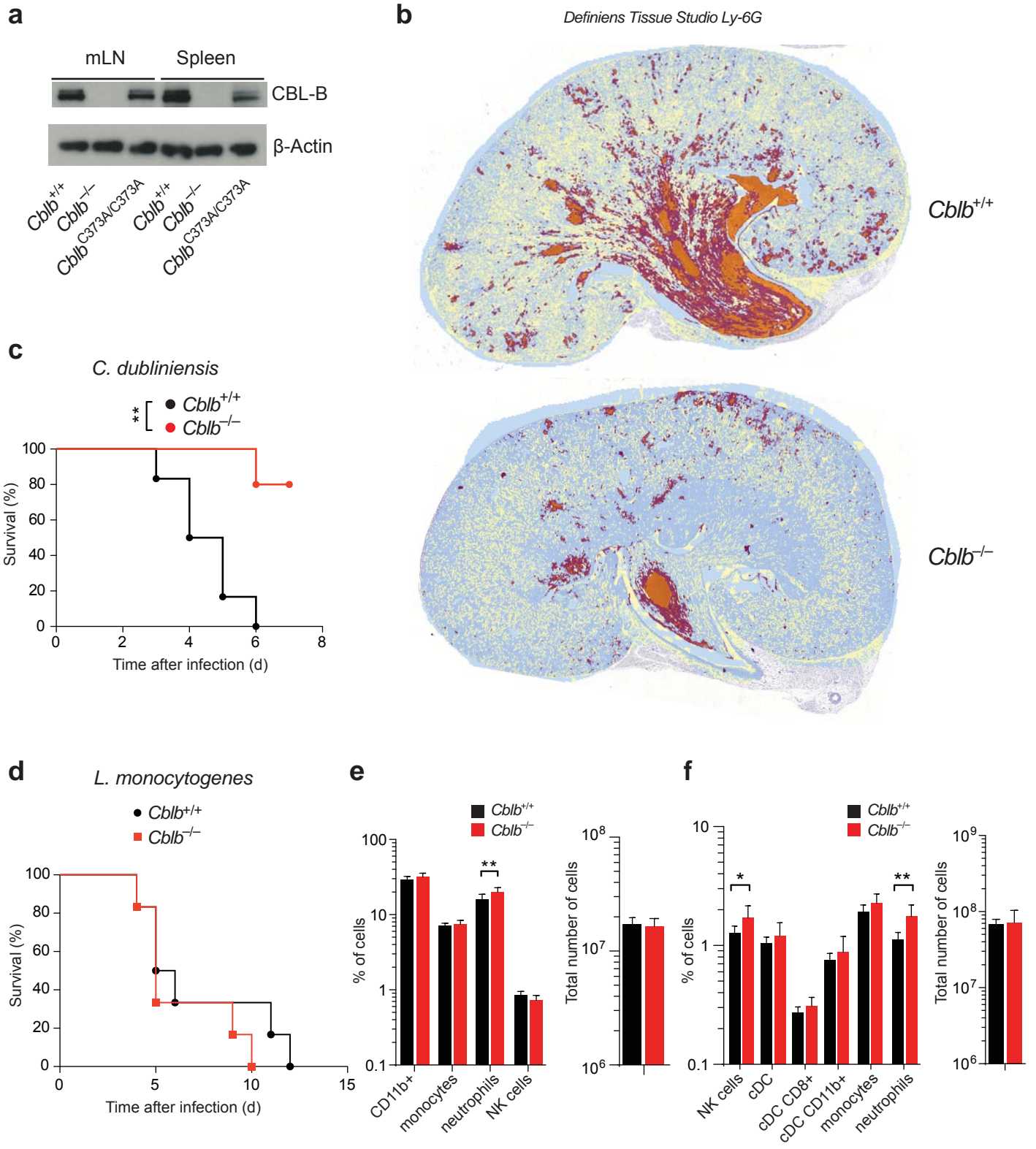


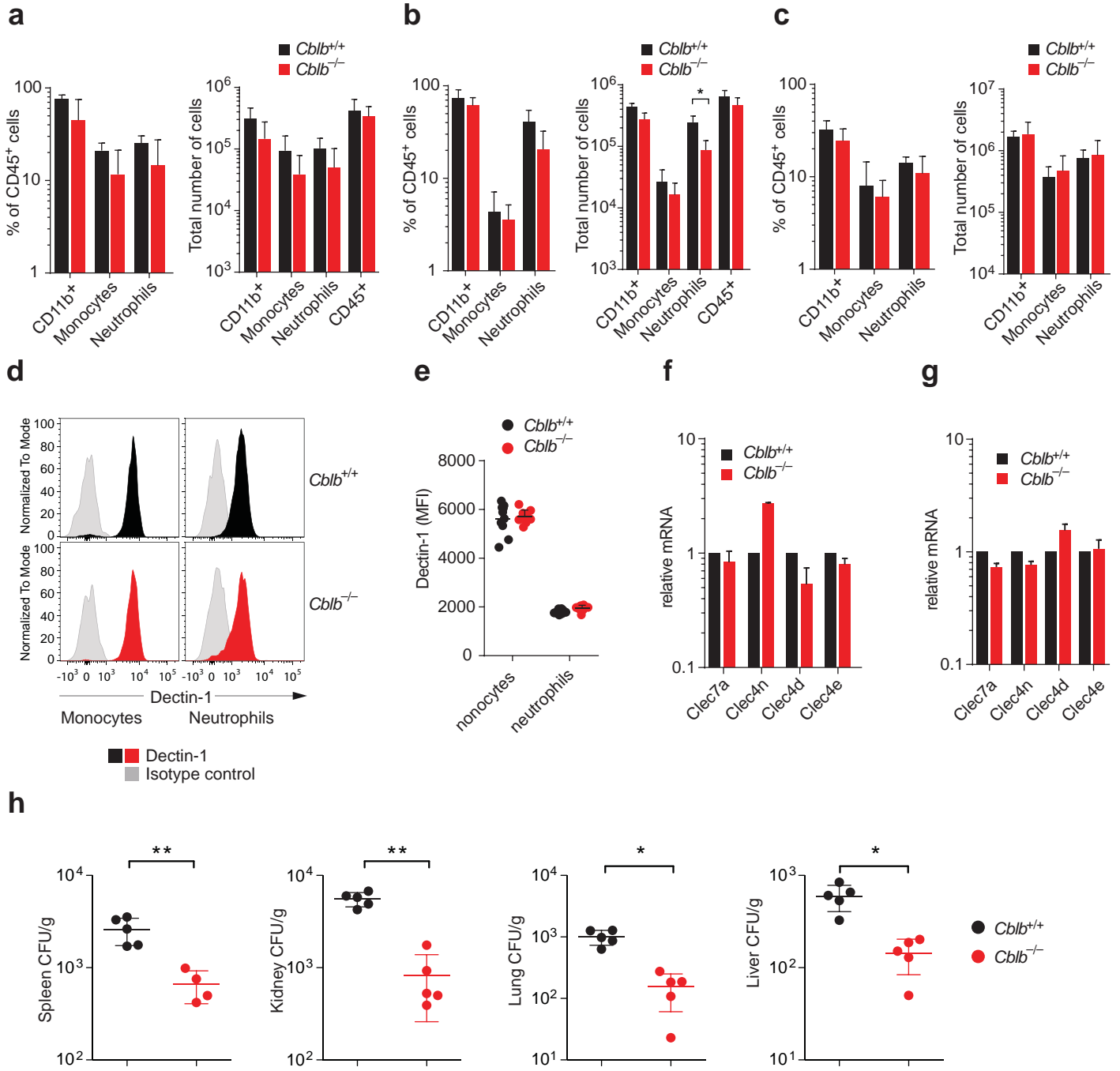
Supplementary Figure 1



Supplementary Figure 1. *Candida ssp.* and *Listeria monocytogenes* infection.

(a) Protein levels of CBL-B in lysates prepared from mesenteric lymph nodes (mLN) or spleens of mice of the indicated genotypes as assessed with Western blotting. β -Actin is shown as loading control. (b) Representative Definiens Tissue Studio software analysis of Ly-6G stained kidney sections from **Figure 1h**. (c) *Cblb*^{+/+} and *Cblb*^{-/-} mice were infected intravenously with *C. dubliniensis* (10^6 CFUs) and monitored for the indicated period of time. Plot depicts survival over time after infection (*P* values assessed with log rank test) *n* = 5 for *Cblb*^{+/+} and *n* = 6 for *Cblb*^{-/-} cohort. (d) *Cblb*^{+/+} and *Cblb*^{-/-} mice were infected intravenously with *L. monocytogenes* (10^5 CFUs) and monitored for the indicated period of time. Plot depicts survival over time after infection. (e,f) Frequency of indicated innate immune cell lineages and total immune cell counts of (e) femurs or (f) spleens of mice of the indicated genotypes as assessed by flow cytometry. *n* = 8 for *Cblb*^{+/+}, *n* = 11 for *Cblb*^{-/-} littermates. Data in e,f are shown as means \pm standard deviation. For panels (a,b,d,e,f) 1 representative of 3, for panel (c) 1 representative of 2 independent experiments is shown. **P* < 0.05, ***P* < 0.01 as calculated with *Student's t-test*, unless stated otherwise.

