

Supplemental Material

Table S1. Genotypes and Sources of *Saccharomyces cerevisiae* Strains

Strain	Genotype	mtDNA	Source
W303-1A	MAT_a <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^+	a
W303-1B	MAT_{α} <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^+	a
MR6	MAT_a <i>ade2-1 arg8::HIS3 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^+ (S288C)	Rak <i>et al.</i> 2007
MRS-3A	MAT_a <i>ade2-1 arg8::HIS3 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^+ (S288C)	(McStay <i>et al.</i> , 2013)
MRS-3B	MAT_{α} <i>ade2-1 arg8::HIS3 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^+ (S288C)	(McStay <i>et al.</i> , 2013)
MRS-3B ρ^0	MAT_{α} <i>ade2-1 arg8::HIS3 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^0	(McStay <i>et al.</i> , 2013)
aDFK ρ^0	MAT_a <i>kar1-1 ade2-10 arg8::URA3 leu2Δ ura3-52 lys2</i>	ρ^0	(McStay <i>et al.</i> , 2013)
DFK ρ^0	MAT_{α} <i>kar1-1 ade2-10 arg8::URA3 leu2Δ ura3-52 lys2</i>	ρ^0	(McStay <i>et al.</i> , 2013)
SCS9S	MAT_{α} <i>kar1ade2-101 arg8::URA3 leu2Δ ura3-52 lys2</i>	ρ^+ COX2-HAC	b
HMD22	MAT_a <i>leu2-3,112 ura3-52 lys2 his3Δ arg8::hisG</i>	ρ^+ cox2::ARG8 ^m	b
MRS Δ COX2	MAT_{α} <i>ade2-1 arg8::HIS3 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	ρ^+ cox2::ARG8 ^m	HMD22 x MRS-3B ρ^0

MRSI ⁰ /COX1-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1arg8::HIS3</i>	Intronless mtDNA with COX3-HAC	(McStay <i>et al.</i> , 2013)
MRS/COX2-pH	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1arg8::HIS3</i>	COX2-pH	This study
MRS/COX2-HA	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1arg8::HIS3</i>	COX2-HA	This study
MRS/COX2-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1arg8::HIS3</i>	COX2-HAC	This study
W303/COX2-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	COX2-HAC	This study
W303 Δ COX1/COX2-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1</i>	COX2-HAC, Δ <i>cox1::ARG8^m</i>	This study
W303/COX9-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox12::HIS3 leu2::pG16/ST14</i>	ρ^+	(McStay <i>et al.</i> , 2013)
W303 Δ COX1/COX9-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox12::HIS3 cox1::ARG8^m leu2::pG16/ST14</i>	ρ^+	This study
aW303/SCO1-CH	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 sco1::URA3 trp1::pG41/ST50</i>	ρ^+	This study
W303 Δ SCO1/COX2-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 sco1::URA3 trp1::pG41/ST50</i>	COX2-HAC	This study
aW303/COA6-CH	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 coa6::HIS3 trp1::pCOA6/ST3</i>	ρ^+	This study
aW303 Δ MSS51/COX12-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox12::HIS3 mss51::HIS3 trp1::pCOX12/ST4</i>	ρ^+	This study

aW303/COX13-HAC	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox12::HIS3 trp1::pCOX13/ST4</i>	ρ^+	(McStay et al, 2013a)
aW303 Δ MSS51/COX13-HAC	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox12::HIS3 mss51::HIS3 trp1::pCOX13/ST4</i>	ρ^+	This study
aW303 Δ COX16	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox16::URA3</i>	ρ^+	(Carlson et al, 2003)
aW303/COX16-CH	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox16::URA3 trp1::pG22/ST13</i>	ρ^+	This study
aW303 Δ MSS51/COX16-CH	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox16::URA3 trp1::pG22/ST13 mss51::HIS3</i>	ρ^+	This study
aW303 Δ COX18	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox18::URA3</i>	ρ^+	(Souza et al, 2000)
aW303 Δ COX18-HAC	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox18::URA3 leu2::pG34/ST13</i>	ρ^+	This study
aW303 Δ COX20	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 cox20::URA3</i>	ρ^+	(Hell et al, 2000)
aW303 Δ COX20-HAC	MATa <i>a ade2-1 his3-11,15 leu2-3,112 trp1-1 ura3-1 cox20::URA3 leu2::pG92/ST20</i>	ρ^+	This study
aW303 Δ PET100	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 pet100::URA3</i>	ρ^+	This study
aW303 Δ PET100-CH	MATa <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 pet100::URA3 trp1::pG135/ST6</i>	ρ^+	This study
W303 Δ PET117	MATα <i>ade2-1 his3-1,15 leu2-3,112 trp1-1 ura3-1 pet117::HIS3</i>	ρ^+	This study

