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Supporting information for article:

Structures of collagen IV globular domains: insight into associated pathologies, folding and network assembly

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Table S1 Primers used for α NC1 amplification.

Product	Primer name	Primer sequences
α 1(IV)NC1	NCI1_Flag1/2_FWD	GACAAGGGCCCCGATGGGTTGCCAGGATCCATGGGGCCC
	NCI1_Flag_FWD	GACTACAAGGACGACGATGACAAGGGCCCCGATGGGTTGCCAGGATCC
	NCI1_REV	TAGCTGAGTCAGGCTTCATTATGTTCT
	NCI1_KpnI_REV	CCATGTTGGTACCTTAGCTGAGTCAGGCTTCATTATGTTCT
α 2(IV)NC1	NCI2_Flag1/2_FWD	GACAAGGGCCGTCCAGGGAGCCCGGGCTGCCGGGTATG
	NCI2_Flag_FWD	GACTACAAGGACGACGATGACAAGGGCCGTCCAGGGAGC
	NCI2_REV	TTCCTGGCACCGCGCCGGCTCACAGGTT
	NCI2_SacI_REV	TTCCTGGCAGAGCTCGGCTCACAGGTT
α 4(IV)NC1	NCI4_Flag1/2_FWD	GACAAGCTAGCTGGTCCCATTGGGATCCTGGGCCAAA
	NCI4_Flag_FWD	GACTACAAGGACGACGATGACAAGCTAGCTGGTCCCATTGGGATCCTGGGCC
	NCI4_REV	GCCGAGGGCCCCTAGCTATACTTCACGCAG
	NCI4_KpnI_REV	ATTGGTACCGCCGAGGGCCCCTAGCTA
α 5(IV)NC1	NCI5_Flag1/2_FWD	GACAAGGGTCCAGATGGATTGCAAGGTCCCCCAGGTCCC
	NCI5_Flag_FWD	GACTACAAGGACGACGATGACAAGGTCCAGATGGATTGCAAGGTCCC
	NCI5_Flag_REV	AAGGAATTCTCAAAATGTTATGTCCT
	NCI5_SacI_REV	GGTGAGCTCAAGGAATTCTCAAAATG
Cloning Bm40 and Flag	Flag_SignalP1_FWD	AGGGCTCTGGCAGCCCCACTAGCCGACTACAAGGACGACGATGACAAG
	SignalP1_SignalP2_FWD	TTCTTCTCCTTGCCCTGGCGGGAGGGCTCTGGCAGCCCCACTAGCC
	SignalP2_BamHI_FWD	AGGGGGATCCATGAGGGCCTGGATCTCTCCTTGCCTG
	SignalP2_XhoI_FWD	AGGGCTCGAGATGAGGGCCTGGATCTCTCCTTGCCTG

Table S2 Residue numbers for the structures of α 1, α 2, α 3, α 4 and α 5 NC1 domains and their corresponding residue numbers according to the UniprotKB accession number.

Collagen α (IV) NC1 domain	Residue numbers in each chain	Corresponding residue numbers in the complete α (IV) chain	
		UniprotKB accession number	Residue number
α 1NC1	1-229	P02462	1441-1669
α 2NC1	1-228	P08572	1485-1712
α 3NC1	1-230	Q01955	1441-1671
α 4NC1	1-230	P53420	1461-1690
α 5NC1	1-229	P29400	1461-1685

Table S3 Root mean square deviations (r.m.s.d.), in Å, for the superposition of the structures of NC1 hexamers.

The number of C α atoms superimposed is indicated between parentheses.

Rmsd (Å)	$\alpha 1\text{NC1}_{\text{homo}}$	$\alpha 3\text{NC1}_{\text{homo}}$	$\alpha 5\text{NC1}_{\text{homo}}$	$\alpha 121\text{NC1}$	$\alpha 121\text{NC1(PDB:1LI1)}^*$
$\alpha 1\text{NC1}_{\text{homo}}$	-	0.62 (1342)	0.55 (1356)	0.59 (1335)	0.73 (1346)
$\alpha 3\text{NC1}_{\text{homo}}$	-	-	0.67 (1344)	0.72 (1327)	0.84 (1329)
$\alpha 5\text{NC1}_{\text{homo}}$	-	-	-	0.67 (1338)	0.70 (1338)
$\alpha 121\text{NC1}$	-	-	-	-	0.48 (1348)

*Human $\alpha 121\text{NC1}$ obtained from placenta, PDB file 1LI1.

Table S4 Root mean square deviations (r.m.s.d.), in Å, for the superposition of the structures of the various recombinant NC1 chains crystallized here and the $\alpha 1\text{-}2\text{NC1}$ chains of human placenta.

The number of C α atoms superimposed is indicated between parentheses.