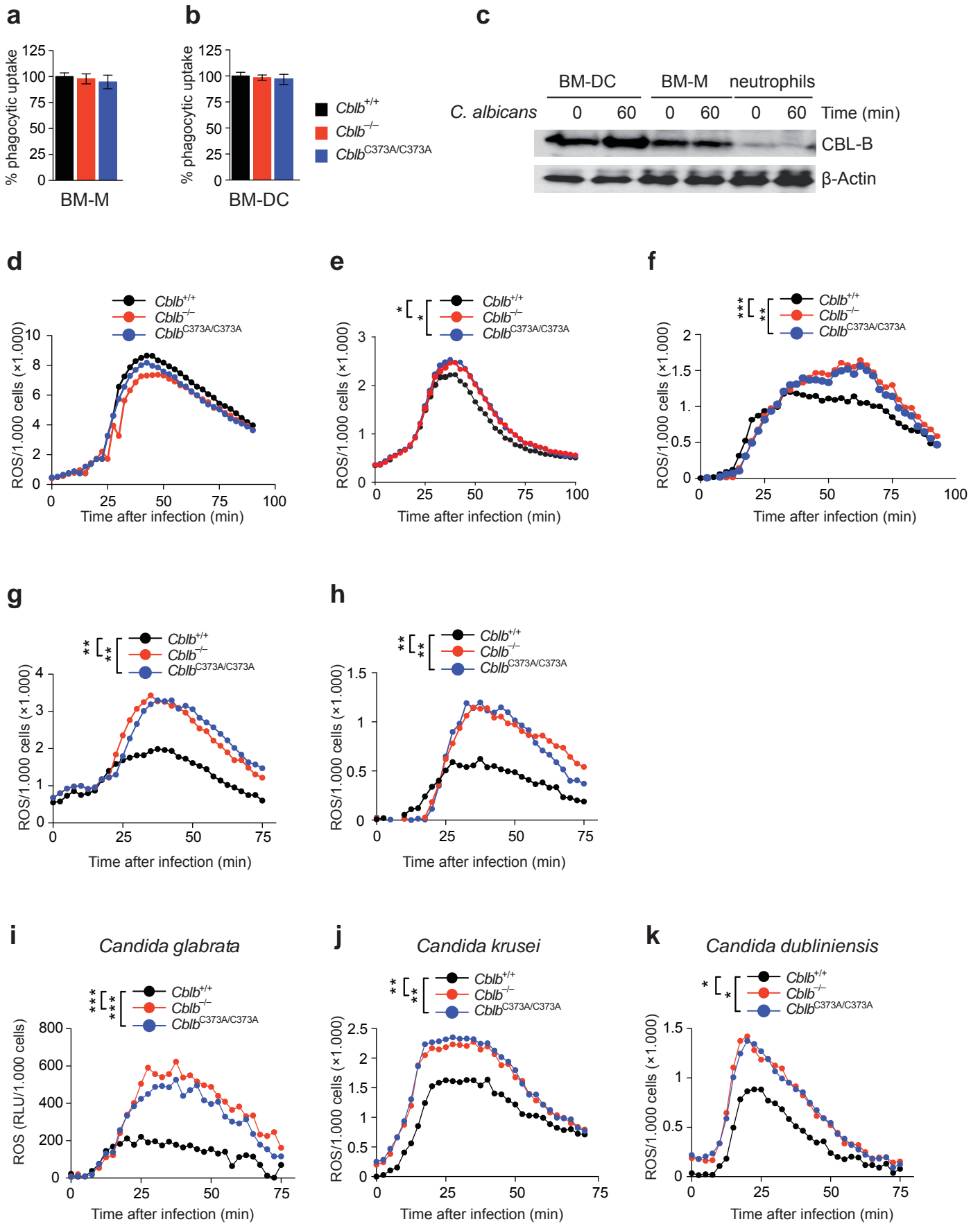
Supplementary Figure 2



Supplementary Figure 2. Immune infiltrations, CLR expression, and early fungal loads.