W303/PET117-CH

MAT α *ade2-1 his3-1,15 leu2-3,112*
trp1-1 ura3-1 pet117::HIS3
trp1::pG8/ST10

ρ^+

This study

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Table S2. Sequences of primers

Primer name	Primer sequence (5' -> 3')
	<i>Primer used to amplify a COX2-HA hybrid gene</i>
cox2-5	ggcgagtccttgctgcatacctttat
cox2-13	ggcggatccttattaagcgtagctcgggacgctgatgggtattgttcatttaattccaaaaattagg
cox2-11	ggcggatccttaattacttattattataatattttaa
cox2-12	ggcctgcagcaaggatactatcaaaaatagttac
	<i>Primers used to amplify the mitochondrial COX2-HAC hybrid gene</i>
cox2-5	ggcgagtccttgctgcatacctttat
HA-proC	ggcggatccttacttaccatcgattaaccgtggatctacctgatcttcacctccagcgtagctcgggacgctgta
	<i>Primers used to construct nuclear genes expressing HAC or CH tagged proteins</i>
cox18-1	gaaaggcggatcccaaatgagtttaataagagatg
cox18-2	ggcgagctcctcaagcgtagctcgggacgctgatcgttggaaggataaatccaa
C-phs	ggcctgcagtcattaccatcgattaatcttggatctacttgatcttctcctccagcgtagctcgggacgctgatgg
cox20-1	ggcggatcccaactagctctattagtt
cox20-2	ggcgagctcctcaagcgtagctcgggacgctgtaatgggaccagaactgtaccatttctt
cox16-1	ggcgagctcctcgggaaggttcaactcat
cox16-2	ggcctgcagccagacattctcagattcatc
pet100-3	ggcgagctccgctccatcattagatccttc
pet100-5	ggcctgcagtcctttttgagagccaaatcccg
pet117-1	ggcgagctccgctccatcattagatccttc
pet117-2	ggcctgcagtcctttttgagagccaaatcccg
cyc1-1	ggcgagctcatggagaaaggagtataatg
cyc1-2	ggcctgcagtttcttagattcttaaccac
sco1-1	ggcgagctccaatcgcgatgctatcactc
sco1-2	ggcggatcctttgaataagaaggagtaccatgc
coa6-5	ggcgagctcgcgatggtatattggcacctgagc
coa6-6	ggcctgcagctgattcgttcctctgttttag

