

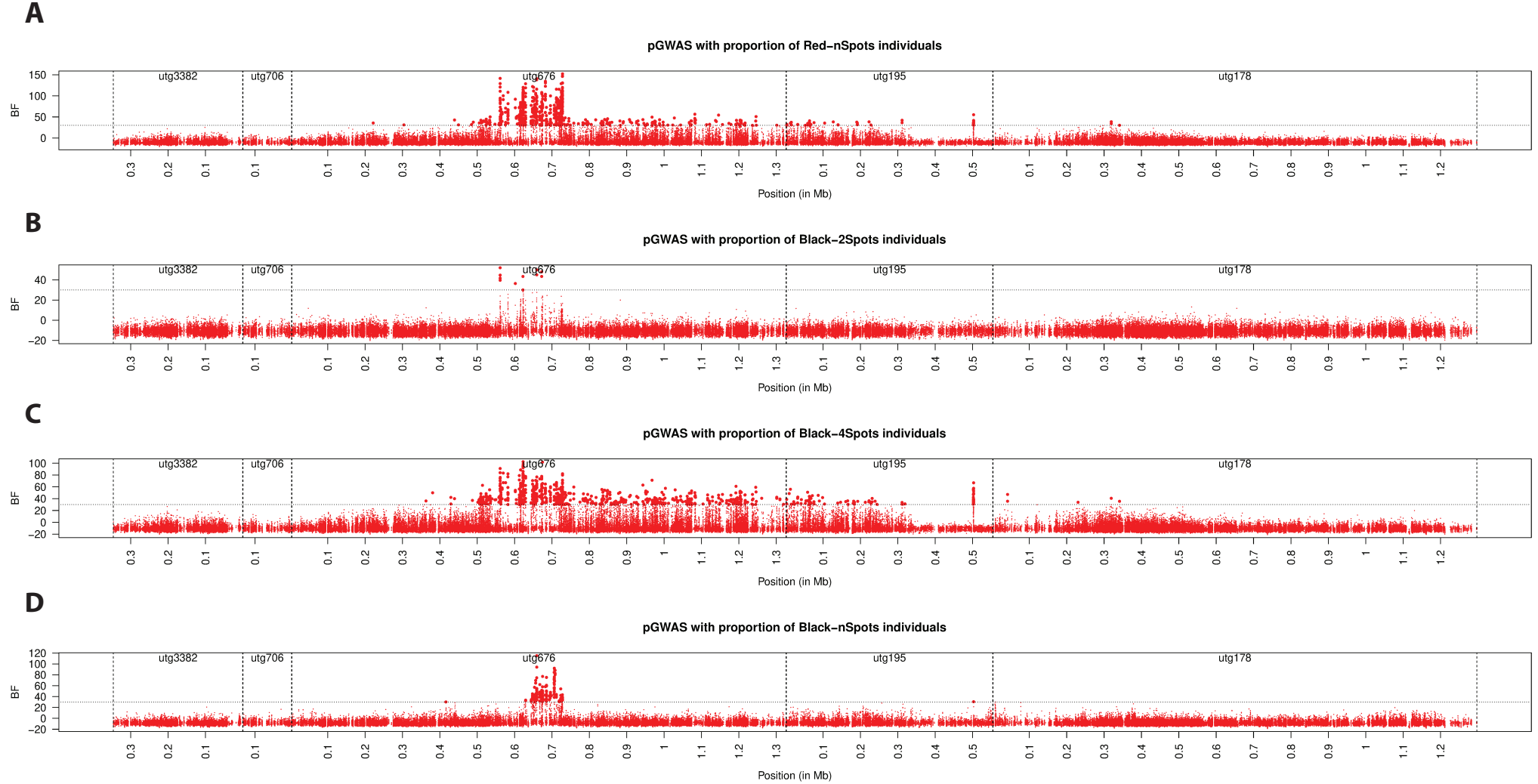
**Current Biology, Volume 28**

## **Supplemental Information**

### **The Genomic Basis of Color Pattern**

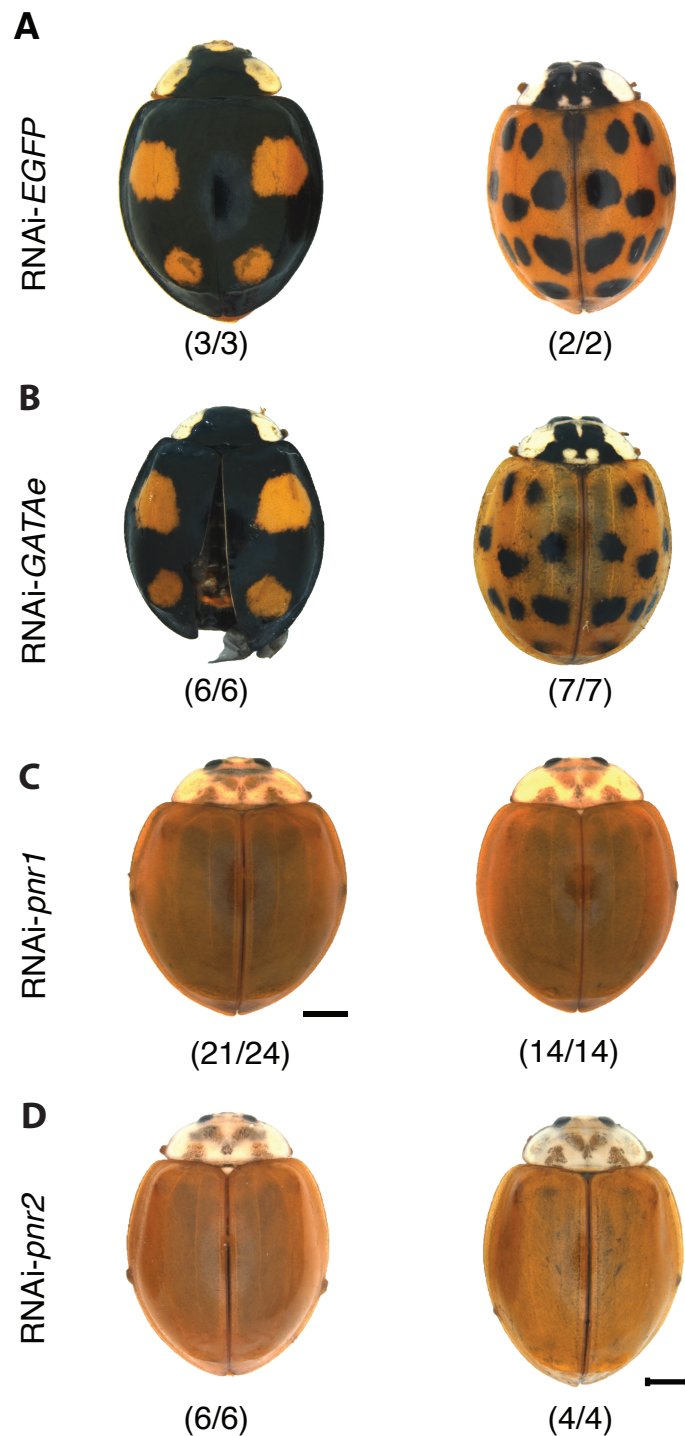
#### **Polymorphism in the Harlequin Ladybird**

**Mathieu Gautier, Junichi Yamaguchi, Julien Foucaud, Anne Loiseau, Aurélien Ausset, Benoit Facon, Bernhard Gschloessl, Jacques Lagnel, Etienne Loire, Hugues Parrinello, Dany Severac, Celine Lopez-Roques, Cecile Donnadiou, Maxime Manno, Helene Berges, Karim Gharbi, Lori Lawson-Handley, Lian-Sheng Zang, Heiko Vogel, Arnaud Estoup, and Benjamin Prud'homme**



**Figure S1. Genome-wide association study focusing on the color pattern genomic region of the *HaxR* assembly. Related to Figure 1.**  
 (A) Association with the proportion of Red-nSpots individuals. (B) Association with the proportion of Black-2Spots individuals.  
 (C) Association with the proportion of Black-4Spots individuals. (D) Association with the proportion of Black-nSpots individuals.

