Supplemental Tables

Table S1. Examples according to the published reports on CRISPR/Cas9-induce exon-skipping and the case reports in clinical.

Reference	Gene (Gene bank)	Method	Nature of mutation	PTC in Exon	Exon skipping
[1]	*KRAS* (NM_021284.6)	CRISPR/Cas9	-2bp in exon 2	Exon 2	Exon 2
			-1bp in exon 2	No	No
	Ctnnb1 (NM_001098209.1)	CRISPR/Cas9	-3bp in exon 3	No	No
			+1bp in exon 3	Exon 3	Exon 3
			+1bp in exon 3	Exon 3	Exon 3, 4
	P65 (NM_001365067.1)	CRISPR/Cas9	+1bp in exon 6	No	No
[2]	*DMD* (NM_000109.3)	CRISPR/Cas9	-18bp in exon 45	exon 45	exon 45
			+1bp in exon 45	exon 45	exon 45
			-10bp in exon 45	No	No
			-8bp in exon 45	No	No
			-7bp in exon 45	No	No
			-1bp in exon 45	No	No
[3]	*Creb3l3a* (NM_001020673.1)	CRISPR/Cas9	g.357C>T in exon 2	Exon 2	Exon 2
	Pla2g12b (NM_213430.1)	CRISPR/Cas9	g.10194A>T in exon 4	No	No
	Smyd1a (NM_205540.2)	CRISPR/Cas9	-7bp in exon 3	Exon 3	Exon 3
			-13bp in exon 2	Exon 2	Exon 2
			-40bp in exon 2	Exon 2	Exon 2
[4]	*PHACTR1* (NM_001242648.2)	CRISPR/Cas9	+4bp in exon 8	Exon 8	Exon 8
			-1bp in exon 8	Exon 8	Exon 8
			-1bp in exon 9	Exon 9	Exon 9
			+22 bp in exon 10	Exon 10	Exon 10
	Adgrl4 (NM_213367.2)	TALEN	-5bp in exon 2	Exon 2	Exon 2
	LGALS8 (NM_006499.4)	TALEN	p.Leu212Ter in exon 9	Exon 9	Exon 9
[5]	*BRCA1* (NM_007294.3)	Case report	p. Glu1694Ter in exon 18	Exon 18	Exon 18
[6]	*FBN1* (NM_000138.4)	Case report	PTC mutations in exon 51	Exon 51	Exon 51
[7]	*CEP290* (NM_025114.3)	Case report	c.1666del in exon 17	Exon 18	Exon 18
			c.508A>T in exon 8	Exon 8	Exon 8
			c. 4090G>T in exon 32	Exon 32	Exon 32
[8]	*NF1* (NM_000267.3)	Case report.	c.4367+1G>C in exon 32	Exon 32	Exon 32
[9]	*LAMB3* (NM_000228.2)	Case report	c.1903C>T in exon 14	Exon 14	Exon 14
[10]	*LAMB3* (NM_000228.2)	Case report	c.1978C>T in exon 15	Exon 15	Exon 15
[11]	*LAMC2* (NM_005562.2)	Case report	c.1045C>T in exon 8	Exon 8	Exon 8
[12]	*LAMA3* (NM_000227.4)	Case report	c.3928A>T in exon 30	Exon 30	Exon 30
[13]	*LAMA2* (NM_000426.3)	Case report	p. Arg744Stop in exon 15	Exon 15	Exon 15

[14]	*COL11A2* (NM_001163771.1)	Case report	p. Arg893Stop in exon 57	Exon 57	Exon 57
[15]	*CFTR* (NM_000492.3)	Case report	p. Arg553X in exon 11	Exon 11	Exon 11
[16]	*AAAS* (NM_001173466.1)	Case report	c.1331+1G>A in exon 14	Exon 14	Exon 14
[17]	*DOCK9* (NM_001130048.1)	Case report	c. 2262A>C in exon 20	Exon 20	Exon 20
[18]	*CASC5* (NM_144508.4)	Case report	c. 6673-19T>A in exon 25	Exon 25	Exon 25
[19]	*NF1* (NM_000267.3)	Case report	p. Trp336X in exon 7	Exon 7	Exon 7
			p. Gln315X in exon 7	Exon 7	Exon 7
			p. Arg304X in exon 7	Exon 7	Exon 7
[20]	*PS* (NM_000267.3)	Case report	p. Ser62Stop in exon 4	Exon 4	Exon 4
[21]	*ACY1* (NM_000666.2)	Case report	c.1001+5del6 in exon 13	Exon 13	Exon 13
[22]	*CYP27B1* (NM_000785.3)	Case report	c.1022-1037del16: p in exon 7	Exon 7	Exon 7
			c. 934_935delAC in exon 7	Exon 7	Exon 7
			c.1215 T>C in exon 7	No	No
[23]	*FANCM* (NM_001308133.1)	Case report	c.5791C>T in exon 22	Exon 22	Exon 22