(**a,b,c**) Cellular recruitment of the indicated immune cell lineages to kidneys (**a**), lungs (**b**), and livers (**c**) of *Cblb*^{+/+} and *Cblb*^{-/-} mice 24 h after infection with *C. albicans* (10⁵ CFU/21.5 g body weight). Plots depict percentages of CD45⁺ hematopoietic cells (left panels) or total numbers of recruited cells per organ (right panels). *n* = 5 for *Cblb*^{+/+}, *n* = 6 for *Cblb*^{-/-} mice. (**d,e**) Dectin-1 protein expression on peripheral blood monocytes and neutrophils isolated from *Cblb*^{+/+} and *Cblb*^{-/-} mice as assessed by flow cytometry. (**d**) Representative stainings and (**e**) Dectin-1 mean fluorescence intensities (MFI) are shown. Dots in (**e**) represent individual mice. (**f,g**) mRNA expression levels of the indicated C-type lectin receptors in (**f**) bone marrow cells and (**g**) splenocytes of *Cblb*^{+/+} and *Cblb*^{-/-} mice as assessed by quantitative PCR. Experiments were done in triplicates. (**h**) *C. albicans* fungal load in depicted organs 24 h after infection, plotted as colony forming units (CFU) / gram organ weight. Each dot represents an individual mouse. For panels (**a,b,c,e,f,g,h**) data are shown as means ± standard deviation. For panels (**a,b,c,f,g**) 1 representative of 3, for panels (**d,e**) 1 representative of 5 independent experiments is shown. **P* < 0.05, ***P* < 0.01 as calculated with *Student's t-test*.