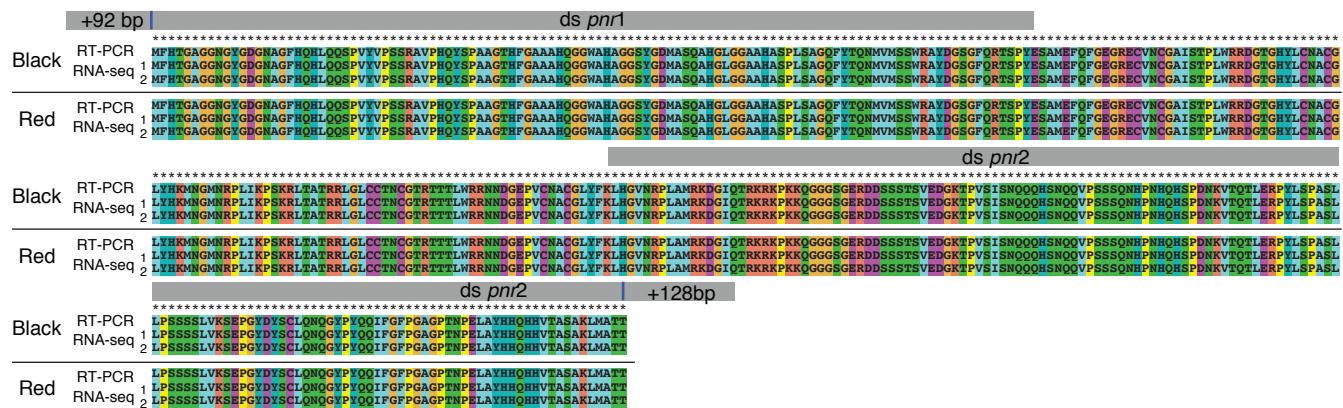




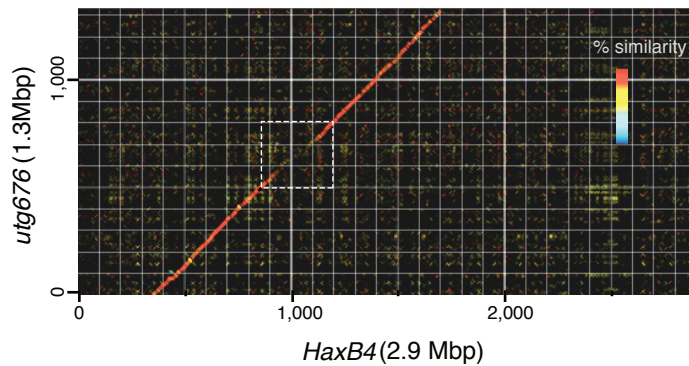
**Figure S2. *pannier*, but not *GATAe*, is necessary for adult pigmentation patterns. Related to Figure 2; Figure S3.**

(A-D) RNAi phenotypes after larval injection of dsRNA in Black-4Spots (left column) or Red-nSpots (right column). (A) dsRNA targeting *eGFP* (negative control). (B) dsRNA targeting *GATAe*. (C) dsRNA targeting *pnr\_1* region. (D) dsRNA targeting *pnr\_2* region. *pnr\_1* and *pnr\_2* are non-overlapping regions of the *pannier* cDNA. Numbers in parentheses indicate the proportion of eclosed adults showing the pigmentation pattern of the representative individual. Scale bar; 1mm.

