR-MS images showing the distribution of selected molecular species in the (B) bile duct lumen (unknown at m/z 560.2414), (C) heme ($[M]^+$) at m/z 616.1775, (D) connective tissue ($[PC (32:0)+K]^+$) at m/z 772.5254), (E) parenchyma ($[PC (38:4)+K]^+$) at m/z 848.5566) and (F) overlay of the selected species. (Designated masses are from MALDI-FTICR-MS imaging measurements, field of view 400 pixels, area 3.8 mm^2 and spatial resolution $100 \mu\text{m}$. Images normalized with TIC).

<i>Sample</i>	<i>Bilirubin (umol/L)</i>	<i>ALT (U/L)</i>	<i>AST (U/L)</i>	<i>ALP (U/L)</i>	<i>GGT (U/L)</i>	<i>INR</i>
1 (mild)	15	15	19	N/A	21	1.05
2 (severe)	48*	76*	79*	365*	229*	1.1
3 (mild)	7	48*	56*	52	61*	N/A
4 (mild/severe)	56*	39	46*	110	21	1.6*
5 (severe)	80*	56*	83*	373*	146*	N/A
6 (severe)	147*	157*	258*	240*	104*	1.4*

Key: Alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), gamma glutamyltransferase (GGT), international normalized ratio (INR). * Represents values above the reference range.

Table S1. Serum biochemistry of PSC patients.

CONNECTIVE TISSUE				PARENCHYMA				CONNECTIVE TISSUE				PARENCHYMA			
Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula
741.5309	2.36E+10	ISM (34:1)H ⁺ K ⁺	C39H79N2O6PNa	741.5308	1.34E+10	ISM (34:1)H ⁺ K ⁺	C39H79N2O6PNa	725.5570	1.61E+10	ISM (34:1)H ⁺ Na ⁺	C39H79N2O6PNa	832.5825	1.27E+09	IPC (34:1)H ⁺ K ⁺	C42H82NO8PNa
851.6399	2.34E+10	C47H93N2O6PNa	C47H93N2O6PNa	772.5251	1.05E+10	IPC (32:0)H ⁺ K ⁺	C40H80NO8PNa	756.5518	7.65E+09	IPC (32:0)H ⁺ Na ⁺	C40H80NO8PNa	848.5564	4.68E+09	IPC (38:4)H ⁺ K ⁺	C46H84NO8PNa