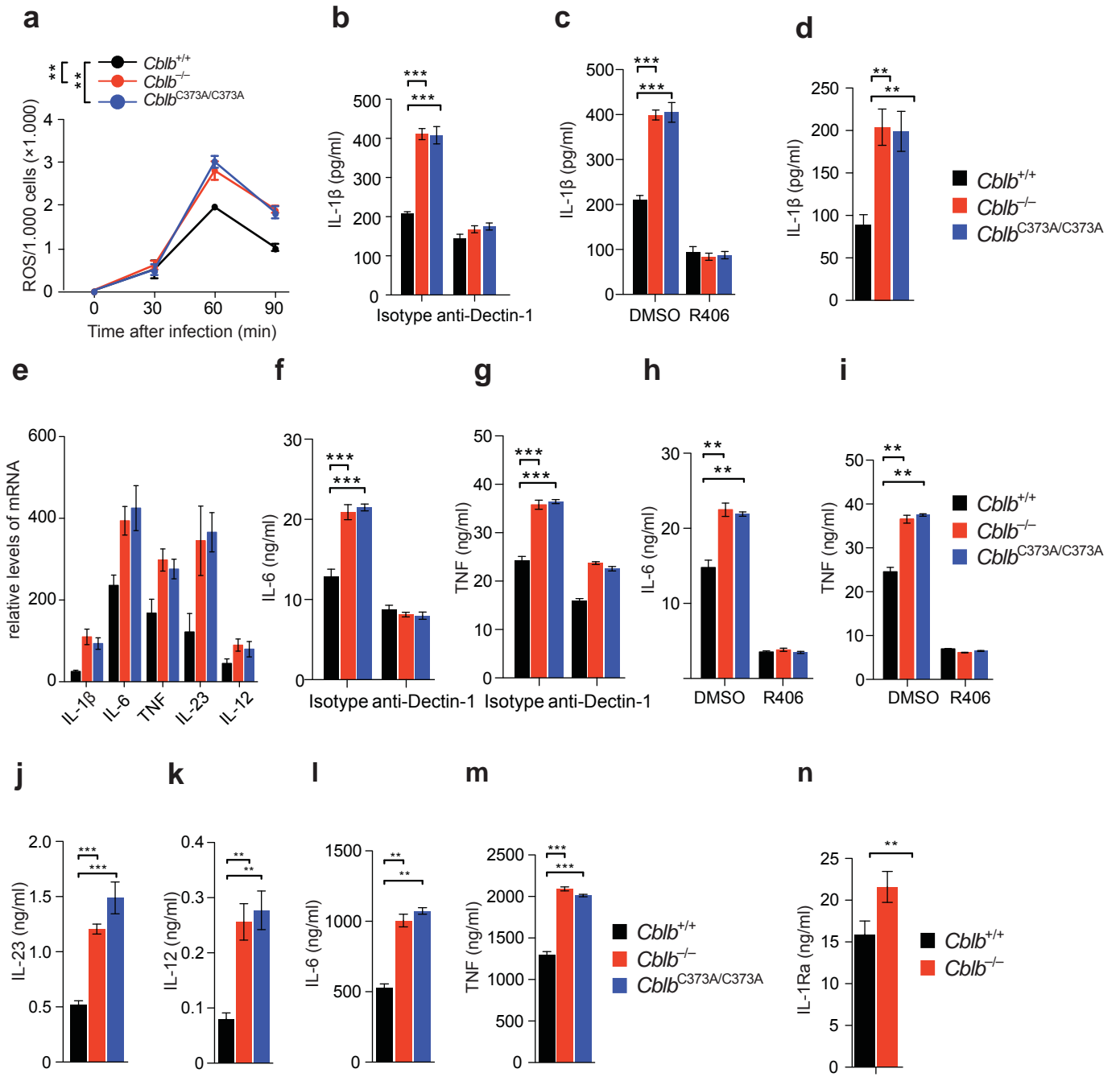
Supplementary Figure 3



Supplementary Figure 3. Phagocytosis and ROS production in response to fungal stimulation.

(a,b) Rate of phagocytosis by (a) BM-M or (b) BM-DC of the indicated genotypes co-cultured with *C. albicans*. (c) CBL-B protein expression as assessed by Western blotting from lysates prepared from *C. albicans* stimulated or unstimulated BM-DC, BM-M or bone marrow neutrophils derived from *Cblb*^{+/+} mice. β -Actin blots are shown as loading control. (d-h) Reactive oxygen species (ROS) production by (d) bone marrow neutrophils, (e) BM-M, (f) BM-DC, (g) bone marrow monocytes, or (h) splenic DC co-cultured with *C. albicans* and monitored in real time over the indicated time periods using the luminol assay. Experiments were performed in triplicates. Values are expressed as relative light units per 1.000 cells. (i-k) ROS production by BM-DC of the indicated genotypes stimulated with (i) *C. glabrata*, (j) *C. krusei*, or (k) *C. dubliniensis*, monitored and analyzed as in (d-h). Data in panels (a,b) are shown as means \pm standard deviation. *P* values for panel (a,b) were calculated with *Student's t-test*, for panels (d-k) with two-way ANOVA. For all experiments 1 representative of 3 independent experiments is shown. **P* < 0.05, ***P* < 0.01, ****P* < 0.001.