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Table S2. PCR primers for genotyping of the gene editing rabbits

Name	Primers	Sequence(5'-3')	Produce size (bp)
DMD	*DMD*-51-F	TAGTTTGGCTCAGATTGTAG	507
	DMD-51-R	AGAATAGACAAAGCAGTGTG	
ANO5	*ANO5*-F	CCCATATGCCTTGTTCTATT	492
	ANO5-R	GCATGATTAGGAACCCTTT	
LMNA	*LMNA*-F	GAAGGGTGGAGAGACAGGAA	490
	LMNA-R	GCTACATCCAATGAGTGAAAGA	
GHR	*GHR*-F	CATTGGTGTCACCTCCTAGATAC	713
	GHR-R	CACACTCACACTCATCCATACA	
DMD	*DMD*-20-F	TCTTTCAGCCTGTGACTTCAG	421
	DMD-20-R	GTGGCTTAGCTAAATCTGTAGGA	
GCK	*GCK*-F	GTGCCCCAGTCCACCATGGAG	626
	GCK-R	CCAACAGCCTCTGCCGGGTTG	
MSTN	*MSTN*-F	GGCCCAGTGGATCTAAATGAA	449
	MSTN-R	AGACTGTCTTTCCTGCTTCTTAC	
TIA1	*TIA1*-F	GGCATTACTGTTACGTTGGTATTT	459
	TIA1-R	GGCAGACATCCAGCATCTT	
TYR	*TYR*-F	ATCCGCTCAAGCAGGTATTG	454
	TYR-R	AGTGAGGTAGGCAAGGAATTTG	
OXT	*OXT-F*	GCAAGGTGAGTGTTCACAGG	609
	OXT-R	TATTATTCCTGGGAGTGGCTGA	

DMD-51, the primer used for Fig.S1. *DMD*-20, the primer used for Fig.S5. *OXT*, the primer used for Fig.S10 .

Table S3. Primers for RT-PCR analysis of exon skipping.

Name	Primers	Sequence(5'-3')	Produce size (bp)
DMD	*DMD*-51-F	GTCAACTATCTACTGCAAGAGC	459
	DMD-51-R	CTGTACTTCATCCCACTGATTC	
ANO5	*ANO5*-F	CGACAAGCCACATTGGAATAC	610
	ANO5-R	ACTGAGTCGTCAGTTCGATTAG	
LMNA	*LMNA*-F	CATCAAGGCCGCCTACGAG	418
	LMNA-R	CAGCTCCTCGCTGTAGATGTT	
GHR	*GHR*-F	CACTAGCAGGGTCAAGTGATG	478
	GHR-R	TCTGCATGAATCCCGGTTAAG	
DMD	*DMD*-20-F	TGGTGGAACAGATGGTGAATG	492
	DMD-20-R	TGGTACTGATCGTCTCCTGATAG	
GCK	*GCK*-F	GAGGACCTGAAGAAGGTGATG	466
	GCK-R	CCACGATGTTGTTCCCTTCA	
MSTN	*MSTN*-F	TTATCACGCTACGACGGAAAC	494
	MSTN-R	GCTCATCACAGTCAAGACCAA	
TYR	*TYR*-F	ACTACGAGCCCAGACTATGT	432
	TYR-R	GCTCTGTCGGCTATTGTACTC	
TIA1	*TIA1*-F	GTTCGGTTCAACTCCCATGA	291
	TIA1-R	GGAAGACTGCGTCTGGTTAAA	
OXT	*OXT*-F	GCTGACCTCGGCCTGCTACAT	331
	OXT-R	GCGCTCGGAGAAGGCGGCCTCG	

DMD-51, the primer used for Fig.S1. *DMD*-20, the primer used for Fig.S5. *OXT*, the primer used for Fig.S10.

Table S4. Primers for qPCR analysis.

Name	Primers	Sequence(5'-3')	Produce size (bp)
MSTN	*MSTN*-F1	GCTCTTTGGAAGATGACGATTA	109
	MSTN-R1	AAGAAGCAACATTTGGGTTT	
	MSTN-F2	CCTGAGGCTCATCAAACCTATG	106
	MSTN-R2	CACATCAATGCTCTGCCAAATAC	
TYR	*TYR*-F1	GCTGTGATGTTTGTACAGATGAG	113
	TYR-R1	CTCCAACCGGCTACAGATAAT	
	TYR-F2	CCATGCGTTTGTTGACAGTATT	111
	TYR-F2	ATGTAGGATTCCCGGTTGTG	
TIA1	*TIA1*-F1	CACAACAGATTGGCCAGTACAT	116
	TIA1-R1	CATCCACGGTGCGGAAGAC	
	TIA1-F2	CGAAGGGATATGGCTTTGTCT	112
	TIA1-F2	GTTGCCCAGTTAGTTCTGATTTG	