725.5568	2.24E+10	ISM (34:1)H ⁺ Na ⁺	C39H79N2O6PNa	851.6407	8.95E+09	ISM (42:2)H ⁺ K ⁺	C47H93N2O6PNa	835.6663	7.47E+09	ISM (42:2)H ⁺ Na ⁺	C47H93N2O6PNa	810.6008	2.93E+09	IPC (18:0) 20:4)H ⁺ H ⁺	IPC (18:0) 20:4)H ⁺ H ⁺
835.6658	2.03E+10	ISM (42:2)H ⁺ Na ⁺	C47H93N2O6PNa	725.5569	7.83E+09	ISM (34:1)H ⁺ Na ⁺	C39H79N2O6PNa	741.5311	6.84E+09	ISM (34:1)H ⁺ K ⁺	C39H79N2O6PNa	792.5690	2.23E+09	IPC (16:0) 20:4)H ⁺ H ⁺	IPC (16:0) 20:4)H ⁺ H ⁺
772.5254	1.96E+10	IPC (32:0)H ⁺ K ⁺	C40H80NO8PNa	756.5518	6.50E+09	IPC (32:0)H ⁺ Na ⁺	C40H80NO8PNa	753.5896	5.34E+09	ISM (36:1)H ⁺ Na ⁺	C41H83NO8PNa	796.5293	1.98E+09	IPC (34:2)H ⁺ K ⁺	IPC (34:2)H ⁺ K ⁺
756.5508	1.59E+10	IPC (32:0)H ⁺ Na ⁺	C40H80NO8PNa	825.6243	5.95E+09	IPC (40:1)H ⁺ K ⁺	C45H91N2O6PNa	808.6510	4.44E+09	ISM (40:1)H ⁺ Na ⁺	C40H80NO8PNa	846.5413	1.55E+09	IPC (38:5)H ⁺ K ⁺	IPC (38:5)H ⁺ K ⁺
734.5691	7.59E+09	IPC (16:0) 16:0)H ⁺ H ⁺	C40H81NO8PNa	835.6659	4.80E+09	ISM (40:1)H ⁺ K ⁺	C45H91N2O6PNa	851.6405	3.64E+09	ISM (42:2)H ⁺ K ⁺	C47H93N2O6PNa	824.5565	1.54E+09	IPC (38:6)H ⁺ K ⁺	IPC (38:6)H ⁺ K ⁺
703.5749	6.53E+09	ISM (18:1) 16:0)H ⁺ H ⁺	C39H80N2O6PNa	734.5694	4.77E+09	IPC (16:0) 16:0)H ⁺ H ⁺	C40H81NO8PNa	703.5750	3.34E+09	IPC (16:0) 16:0)H ⁺ H ⁺	C39H80N2O6PNa	844.5293	1.51E+09	IPC (38:4)H ⁺ Na ⁺	IPC (38:4)H ⁺ Na ⁺
769.5626	6.18E+09	ISM (36:1)H ⁺ K ⁺	C41H83NO8PNa	703.5744	3.78E+09	ISM (18:1) 16:0)H ⁺ H ⁺	C39H80N2O6PNa	800.5570	3.89E+08	IPC (34:0)H ⁺ K ⁺	C41H83NO8PNa	832.5827	1.45E+09	IPC (38:4)H ⁺ Na ⁺	IPC (38:4)H ⁺ Na ⁺
762.6005	8.07E+08	IPC (16:0) 18:0)H ⁺ H ⁺	C42H85NO8PNa	809.6505	3.21E+09	ISM (40:1)H ⁺ Na ⁺	C45H91N2O6PNa	820.5255	2.81E+10	IPC (36:4)H ⁺ K ⁺	C44H80NO8PNa	796.5409	1.27E+09	IPC (34:1)H ⁺ K ⁺	IPC (34:1)H ⁺ K ⁺

