

Figure S1. Related to Figure 1. CO₂ response in *C. elegans* is experience-dependent. (A) Schematic of the chemotaxis assay. Animals were placed in the center of a 100-mm agar plate (84.4-mm inside base diameter). A CO₂ gradient was established by delivering gas stimuli through holes on either side of the plate. At the end of the assay, the number of animals in a 20-mm diameter circle centered under each hole was counted and used to determine the chemotaxis index (CI) according to the formula shown above. (B-C) Animals raised at ambient CO₂ avoid CO₂ across concentrations (B), while animals raised at 2.5% CO₂ are attracted to CO₂ across concentrations (C). ***p*<0.01, ****p*<0.001, Kruskal-Wallis test with Dunn's post-test. For each graph, significance was determined relative to the 0% CO₂ condition. *n*=8-20 trials per condition. (D) Animals raised at either ambient or high (2.5%) CO₂ were tested in 0-2.5% (left), 2.5-10% (middle), or 2.5-40% (right) CO₂ gradients. Animals grown at ambient CO₂ migrate toward the lower CO₂ concentration. Animals grown at high CO₂ are attracted to the higher CO₂ concentration when tested in a 0-2.5% or 2.5-10% CO₂ gradient, but not when tested in a 2.5-40% CO₂ gradient. *n*=8-16 trials per condition. (E) BAG activity regulates behavioral sensitivity to CO₂. In animals raised at ambient CO₂, genetic ablation of the BAG neurons (BAG^{-/-}) eliminates CO₂ avoidance across concentrations. By contrast, animals with more active BAG neurons due to BAG-specific expression of *pkc-1(gf)* show enhanced CO₂ avoidance. **p*<0.05, ****p*<0.001, two-way ANOVA with Dunnett's post-test. *n*=8-16 trials per condition. For B-E, graphs depict medians with interquartile ranges. (F) The BAG response to CO₂ is concentration-dependent. Graph shows the calcium responses of BAG neurons to 15% CO₂ (orange) or 5% CO₂ (red), for animals raised at high (2.5%) CO₂, measured using the ratiometric calcium indicator yellow cameleon YC3.60. Solid lines indicate average calcium responses; shading represents SEM. Black line indicates the CO₂ pulse.