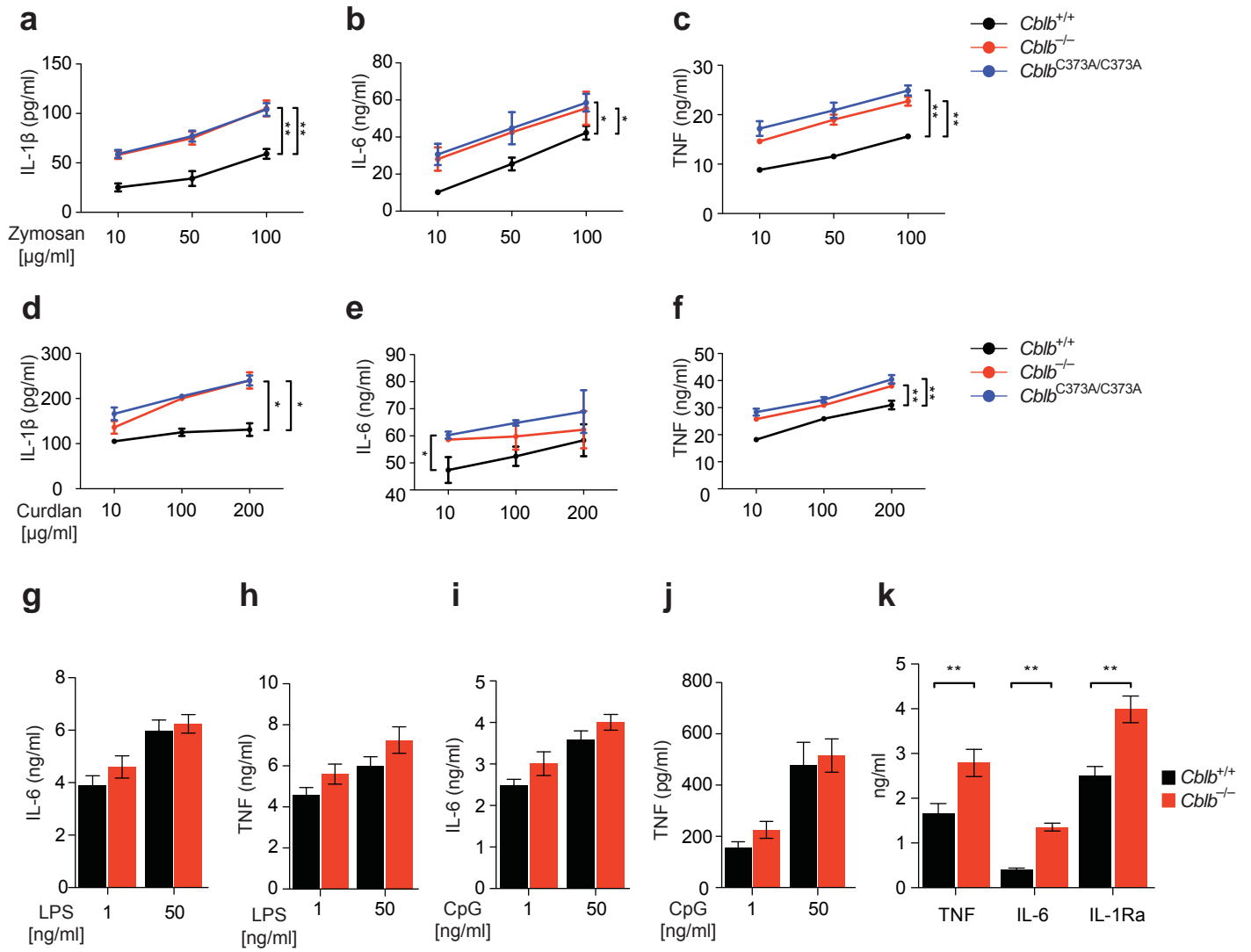
Supplementary Figure 4



Supplementary Figure 4. Cytokine release in response to fungal stimulation.

(a) Activation of Caspase-8 in BM-DC of the indicated genotypes monitored at the indicated time points after stimulation with *C. albicans*, using the CaspGLOW assay. **(b,c)** Secretion of IL-1 β by BM-DC of the indicated genotypes stimulated with *C. albicans* for 18 h in the presence of **(b)** an anti-Dectin-1, or isotype control antibody, or **(c)** the SYK inhibitor R406, or DMSO as assessed by ELISA. **(d)** Secretion of IL-1 β by BM-M upon 18 h co-culture with *C. albicans* as assessed with ELISA. **(e)** Induction of cytokine RNA expression in BM-DC of the indicated genotypes stimulated for 2 h with *C. albicans*, as assessed by quantitative PCR. Experiments were done in triplicates. **(f-i)** Secretion of the indicated cytokines by BM-DC of the indicated genotypes upon 18 h co-culture with *C. albicans* in the presence of **(f,g)** an anti-Dectin-1, or isotype control antibody, or **(h,i)** the SYK inhibitor R406, or DMSO as assessed with ELISA. **(j-n)** Secretion of the indicated cytokines by **(j,k,n)** BM-DC, or **(l,m)** BM-M of the indicated genotypes upon 18 h co-culture with *C. albicans* as assessed with ELISA. Data in all panels are shown as means \pm standard deviation. *P* values for all panels except **(a)** were calculated with *Student's t-test*, for panel **(a)** with two-way ANOVA. For panels **(a-d,f-n)** 1 representative of 4, for panels **(e)** 1 representative of 5 independent experiments is shown. ***P* < 0.01, ****P* < 0.001.