CONNECTIVE TISSUE				PARENCHYMA				CONNECTIVE TISSUE				PARENCHYMA			
Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula
796.5252	2.54E+11	IPC (34:2)H ⁺ K ⁺	C42H80NO8PNa	796.5254	8.38E+10	IPC (34:2)H ⁺ K ⁺	C42H80NO8PNa	725.5570	1.61E+10	ISM (34:1)H ⁺ Na ⁺	C39H79N2O6PNa	772.5254	4.68E+08	IPC (32:0)H ⁺ K ⁺	C40H80NO8PNa
789.5410	1.44E+11	IPC (34:1)H ⁺ K ⁺	C42H80NO8PNa	782.5692	1.39E+10	IPC (16:0) 20:4)H ⁺ H ⁺	C42H81NO8PNa	756.5518	7.65E+09	IPC (32:0)H ⁺ Na ⁺	C40H80NO8PNa	851.6403	3.17E+08	ISM (42:2)H ⁺ K ⁺	C47H93N2O6PNa
780.5513	1.42E+11	IPC (34:2)H ⁺ Na ⁺	C42H80NO8PNa	798.5410	1.33E+10	IPC (34:1)H ⁺ K ⁺	C44H80NO8PNa	741.5311	6.84E+09	ISM (34:1)H ⁺ K ⁺	C39H79N2O6PNa	734.5694	2.85E+08	IPC (16:0) 18:0)H ⁺ H ⁺	IPC (16:0) 18:0)H ⁺ H ⁺
824.5561	1.11E+11	IPC (36:2)H ⁺ K ⁺	C44H80NO8PNa	780.5514	1.24E+10	IPC (36:4)H ⁺ K ⁺	C44H80NO8PNa	753.5896	5.34E+09	ISM (36:1)H ⁺ Na ⁺	C41H83NO8PNa	825.6249	2.24E+08	ISM (40:1)H ⁺ K ⁺	ISM (40:1)H ⁺ K ⁺
820.5248	8.76E+10	IPC (18:0) 20:5)H ⁺ H ⁺	C46H83NO8PNa	808.5851	9.35E+09	IPC (18:0) 20:5)H ⁺ H ⁺	C46H83NO8PNa	808.6510	4.44E+09	ISM (40:1)H ⁺ Na ⁺	C40H80NO8PNa	756.5515	1.74E+08	IPC (32:0)H ⁺ Na ⁺	IPC (32:0)H ⁺ Na ⁺
808.5847	7.15E+10	IPC (16:0) 18:2)H ⁺ H ⁺	C44H82NO8PNa	824.5512	5.17E+09	IPC (36:2)H ⁺ K ⁺	C44H82NO8PNa	851.6405	3.64E+09	ISM (42:2)H ⁺ K ⁺	C47H93N2O6PNa	725.5568	1.38E+08	ISM (34:1)H ⁺ Na ⁺	ISM (34:1)H ⁺ Na ⁺
758.5695	7.15E+10	IPC (16:0) 18:2)H ⁺ H ⁺	C44H82NO8PNa	804.5512	5.72E+09	IPC (36:4)H ⁺ Na ⁺	C44H83NO8PNa	772.5254	3.34E+09	IPC (32:0)H ⁺ K ⁺	C40H80NO8PNa	800.5569	1.13E+08	IPC (34:0)H ⁺ K ⁺	IPC (34:0)H ⁺ K ⁺
822.5406	6.76E+10	IPC (36:3)H ⁺ K ⁺	C44H83NO8PNa	820.5252	5.86E+09	IPC (36:4)H ⁺ K ⁺	C44H83NO8PNa	703.5750	3.02E+09	ISM (18:1) 16:0)H ⁺ H ⁺	C41H83NO8PNa	769.5623	8.69E+07	ISM (36:1)H ⁺ K ⁺	ISM (36:1)H ⁺ K ⁺
806.5672	4.83E+10	IPC (36:3)H ⁺ Na ⁺	C44H83NO8PNa	822.5411	4.79E+09	IPC (36:3)H ⁺ K ⁺	C44H82NO8PNa	734.5695	3.02E+09	IPC (16:0) 16:0)H ⁺ H ⁺	C42H85NO8PNa	762.6008	4.30E+07	IPC (16:0) 16:0)H ⁺ H ⁺	IPC (16:0) 16:0)H ⁺ H ⁺
848.5562	4.53E+10	IPC (38:4)H ⁺ K ⁺	C46H84NO8PNa	760.5851	4.49E+09	IPC (16:0) 18:1)H ⁺ H ⁺	C42H83NO8PNa								