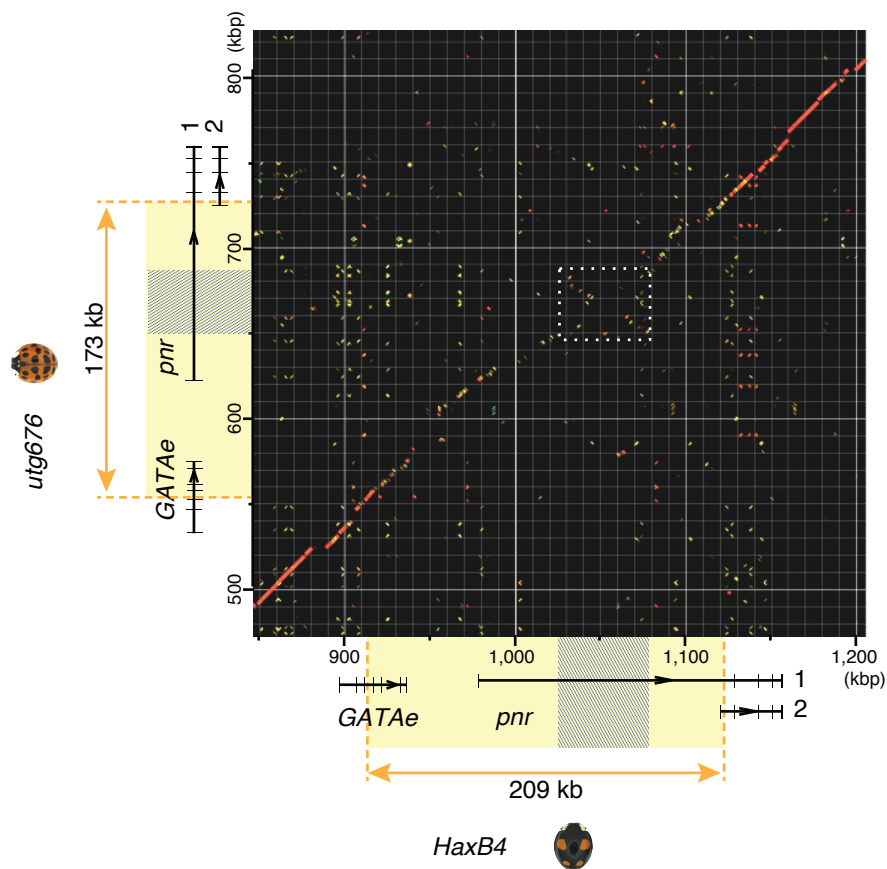
**A**



**B**

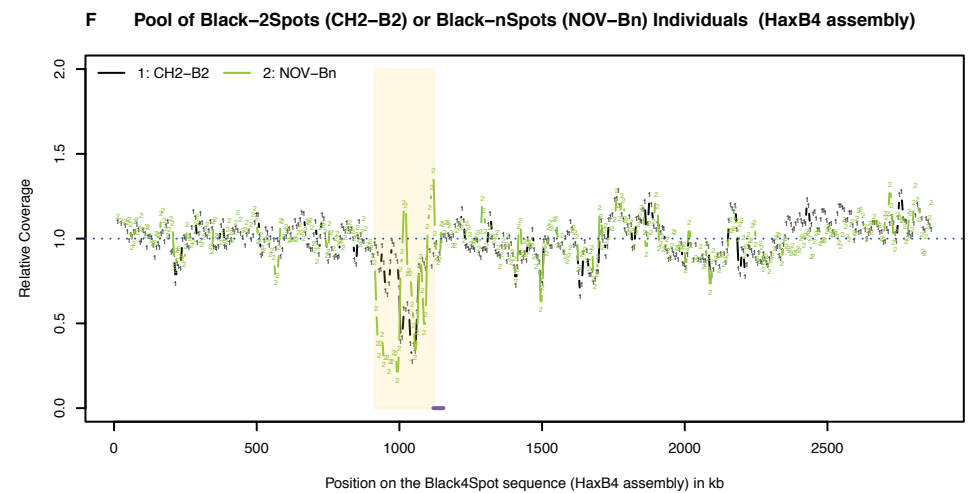