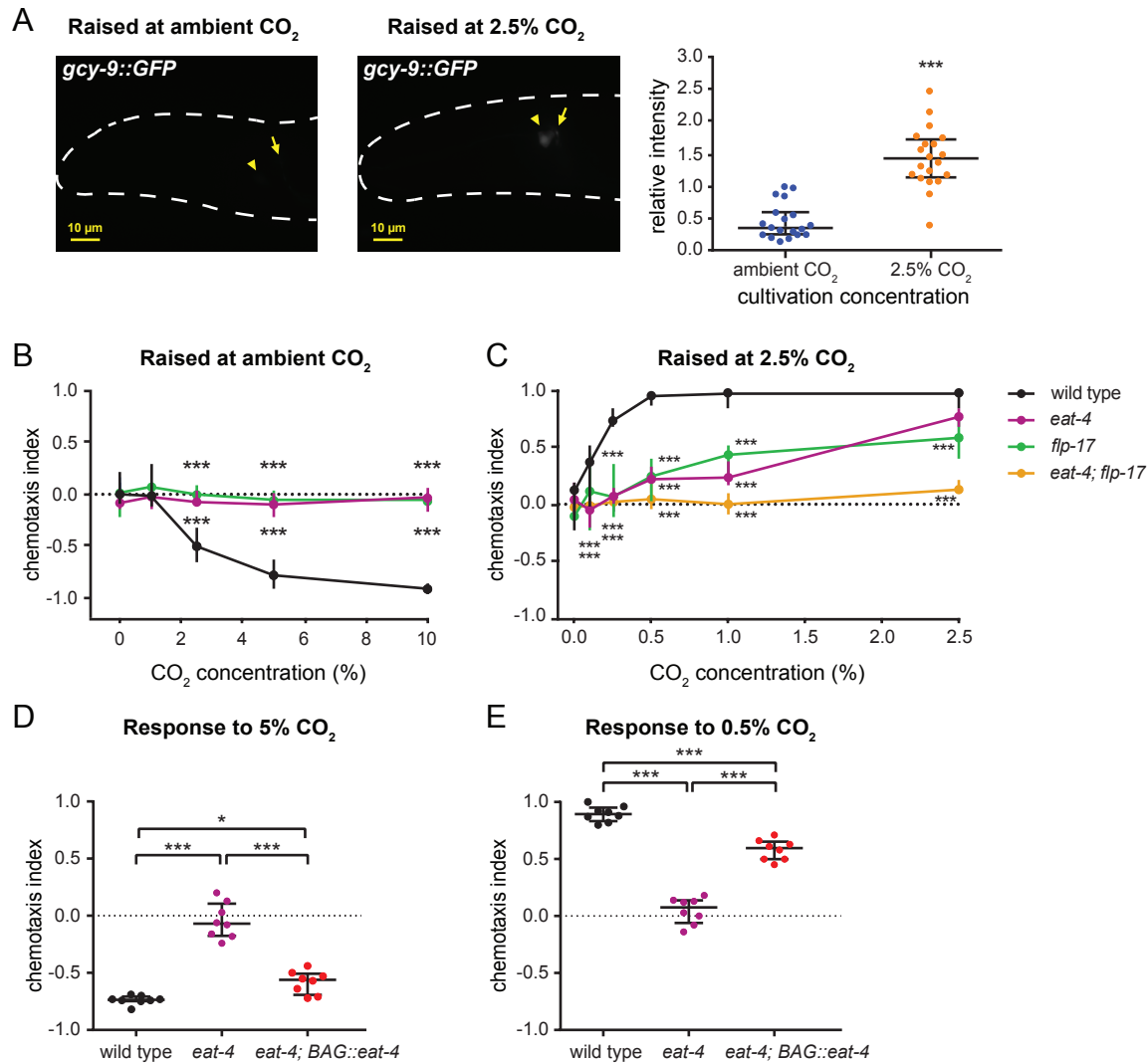


Figure S2. Related to Figure 1. BAG neurons mediate both attractive and aversive CO₂ responses. (A) Epifluorescence images of *gcy-9* expression in the BAG neurons of L4 animals raised at ambient CO₂ (left) or high (2.5%) CO₂ (middle). *gcy-9* expression was measured in animals containing a *gcy-9::GFP* transgene. Arrowheads indicate the location of the BAG neuron cell body; arrows indicate the location of the nerve ring. Anterior is to the left; dorsal is up. GFP expression is faint but detectable in animals raised at ambient CO₂, and brighter in animals raised at high CO₂. Graph (right) shows the relative intensity of expression of the *gcy-9::GFP* transgene in animals raised at ambient vs. high CO₂. ****p*<0.001, unpaired t test. *n*=19-20 animals per condition. (B-C) *eat-4* and *flp-17* are required for normal CO₂ response. (B) Mutation of *eat-4* or *flp-17* abolishes CO₂ avoidance in animals raised at ambient CO₂. ****p*<0.001, two-way ANOVA with Dunnett's post-test. *n*=6-16 trials per genotype and condition. (C) Mutation of either *eat-4* or *flp-17* reduces CO₂ attraction, and mutation of both genes abolishes CO₂ attraction, in animals raised at high (2.5%) CO₂. ****p*<0.001, two-way ANOVA with Dunnett's post-test. *n*=8-26 trials per genotype and condition. (D-E) *eat-4* acts in the BAG neurons to mediate CO₂ avoidance and CO₂ attraction. Restoring *eat-4* expression specifically in the BAG neurons of *eat-4* mutants partially restores CO₂ avoidance (D) and attraction (E). Animals were raised at ambient CO₂ (D) or 2.5% CO₂ (E) and tested for their response to 5% CO₂ or 0.5% CO₂, respectively. **p*<0.05, ****p*<0.001, one-way ANOVA with Sidak's post-test. *n*=8 trials per genotype and condition. For A-E, graphs show medians with interquartile ranges.