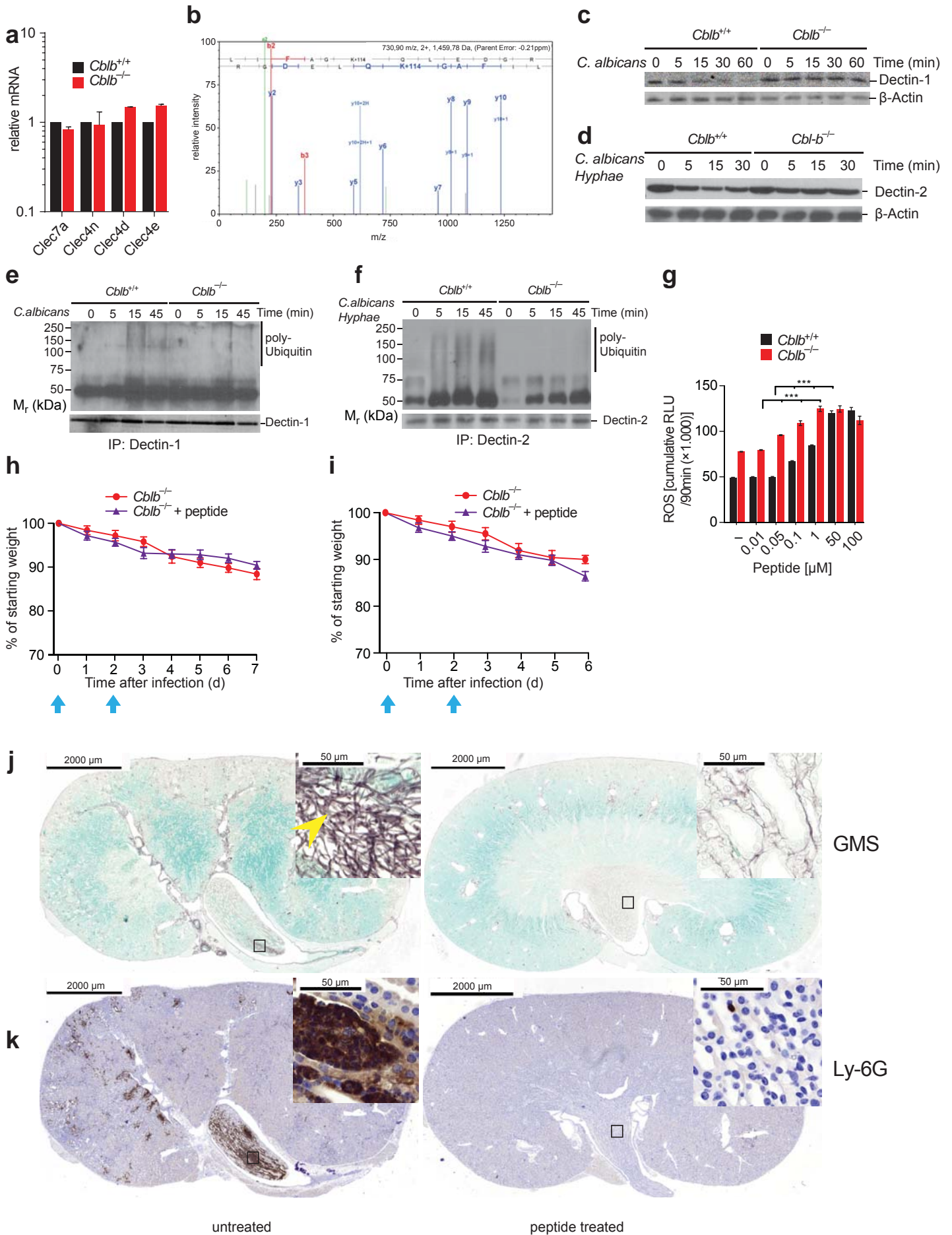
Supplementary Figure 5



Supplementary Figure 5. Zymosan, Curdlan, LPS, CpG, and *C. albicans* triggered cytokine release by BM-DC and intraperitoneal inflammatory infiltrates.

(a-f) Secretion of the indicated cytokines by *Cblb*^{+/+}, *Cblb*^{C373A/C373A} or *Cblb*^{-/-} BM-DC stimulated with increasing doses of either Zymosan (a-c), or Curdlan (d-f) for 18 h, as assessed by ELISA. (g-j) Secretion of the indicated cytokines by *Cblb*^{+/+} or *Cblb*^{-/-} BM-DC stimulated with the indicated doses of either LPS (g,h), or CpG (i,j) for 18 h, as assessed by ELISA. (k) Expression of the indicated cytokines in the intraperitoneal lavages of *Cblb*^{+/+} or *Cblb*^{-/-} mice 24 h after intraperitoneal infection with *C. albicans* as in **Figure 3h-I** as assessed by ELISA. Data in all panels are shown as means ± standard deviation. *P* values as calculated with (a-f) two-way ANOVA, or (k) *Student's t-test*. For all panels 1 representative of 3 independent experiments is shown. **P* < 0.05, ***P* < 0.01

Supplementary Figure 6



Supplementary Figure 6. SYK and Dectin-1, -2 ubiquitination and CBL-B inhibitory peptide treatment.