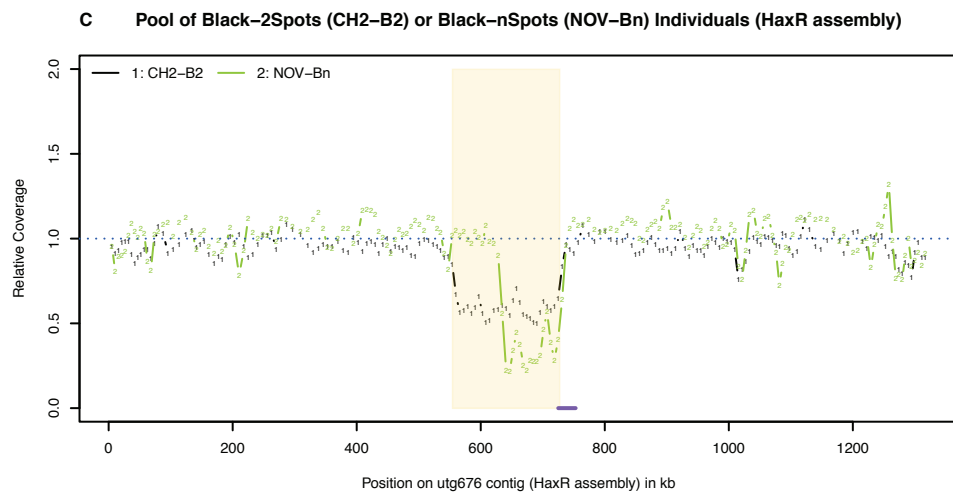
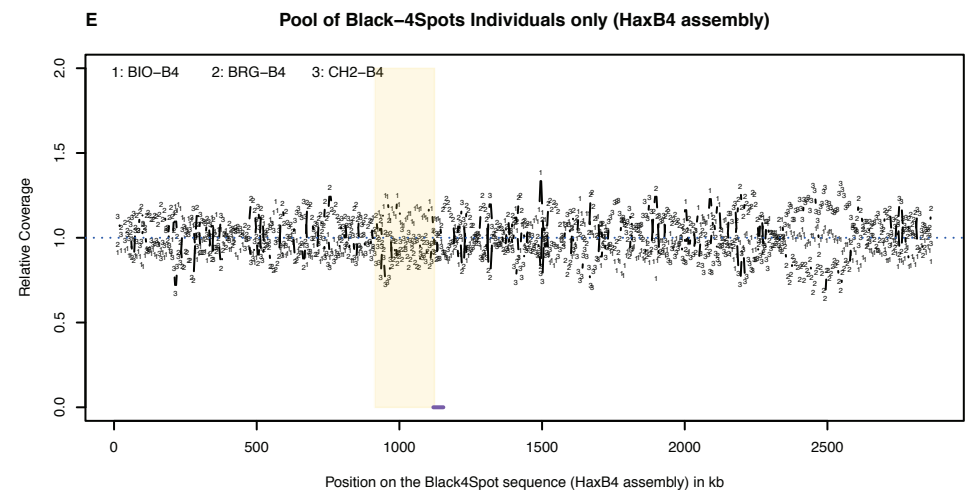
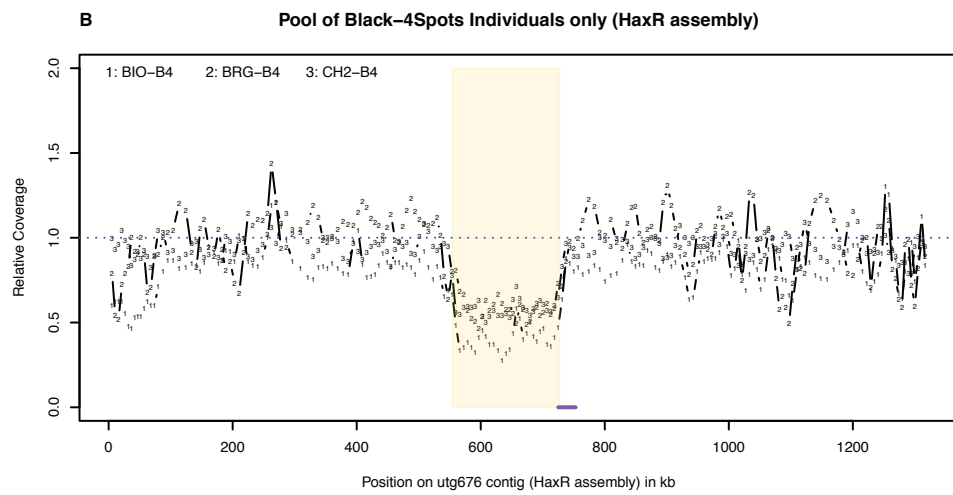