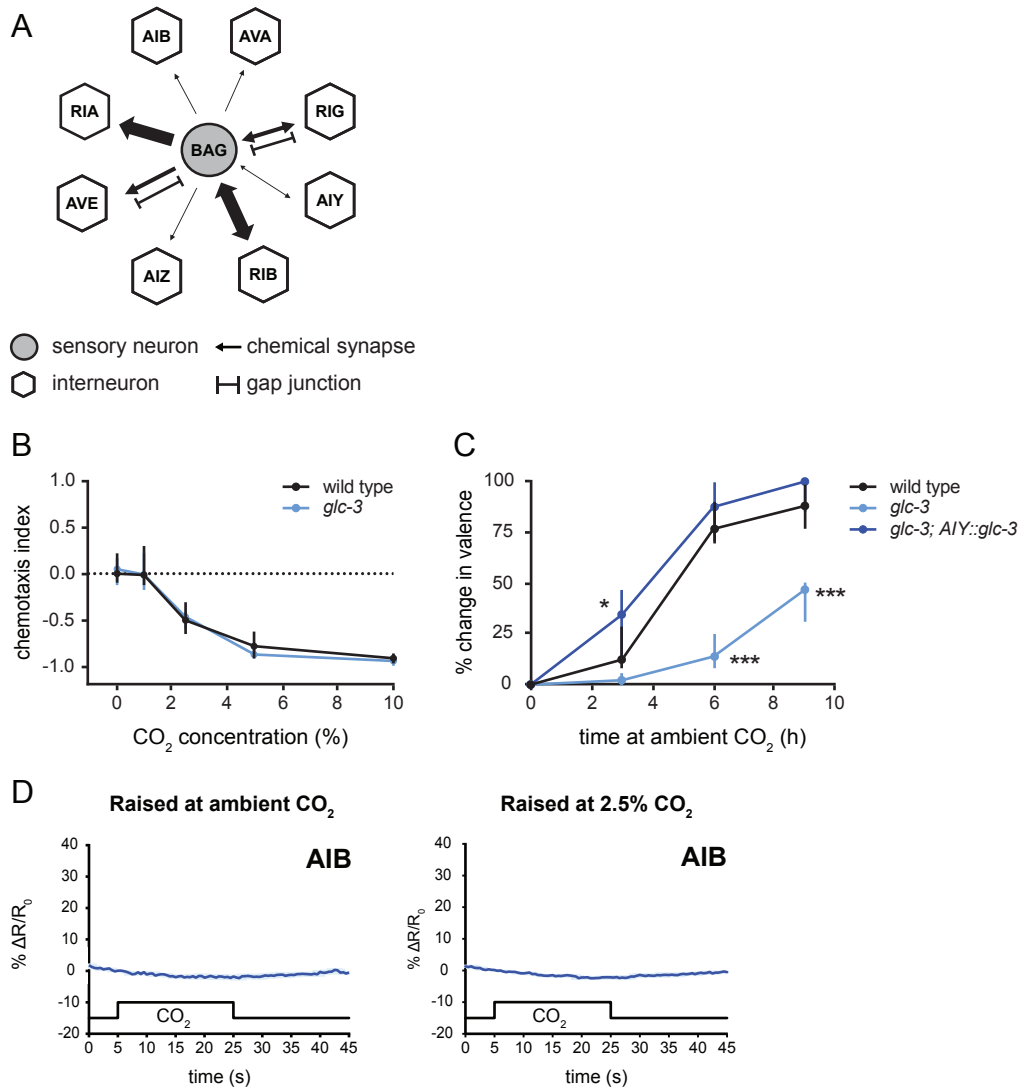


Figure S3. Related to Figure 2, Figure 3 and Figure 4. Distinct interneurons regulate CO₂ avoidance and attraction. (A) Structural connectivity of the CO₂-detecting BAG neurons. Arrow thickness reflects the number of synaptic connections (1-2 synapses, 4-7 synapses, or 10+ synapses) from BAG to the downstream interneurons [S1, S2]. The interneurons shown have documented functions in other chemosensory microcircuits and were therefore investigated in this study. The BAG sensory neurons are also presynaptic to several other interneurons not shown. (B) *glc-3* mutants show normal CO₂ avoidance when raised at ambient CO₂. (C) *glc-3* mutants show a delayed shift from CO₂ attraction to avoidance when raised at high (2.5%) CO₂ and transferred to ambient CO₂. Restoring *glc-3* function specifically to AIY (*AIY::glc-3*) rescues the shift in CO₂ response valence. Animals were tested for their response to 2.5% CO₂. Graph shows the percent change in valence as a function of time (see Methods). **p*<0.05, ****p*<0.001, two-way ANOVA with Sidak's post-test (B) or two-way ANOVA with Dunnett's post-test (C). n=8-24 trials per genotype and condition. For B-C, graphs show medians with interquartile ranges. (D) AIB is not activated by CO₂ in animals raised at ambient CO₂ or high (2.5%) CO₂. n=8 animals per genotype and condition. Calcium responses were measured using the ratiometric calcium indicators yellow cameleon YC3.60. Graphs show composite calcium responses to a 20-s pulse of 15% CO₂. Solid lines indicate average calcium responses; shading represents SEM.

