An autoimmune disease risk SNP, rs2281808, in SIRPG is associated with reduced expression of SIRPγ and heightened effector state in human CD8 T-cells

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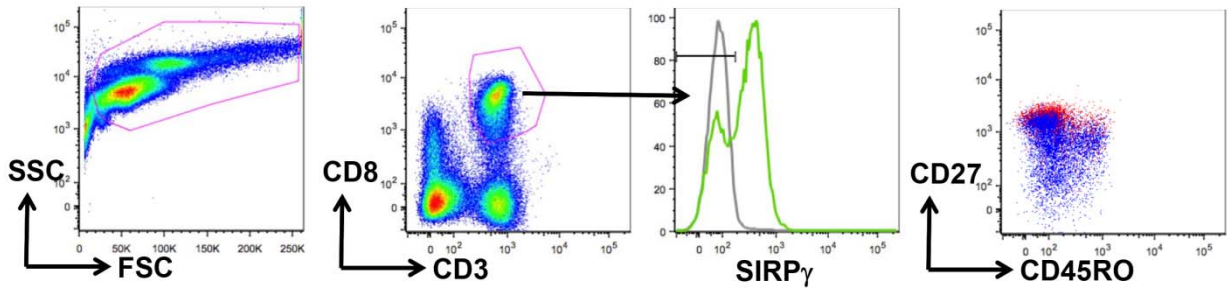
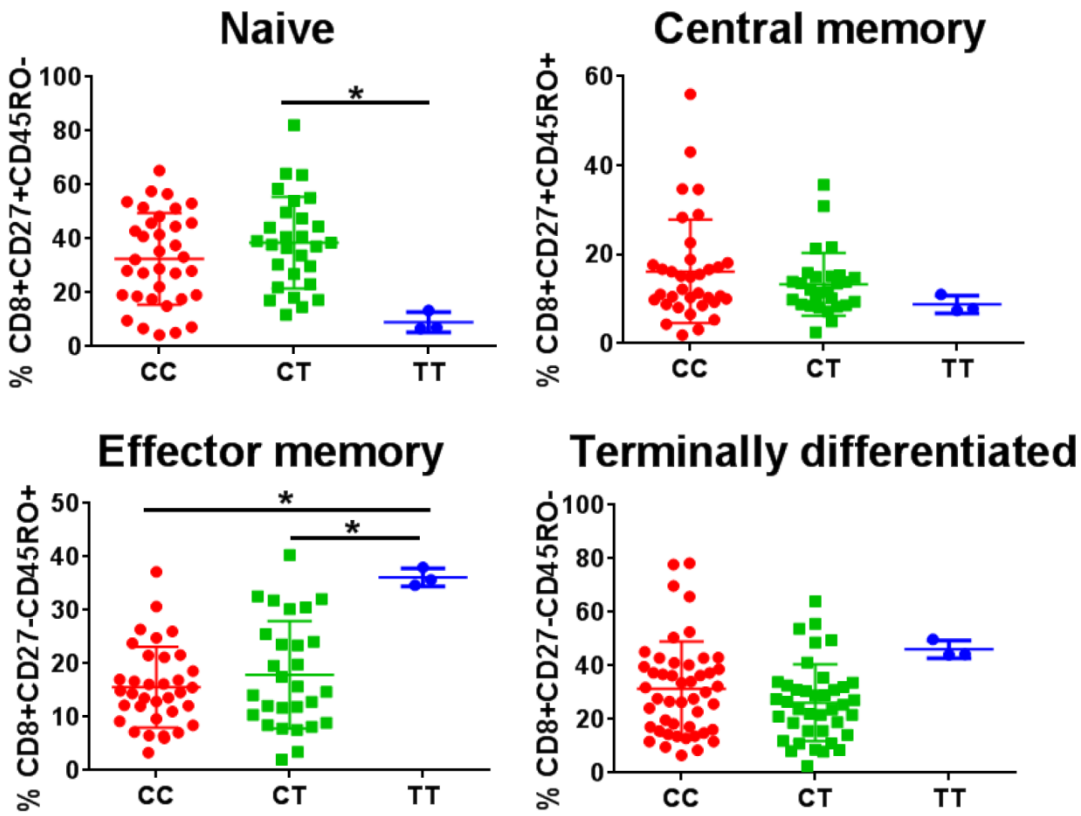
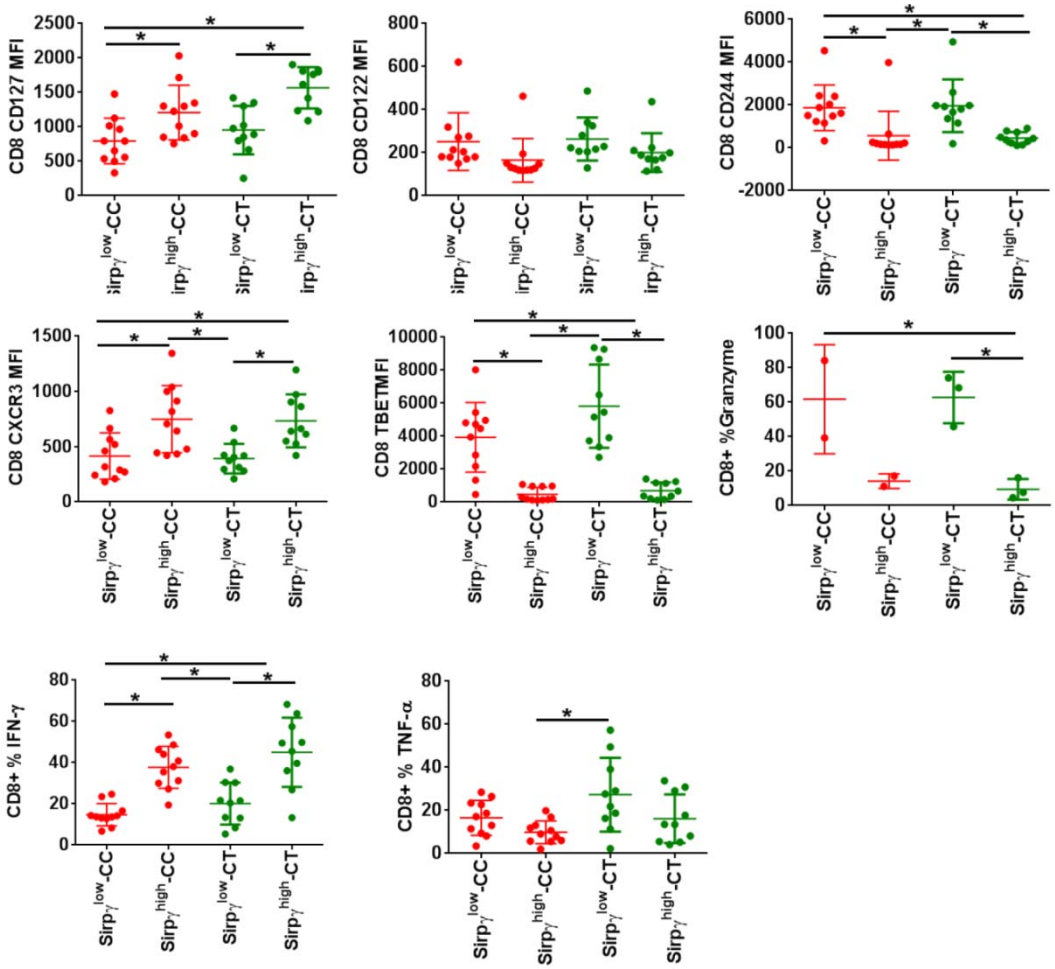