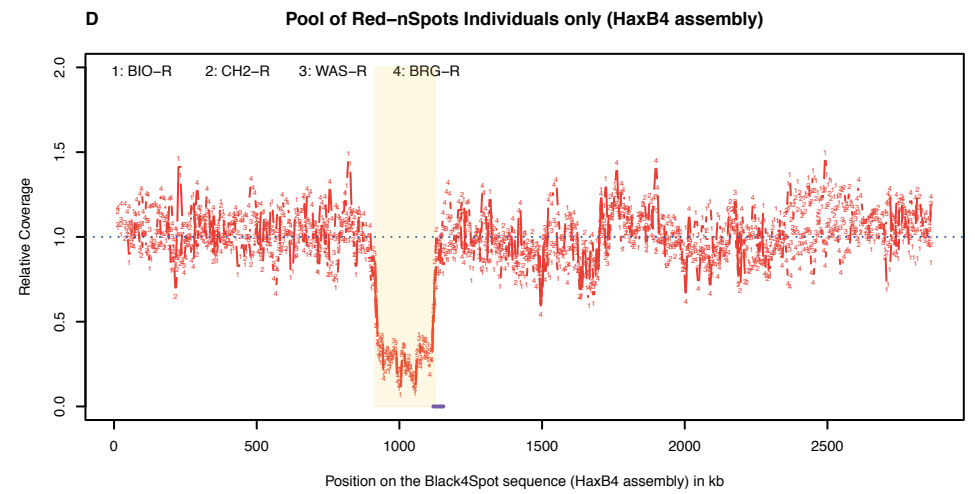
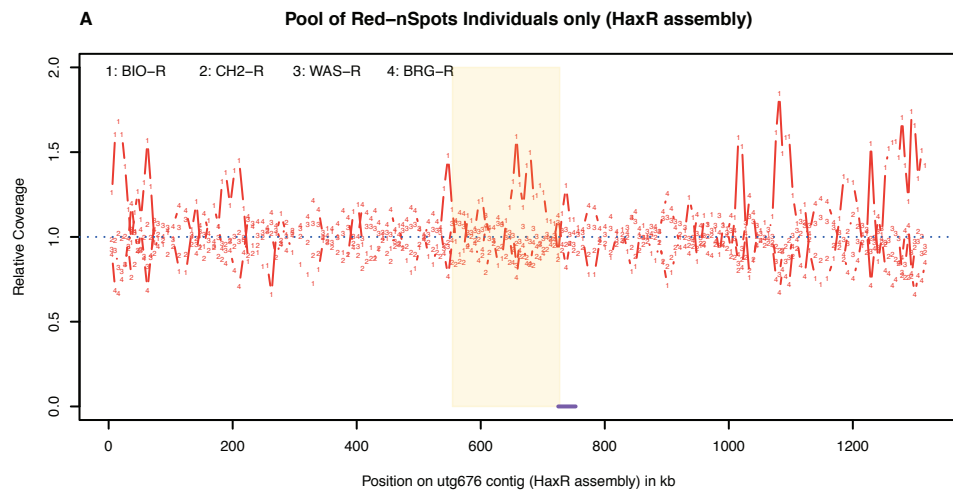