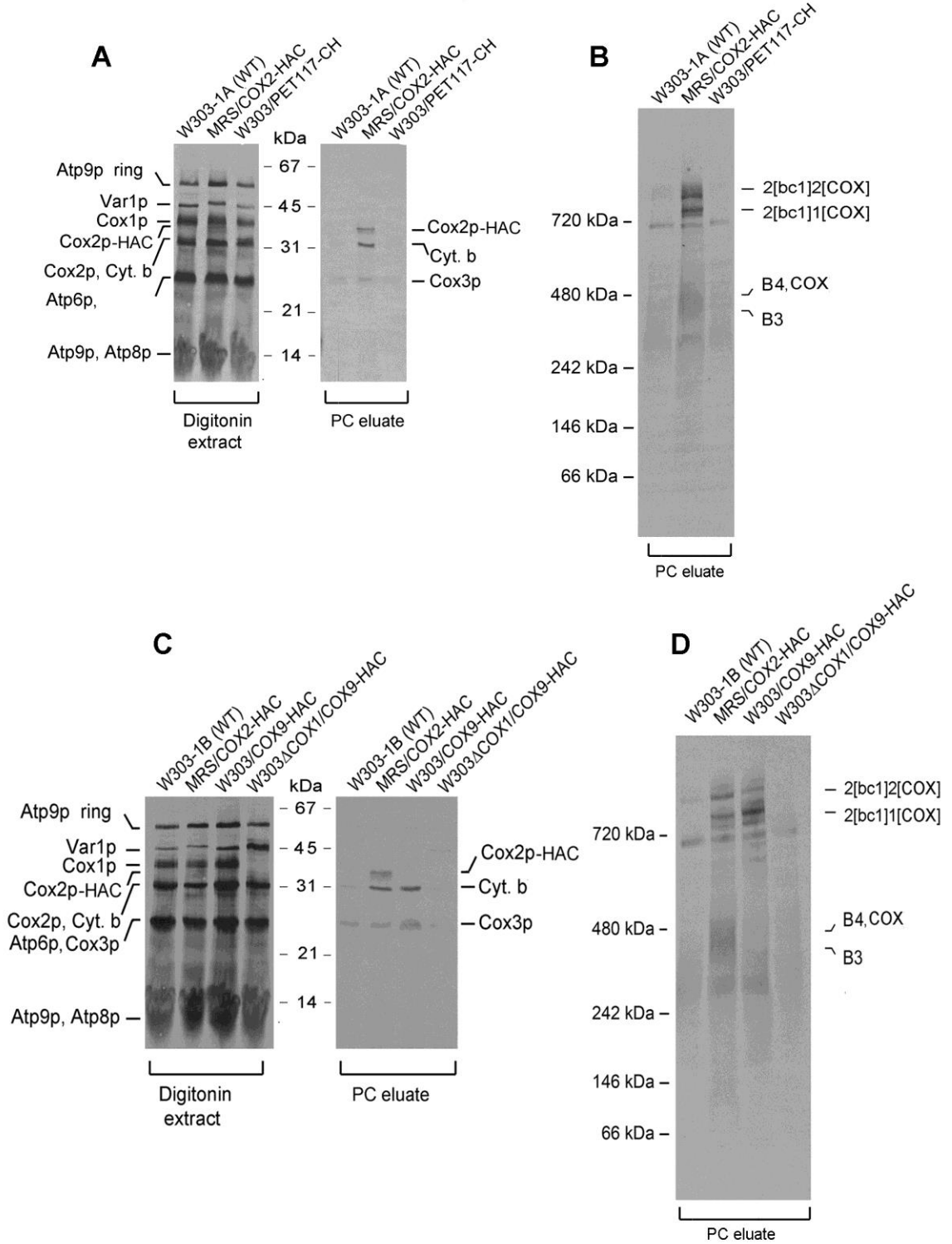


Fig. S1. Pull-down assays of radiolabeled mitochondrial gene products of strains expressing Cox9p-HAC and Pet117p-CH. Mitochondria of the respiratory competent strain W303-1B and of strains expressing tagged Cox2 (MRS/COX2-HAC), Cox9 (W303/COX9-HAC) and Pet117 (W303/PET117-CH) with and without an additional *mss51* mutation, were grown in 2% YPGal to early stationary phase. The cells were harvested and inoculated into the same volume of fresh YPGal plus 2 mg/ml chloramphenicol and shaken for an additional 2 hours at 30°C. **A.** Mitochondria were labeled for 20 min and chased for 10 min after addition of puromycin and excess cold methionine. The labeled mitochondria were extracted with digitonin and protein C tagged proteins were purified on protein C antibody beads. The digitonin extracts and eluates from the protein C antibody beads (PC eluates) were separated by SDS-PAGE on a 12% polyacrylamide gel. Proteins were blotted onto nitrocellulose and exposed to X-ray film. **B.** The PC eluates from **A** were separated by BN-PAGE on a 4-13% polyacrylamide. Proteins were transferred to a PVDF membrane and exposed to X-ray film. **C.** Mitochondria from the indicated strains were labeled and further processed as in **A**. **D.** The PC eluates from **C** were transferred to a PVDF membrane and exposed to X-ray film. The radiolabeled mitochondrial products and assembly intermediates are marked in the margins of the autoradiograms.