(a) mRNA expression levels of the indicated C-type lectin receptors in *Cblb*^{+/+} and *Cblb*^{-/-} BM-DC assessed by qPCR. Experiments were done in triplicates. (b) Mass spectrometric analysis of the ubiquitin chains generated by *in vitro* ubiquitination by CBL-B. A representative MS/MS spectrum acquired for a peptide derived from K48-linked Ubiquitin chains, using a sample processed identically and in parallel to the sample shown in **Figure 5a**, is shown. Both β -type (red) and γ -type (blue) ions of the selected peptide are indicated. (c,d) Immunoblot analysis of (c) Dectin-1 and (d) Dectin-2 protein expression in *Cblb*^{+/+} and *Cblb*^{-/-} BM-M upon infection with (c) *C. albicans* or (d) *C. albicans* hyphae with an anti-Dectin-1 or anti-Dectin-2 antibody, respectively. β -Actin blots are shown as loading control. (e,f) Immunoblot analysis of (e) Dectin-1 and (f) Dectin-2 ubiquitination in E64-pre-treated *Cblb*^{+/+} and *Cblb*^{-/-} BM-M after immunoprecipitation with an anti-Dectin-1 or anti-Dectin-2 antibody, respectively, with an anti-ubiquitin antibody. Dectin-1 and Dectin-2 immunoblots are shown as controls. (g) ROS production by *Cblb*^{+/+} and *Cblb*^{-/-} BM-DC co-cultured with *C. albicans* in the presence of the indicated concentrations of the TKB-binding peptide. Plot depicts cumulative ROS production over 75 minutes for the indicated peptide concentrations. Experiments were performed in triplicates. Values are expressed as relative light units per 1.000 cells. (h,i) *Cblb*^{-/-} mice were infected intravenously with *C. albicans* (10^5 CFU/21.5g body weight), injected intraperitoneally with (h) 100 μ g or (i) 20 μ g of the TKB-binding peptide at day 0 and day 2 after infection, and monitored for weight loss over time after infection as compared to starting weight (*P* values assessed by two-way ANOVA). *n* = 5 for treated and untreated cohorts. (j,k) Representative kidney sections from *Cblb*^{+/+} mice treated as in **Figure 6g**, and stained with (j) Gomori methenamine silver (GMS) or (k) for the neutrophil marker Ly-6G. Yellow arrowhead in (j) indicates fungal hyphae. *n* = 3 for peptide treated and untreated cohorts. 4 kidney sections per mouse were analyzed. Data in (a,g) are shown as means \pm standard deviation. For panel (a,b) 1 representative of 4, for panel (c-k) 1 representative of 3 independent experiments is shown. ****P* < 0.001 as calculated with *Student's t-test*.

Antibodies and Primers used in this study

Antibodies

Antigen	Clone	Distributor
Syk/SYK	D1I5Q	Cell Signaling Technologies
phospho Syk	C87C1	Cell Signaling Technologies
Src	36D10	Cell Signaling Technologies
phospho Src	D7F2Q	Cell Signaling Technologies
Plc- γ 2	Q-20	Santa Cruz Biotechnology
phospho-Plc- γ 2	Tyr1217	Cell Signaling Technologies
NF- κ B p65	E498	Cell Signaling Technologies
phospho-NF- κ B p65	93H1	Cell Signaling Technologies
Shp-2	C-18	Santa Cruz Biotechnology
phospho-Shp-2	Tyr542	Cell Signaling Technologies
β -Actin	13 E 6	Cell Signaling Technologies
Ubiquitin	P4D1	Santa Cruz Biotechnology
Ly-6G FITC	1A8	BD Biosciences
CD8 Brilliant Violet 605	53-6.7	Biolegend
TCR β Brilliant violet 421	H57-597	BD Biosciences
NK-1.1 APC	PK136	BD Biosciences
CD19 PE	6D5	BD Biosciences
CD11c FITC	HL3	Biolegend
CD16/32 purified	2.4G2	BD Biosciences
Ly-6C PerCP-Cy5.5	AL-21	BD Biosciences
anti-rabbit Alexa555	polyclonal	Invitrogen
anti-Dectin-1	GE2	Abcam
anti-Dectin-2	217611	R&D Systems

PCR Primers

Target	fwd	rev	amplicon
Clec4n	CTGCCCAAATCACTGGAAGT	GAAATTCTGCTCCGCTTCAG	142bp
Clec7a	GAACCACAAGCCCACAGAAT	TAGGAAGGCAAGGCTGAGAA	95bp
Clec4d	TACGCTGGACGAGAGGAAGT	ACAGGACAGCAGGTCCAAGT	112bp
Clec4e	CAAGTGCTCTCCTGGACGAT	CTGTAAGTTCTGCCCGGAAA	114bp
TNF	CAAAATTCGAGTGACAAGCCTG	GAGATCCATGCCGTTGGC	88bp
IL-6	GAGGATACCACTCCCAACAGACC	AAGTGCATCATCGTTGTTTCATACA	140bp
IL-12	GGAAGCACGGCAGCAGAATA	AACTTGAGGGAGAAGTAGGAATGG	159bp
IL-23	GCAGATTCCAAGCCTCAGTC	TTCAACATATGCAGGTCCCA	86bp
IL-1 β	CAACCAACAAGTGATATTCTCCATG	GATCCACACTCTCCAGCTGCA	120bp