**C**



**Figure S3. Comparison of Pannier sequences and *utg676* and *HaxB4* sequences. Related to Figure 1; Figure 2; Figure S2.**

(A) Alignment of Pannier sequences, resulting from conceptual translation of cDNA obtained by RT-PCR or by mapping two independent RNA-seq reads from Black-4Spots or Red-nSpots individuals. Asterisks indicate conserved amino acids. The positions of the two fragments amplified for dsRNAs (*ds\_pnr1* and *ds\_pnr2*) are shown in grey. (B) Dot-plot between genomic scaffolds from the Black-4Spots (assembly *HaxB4*) and Red-nSpots (assembly *HaxR*, contig *utg676*) forms. The most divergent sequence (white dashed box in B) is shown at a higher magnification in C). (C) Traces of a sequence inversion are visible in the divergent region (white dashed box).



**Figure S4. Relative read mapping coverage for window of 10,000 positions. Related to STAR Methods.**

(A-C) relative read mapping coverage over the extended Red-nSpots (*HaxR* assembly), and (D-F) relative read mapping coverage over the Black-4Spots allele sequence (*HaxB4* assembly) for i) the four pools including Red-nSpots individuals only (A, D); ii) the three pools including Black-4Spots individuals only (B, E); and iii) the CH2-B2 and NOV-Bn pools including respectively Black-2Spots individuals only and Black-nSpots individuals only (C, F). The horizontal blue dotted line gives the value of 1 expected for the absence of departure in the relative coverage. The genomic region where the sequences of the Red-nSpots and Black-4Spots alleles diverge is represented as a shaded orange area. A purple line represents the position of the gene *pannier*. As expected, no obvious departure of the relative window coverage (from the expected value of 1) was observed over the entire Red-nSpots *utg676* contig for the four pools including Red-nSpots individuals only, although the BIO-R pools displayed several regions with high relative coverage values (A). In contrast, for the three pools including Black-4Spots individuals only, we found that the relative window coverage was almost halved in the ca. 170 kb region upstream *pannier*, with slight pattern differences likely explained by the pool frequency of heterozygous individuals carrying a Red-nSpots allele (B). When considering the extended Black-4Spots allele sequence as a reference (assembly *HaxB4*), the pattern was reverted: no obvious departure of the relative window coverage was observed for Black-4Spots pools (E) and the coverage was almost halved for Red-nSpots pools in the genomic region harbouring strong sequence divergence between the two color pattern form alleles (D). For the CH2-B2 pool including Black-2Spots individuals only, we observed a clear reduction in relative coverage over the entire genomic region where the sequences of the Red-nSpots and Black-4Spots alleles diverge on the *HaxR* assembly (C). Interestingly, the reduction in relative coverage concerned a more restricted region (located closer to *pannier*) on the *HaxB4* assembly (F). This suggests that the Black-2Spots allele might be more similar to the Black-4Spots allele, over at least a part of the upstream region of *pannier*, although it should be kept in mind that additional large scale variation (insertion or deletion) specific to the Black-2Spots allele are not identifiable with this approach. Finally, for the NOV-Bn pool including Black-nSpots individuals only, we observed a clear reduction in relative coverage over the second half of the region where the sequences of the Red-nSpots and Black-4Spots alleles diverge on the *HaxR* assembly (C). This pattern suggests a close similarity in sequence of the Black-nSpots allele with the Red-nSpots allele over the first half of its sequence. On the *HaxB4* assembly we also observed a clear reduction of the NOV-Bn relative coverage over the first half of the region where the sequences of the Red-nSpots and Black-4Spots alleles diverge, whereas coverage reduction was less obvious in the second half of the region (F). This suggests some sequence similarity between the Black-nSpots and the Black-4Spots allele in the latter genomic region. It is worth noting, however, that for the Black-nSpots allele too, additional specific large-scale variations (insertion or deletion) are not identifiable with this approach.

	Assembly <i>HaxR</i>	Assembly <i>HaxB4</i>
Data	MinION long reads (65X) Illumina PE reads (100X)	Illumina PE reads (65X) Illumina MP reads (24X)
Assembler	SMARTdenovo	ALLPATH-LG
Nb. of sequences	1,071 contigs	6,586 scaffolds
Total length (Mbp)	429	393.1
Average length (Kbp)	400.9	59.7
Max size (Kbp)	7,499	5,635
Total Ns (bp)	22	22,814,986
N50 (Kbp)	1,434	978.4
BUSCO (complete)	97.2 %	86.0 %
BUSCO (fragmented)	1.3 %	8.7 %
BUSCO (missing)	1.5 %	5.3 %

**Table S1 Statistics characterizing the *HaxR* assembly obtained from Red-nSpots individuals and the *HaxB4* assembly obtained from Black-4Spots individuals. Related to STAR Methods.**  
More details can be found in DataS1.

Pooled-sequencing sample code	Population sampling site		Sampling year	Colour form in the pool	No. of sequenced individuals
	Country	Region or city			
CH1-R	China	Jilin	2013	Red-nSpots only	100
CH1-B				30 Black-4Spots 28 <b>Black-2Spots</b>	58
CH2-R	China	Changchun	2015	Red-nSpots only	100
CH2-B4				Black-4Spots only	67
CH2-B2				<b>Black-2Spots only</b>	73
JP-R	Japan	Kyoto and other cities	2009	Red-nSpots only	57
JP-B4				Black-4Spots only	58
NOV-Bn	Russia	Novosibirsk	2007	Black-nSpots only	44
BRG-R	France	Bourgogne	2013	Red-nSpots only	50
BRG-B4				Black-4Spots only	50
ENA-R	USA	Georgia	2007	Red-nSpots only	45
WAS-R	USA	Washington	2007	Red-nSpots only	40
BIO-R	France	Biological control population (BIOTOP)	2012	Red-nSpots only	100
BIO-B4				Black-4Spots only	100

**Table S2 Sequenced pools of individuals representative of the world-wide diversity and the four main color pattern forms of *H. axyridis*. Related to STAR Methods.**

The colour pattern forms **Red-nSpot**, Black-nSpots, Black-4Spots and **Black-2Spots** correspond to the forms *f. succinea*, *f. axyridis*, *f. conspicua*, and *f. spectabilis*, respectively. Because of the hierarchical dominance between color form alleles (i.e. Black-2Spots > Black-4Spots > Black-nSpots > Red-nspot), a population pool sample including individuals of a single colour pattern form is characterized by a high proportion of the allele associated to that form but also contains alleles associated to other more recessive forms if present in the population. Only population pool samples with Red-nSpots individuals contain 100% of Red-nSpots alleles.