

Supplementary Figure 1: Construction of deletion mutants in DSM12444. Deletion plasmids were synthesized *de novo*, with 450 bp regions of homology (orange and purple) flanking the gene to be deleted (green). Approximately 15 bp of the 5' and 3' ends of the gene were retained to minimize polar effects, expanding as necessary to avoid disrupting adjacent genes. Plasmids were introduced into DSM12444 through biparental mating with DAP-auxotrophic *E. coli* containing the appropriate deletion plasmid. Strains with a single crossover integrating the plasmid into the genome are kanamycin-resistant and streptomycin-sensitive (due to a sensitive

rpsL allele). Sequential selection for kanamycin resistance and then streptomycin resistance selects for integration and then loss of the plasmid, either reverting to the WT genotype or producing the desired deletion. The deletion strains can then be identified by colony PCR.



Supplementary Figure 2: Gene deletions affect growth with PCA. Clean deletions were constructed in *ligA*, *ligU*, and *xylG*, and growth with 1 g/L PCA as the sole source of carbon and energy. As predicted from the pooled fitness data, deletion of *ligA* did not affect growth with PCA, while deletion of *ligU* and *xylG* reduced growth. Error bars show one standard deviation, calculated from three biological replicates. * : p<0.05. ***: p<0.001.



Supplementary Figure 3: Genes involved in deacetylation of sinapate to syringate.

Phenylpropanoids can be deacetylated through the combined action of a CoA-ligase and hydratase/lyase, FerAB, and then oxidized by LigV. Disruption of any of the associated genes significantly diminishes fitness with the phenylpropanoid but not with the associated benzoate derivative. Error bars show one standard deviation, calculated from three biological replicates.



Supplementary Figure 4: Gene disruptions do not affect growth with other phenylpropanoids. Transposon insertions in *desC* and *desD* diminish fitness with sinapate, but not with ferulate or coumarate. The operon structure of the genetic loci is shown above the graph. Error bars show one standard deviation, calculated from three biological replicates.





Gene	Locus	Annotation	SYK-6	SYK-6
name			homolog	%AA
			0	identity
ligA	SARO_RS14270	Protocatechuate 4 5-	SLG_RS06125	68
		dioxygenase subunit alpha		
ligB	SARO_RS14265	Protocatechuate 3 4-	SLG_RS06120	70
		dioxygenase subunit beta		
ligC	SARO_RS14260	Oxidoreductase	SLG_RS06115	87
ligI	SARO_RS14300	2-pyrone-4 6-dicarboxylate	SLG_RS06155	83
		hydrolase		
ligU	SARO_RS14295	4-oxalomesaconate	SLG_RS06150	78
		tautomerase		
ligJ	SARO_RS14275	4-oxalomesaconate hydratase	SLG_RS06130	91
ligK	SARO_RS14290	4-carboxy-4-hydroxy-2-	SLG_RS06145	91
		oxoadipate		
		aldolase/oxaloacetate		
		decarboxylase		
	SARO_RS12260	4-hydroxybenzoate 3-	SLG_RS08320	72
		monooxygenase		
ligM	SARO_RS14510	Vanillate demethylase	SLG_RS06245	78
metF	SARO_RS14505	5,10-methylenetetrahydrofolate	SLG_RS06250	51
		reductase		10
purU	SARO_RS14515	Formyltetrahydrofolate		<40
6.10	GADO DG10020	deformylase		~ 1
folD	SARO_RS19830	bifunctional 5 10-methylene-	SLG_RS01620	54
		tetrahydrofolate		
		denydrogenase/5 10-		
		avalobydrologa (folD)		
for	SADO DS02265	Equiport CoA synthetese	SI C DS12205	67
forD	SARU_RS03303	n hydroxyginnamoyl CoA	SLO_KS12393	61
јегь	SARU_R500505	p-flydroxychillanioyi COA	SLO_K512400	01
ligV	SARO RS08360	Salicylaldebyde debydrogenase	SI G PS03480	87
des A	SARO R\$12005	Suringate O_demethylase	SLG_RS12385	71
desC	SARO RS1/1525	Alpha/beta hydrolase	SLG_RS12303	47
dest	SARO RS1/1530	Glutathione S-transferase	<u>510_K500255</u>	<40
ucsD	SARO RS14535	Short chain dehvdrogenase	SLG R\$06215	5 9
	SARO RS1/15/15	4-hydroxyphenyl_beta_	SLG_RS06210	83
	5/110_1014343	ketoacyl-CoA hydrolase		05
	SARO RS14550	Aldehyde dehydrogenase	SLG RS04675	50
xvlE	SARO RS17090	Catechol 2.3-dioxygenase	220_1001073	<40
xylG	SARO RS17090	2-hydroxymuconic		<40
		semialdehyde dehydrogenase		
xvlH	SARO RS17055	4-oxalocrotonate tautomerase		<40
ligM metF purU folD folD ferA ferB ligV desA desC desD xylE xylG xylH	SARO_RS12260 SARO_RS14510 SARO_RS14505 SARO_RS14515 SARO_RS19830 SARO_RS19830 SARO_RS08365 SARO_RS08365 SARO_RS08360 SARO_RS08360 SARO_RS14525 SARO_RS14525 SARO_RS14535 SARO_RS14535 SARO_RS14535 SARO_RS14550 SARO_RS14550 SARO_RS17090 SARO_RS17080 SARO_RS17080	 4-hydroxybenzoate 3- monooxygenase Vanillate demethylase 5,10-methylenetetrahydrofolate reductase Formyltetrahydrofolate deformylase bifunctional 5 10-methylene- tetrahydrofolate dehydrogenase/5 10- methylene-tetrahydrofolate cyclohydrolase (foID) Feruloyl-CoA synthetase p-hydroxycinnamoyl CoA hydratase/lyase Salicylaldehyde dehydrogenase Syringate O-demethylase Alpha/beta hydrolase Glutathione S-transferase Short chain dehydrogenase 4-hydroxyphenyl-beta- ketoacyl-CoA hydrolase Aldehyde dehydrogenase 2-hydroxymuconic semialdehyde dehydrogenase 4-oxalocrotonate tautomerase 	SLG_RS08320 SLG_RS06245 SLG_RS06250 SLG_RS01620 SLG_RS01620 SLG_RS12395 SLG_RS12400 SLG_RS03480 SLG_RS03480 SLG_RS06235 SLG_RS06210 SLG_RS06210 SLG_RS04675	72 78 51 <40 54 67 61 87 71 47 <40 59 83 50 <40 <40 <40

Supplementary Table 1: Genes referenced in this study

xylI	SARO_RS17060	4-oxalocrotonate decarboxylase	<40
xylJ	SARO_RS17075	2-keto-4-pentenoate hydratase	<40
xylK	SARO_RS17065	4-hydroxy-2-oxovalerate	<40
		aldolase	
xylQ	SARO_RS17070	Acetaldehyde dehydrogenase	<40