Supplementary Fig 1. Gating strategy for Fig 2. Live gated lymphocytes were gated on CD3+ CD8 T cells. SIRPγ expression on CD8 T cells was analyzed and SIRPγ low CD8 T-cells were gated. SIRPγ low CD8 T-cells were then overlaid over total CD8 T-cells displayed in the dot plot showing CD27 and CD45RO.



Supplementary Fig 2. CD8 T cell subsets with respect to rs2281808 genotype. PBMC samples were genotyped for rs2281808 by TaqMan PCR. PBMC samples were also subjected to flow cytometry staining. Gated CD3+CD8+ T-cells were analyzed for naïve (CD27+CD45RO-), central memory (CD27+CD45RO+), effector memory (CD27-CD45RO+), and terminally differentiated markers (CD27-CD45RO-) and plotted with respect to their rs2281808 genotype status. One-way ANOVA with Tukey's posthoc was performed and $p<0.05$ was considered significant.



Supplementary Fig 3. Phenotypic profile of SIRPγ-high vs. low CD8 T-cells from CC and CT carriers are displayed separately. CD8 T-cells from CC, CT and TT carriers were evaluated for their expression of the indicated molecules by flow cytometry. Intracellular staining was performed for T-bet. In case of CC and CT, cells were gated for SIRPγ-high vs. low cells. PBMC samples from CC and CT carriers were stimulated with PMA/Ionomycin/brefeldin A for 5 hr before intracellular staining for IFN-γ and TNF-α. CD8 T-cells from CC and CT carriers were gated on SIRPγ^{low} vs high cells for and evaluated for frequency of cells producing the indicted

cytokines. 1-way ANOVA with Tukey's posthoc analysis was performed and $p<0.05$ was considered significant.