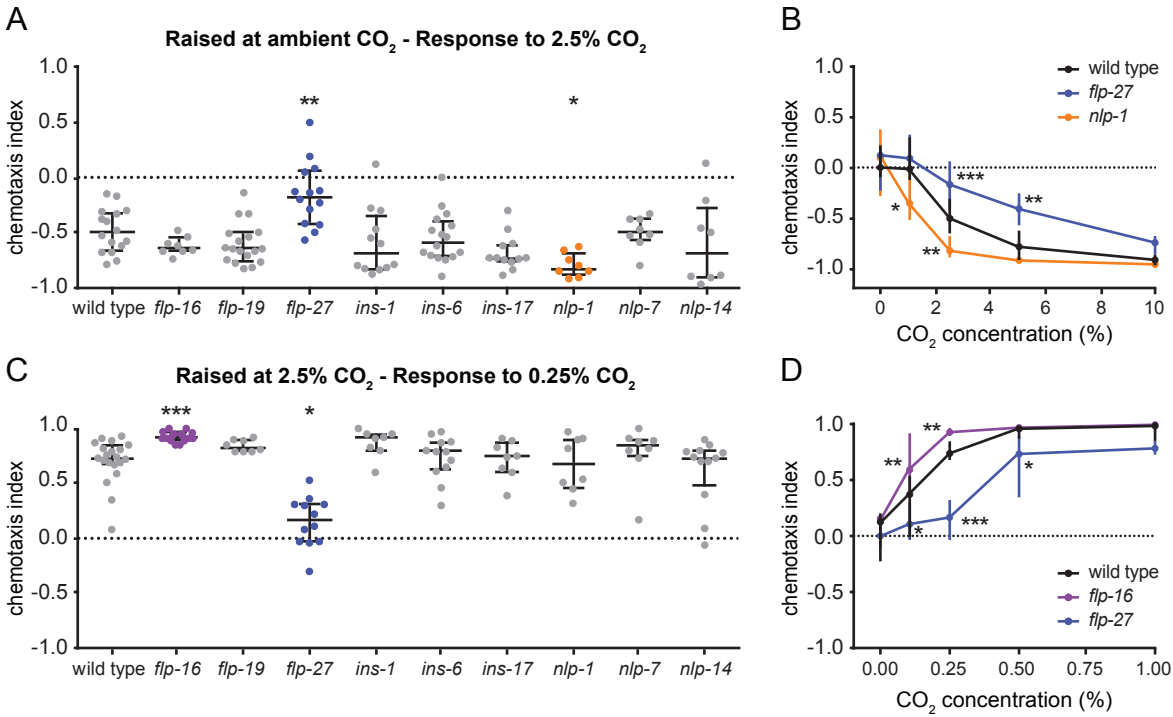


Figure S4. Related to Figure 4. A combinatorial code of neuropeptides regulates CO₂ response valence and sensitivity. (A-B) In animals raised at ambient CO₂, *flp-27* mutants show reduced CO₂ avoidance and *nlp-1* mutants show enhanced CO₂ avoidance. Graphs show responses to 2.5% CO₂ (A) or across CO₂ concentrations (B). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, one-way ANOVA with Dunnett's post-test (A) or two-way ANOVA with Dunnett's post-test (B). $n = 8-18$ trials per genotype and condition. (C-D) In animals raised at high (2.5%) CO₂, *flp-16* mutants show enhanced CO₂ attraction and *flp-27* mutants show reduced attraction. Graphs show responses to 0.25% CO₂ (C) or across CO₂ concentrations (D). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, Kruskal-Wallis test with Dunn's post-test (C) or two-way ANOVA with Dunnett's post-test (D). $n = 6-20$ trials per genotype and condition. For A-D, graphs show medians and interquartile ranges.

Supplemental References

- S1. White, J.G., Southgate, E., Thomson, J.N., and Brenner, S. (1986). The structure of the nervous system of the nematode *Caenorhabditis elegans*. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 314, 1-340.
- S2. Xu, M., Jarrell, T.A., Wang, Y., Cook, S.J., Hall, D.H., and Emmons, S.W. (2013). Computer assisted assembly of connectomes from electron micrographs: application to *Caenorhabditis elegans*. *PLoS One* 8, 1-6.