CONNECTIVE TISSUE				PARENCHYMA				CONNECTIVE TISSUE				PARENCHYMA			
Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula	Mass	Intensity	Identification	Formula
796.5252	2.54E+11	IPC (34:2)H ⁺ K ⁺	C42H80NO8PNa	796.5254	8.38E+10	IPC (34:2)H ⁺ K ⁺	C42H80NO8PNa	725.5570	1.61E+10	ISM (34:1)H ⁺ Na ⁺	C39H79N2O6PNa	772.5254	4.68E+08	IPC (32:0)H ⁺ K ⁺	C40H80NO8PNa
789.5410	1.44E+11	IPC (34:1)H ⁺ K ⁺	C42H80NO8PNa	782.5692	1.39E+10	IPC (16:0) 20:4)H ⁺ H ⁺	C42H81NO8PNa	756.5518	7.65E+09	IPC (32:0)H ⁺ Na ⁺	C40H80NO8PNa	851.6403	3.17E+08	ISM (42:2)H ⁺ K ⁺	C47H93N2O6PNa
780.5513	1.42E+11	IPC (34:2)H ⁺ Na ⁺	C42H80NO8PNa	798.5410	1.33E+10	IPC (34:1)H ⁺ K ⁺	C44H80NO8PNa	741.5311	6.84E+09	ISM (34:1)H ⁺ K ⁺	C39H79N2O6PNa	734.5694	2.85E+08	IPC (16:0) 18:0)H ⁺ H ⁺	IPC (16:0) 18:0)H ⁺ H ⁺
824.5561	1.11E+11	IPC (36:2)H ⁺ K ⁺	C44H80NO8PNa	780.5514	1.24E+10	IPC (36:4)H ⁺ K ⁺	C44H80NO8PNa	753.5896	5.34E+09	ISM (36:1)H ⁺ Na ⁺	C41H83NO8PNa	825.6249	2.24E+08	ISM (40:1)H ⁺ K ⁺	ISM (40:1)H ⁺ K ⁺
820.5248	8.76E+10	IPC (18:0) 20:5)H ⁺ H ⁺	C46H83NO8PNa	808.5851	9.35E+09	IPC (18:0) 20:5)H ⁺ H ⁺	C46H83NO8PNa	808.6510	4.44E+09	ISM (40:1)H ⁺ Na ⁺	C40H80NO8PNa	756.5515	1.74E+08	IPC (32:0)H ⁺ Na ⁺	IPC (32:0)H ⁺ Na ⁺
808.5847	7.15E+10	IPC (16:0) 18:2)H ⁺ H ⁺	C44H82NO8PNa	824.5512	5.17E+09	IPC (36:2)H ⁺ K ⁺	C44H82NO8PNa	851.6405	3.64E+09	ISM (42:2)H ⁺ K ⁺	C47H93N2O6PNa	725.5568	1.38E+08	ISM (34:1)H ⁺ Na ⁺	ISM (34:1)H ⁺ Na ⁺
758.5695	7.15E+10	IPC (16:0) 18:2)H ⁺ H ⁺	C44H82NO8PNa	804.5512	5.72E+09	IPC (36:4)H ⁺ Na ⁺	C44H83NO8PNa	772.5254	3.34E+09	IPC (32:0)H ⁺ K ⁺	C40H80NO8PNa	800.5569	1.13E+08	IPC (34:0)H ⁺ K ⁺	IPC (34:0)H ⁺ K ⁺
822.5406	6.76E+10	IPC (36:3)H ⁺ K ⁺	C44H83NO8PNa	820.5252	5.86E+09	IPC (36:4)H ⁺ K ⁺	C44H83NO8PNa	703.5750	3.02E+09	ISM (18:1) 16:0)H ⁺ H ⁺	C41H83NO8PNa	769.5623	8.69E+07	ISM (36:1)H ⁺ K ⁺	ISM (36:1)H ⁺ K ⁺
806.5672	4.83E+10	IPC (36:3)H ⁺ Na ⁺	C44H83NO8PNa	822.5411	4.79E+09	IPC (36:3)H ⁺ K ⁺	C44H82NO8PNa	734.5695	3.02E+09	IPC (16:0) 16:0)H ⁺ H ⁺	C42H85NO8PNa	762.6008	4.30E+07	IPC (16:0) 16:0)H ⁺ H ⁺	IPC (16:0) 16:0)H ⁺ H ⁺
848.5562	4.53E+10	IPC (38:4)H ⁺ K ⁺	C46H84NO8PNa	760.5851	4.49E+09	IPC (16:0) 18:1)H ⁺ H ⁺	C42H83NO8PNa								

Table S2. Comparison of lipids present in the different areas of the liver across different species in positive ion mode.

<i>Identity</i>	<i>Mass</i>	<i>ROC value</i>
[TCDCA/TDCA-H] ⁻	498,2896	0.626
[TCA-H] ⁻	514,2844	0.721
[Bilirubin-H] ⁻	583,2561	0.568
[PS (18:0-18:1)-H] ⁻	788,5449	0.684
[PI (18:0-18:2)-H] ⁻	861,5502	0.551
[ST-OH (18:1-24:0)-H] ⁻	906,6352	0.439

Table S4. ROC values for other molecular markers.