Rmsd (Å)	$\alpha 1\text{NC1}_{\alpha 121}$	$\alpha 2\text{NC1}^*$	$\alpha 2\text{NC1}_{\alpha 121}$	$\alpha 2\text{NC1}_{\text{homo}}$	$\alpha 3\text{NC1}_{\text{homo}}$	$\alpha 4\text{NC1}_{\text{homo}}$	$\alpha 5\text{NC1}_{\text{homo}}$
$\alpha 1\text{NC1}^*$	0.35 (226)	-	-	-	0.65 (224)	-	0.47 (226)
$\alpha 1\text{NC1}_{\alpha 121}$	-	-	0.75 (218)	1.44 (166)	0.53 (225)	1.27 (178)	0.41 (226)
$\alpha 2\text{NC1}^*$	-	-	0.45 (224)	1.20 (169)	-	1.27 (174)	-
$\alpha 2\text{NC1}_{\alpha 121}$	0.84 (219)	0.45 (224)	-	1.22 (168)	0.88 (219)	1.25 (175)	0.79 (219)
$\alpha 1\text{NC1}_{\text{homo}}$	0.36 (226)	-	-	-	0.53 (225)	-	0.41 (226)
$\alpha 2\text{NC1}_{\text{homo}}$	-	-	-	-	1.45 (167)	1.40 (164)	1.46 (167)
$\alpha 3\text{NC1}_{\text{homo}}$	-	-	-	-	-	1.26 (179)	0.58 (225)
$\alpha 4\text{NC1}_{\text{homo}}$	-	-	-	-	-	-	1.23 (178)
$\alpha 5\text{NC1}_{\text{homo}}$	-	-	-	-	-	-	-

*From human placenta, PDB file 1LI1.

Table S5 Missense mutations found in Alport's syndrome patients in α 5NC1, α 3NC1 and α 4NC1. α 5NC1 mutations have been extracted from The Alport syndrome COL4A5 variant database (www.arup.utah.edu/database/ALPORT/ALPORT_welcome.php).

The mutations in chains α 3NC1 and α 4NC1 were reported in the publications given as footnotes to the Table.

Protein Change	Protein residue	α NC1
p.Cys1476Phe	C20F	α 5NC1
p.Thr1480Arg	T24R	α 5NC1
p.Gly1486Ala	G30A	α 5NC1
p.Ser1488Phe	S32F	α 5NC1
p.Ala1498Asp	A42D	α 5NC1
p.Arg1511His	R55H	α 5NC1
p.Pro1517Thr	P61T	α 5NC1
p.Trp1538Ser	W82S	α 5NC1
p.Pro1559Ala	P103A	α 5NC1
p.Arg1563Gln	R107Q	α 5NC1
p.Cys1564Arg	C108R	α 5NC1
p.Cys1564Ser	C108S	α 5NC1
p.Cys1567Arg	C111R	α 5NC1
p.CYS1567Ser	C111R	α 5NC1
p.Glu1568Gln	E112Q	α 5NC1
p.Pro1584Leu	P128L	α 5NC1
p.Cys1586Arg	C130R	α 5NC1
p.Cys1586Phe	C130F	α 5NC1
p.Gly1589Val	G133V	α 5NC1
p.Trp1590Gly	W134G	α 5NC1
p.Gly1596Asp	G140D	α 5NC1
p.Tyr1597Cys	Y141C	α 5NC1
p.Met1601Ile	M145I	α 5NC1
p.Leu1621Ser	L165S	α 5NC1
p.Cys1632Gly	C176G	α 5NC1
p.Cys1632Trp	C176W	α 5NC1
p.Cys1638Tyr	C182Y	α 5NC1
p.Leu1649Arg	L193R	α 5NC1
p.Ser1659Asn	S203N	α 5NC1
p.Arg1677Gln	R221Q	α 5NC1
p.Arg1677Pro	R221P	α 5NC1
p.Arg1677Leu	R221L	α 5NC1
p.Cys1678Arg	C222R	α 5NC1
p.Cys1678Trp	C222W	α 5NC1
p.Gln1679Pro	Q223P	α 5NC1
p.Cys1681Phe	C225F	α 5NC1
p.Gly1508Ser ¹	G48S	α 4NC1
p.Cys1513Thr ¹	C53T	α 4NC1
p.Cys1588Thr ¹	C128T	α 4NC1
p.Leu1474Pro ²	L34P	α 3NC1
p.Leu1474Cys ³	L34C	α 3NC1

p.Phe1475Ser ⁴	F35S	α 3NC1
p.Cys1511Ile ⁵	C71I	α 3NC1
p.Cys1548Tyr ⁴	C108Y	α 3NC1
p.Val1550Gly ⁴	V110G	α 3NC1
p.Trp1578Ser ⁶	W138S	α 3NC1
p.Qln1495Arg ⁶	Q156R	α 3NC1

¹ Storey, H., Savige, J., Sivakumar, V., Abbs, S., Flinter, F. A. (2013). J Am Soc Nephrol 24, 1945-1954

² Lemmink, H. H., Mochizuki, T., van den Heuvel, L. P., Schröder, C. H., Barrientos, A., Monnens, L. A., van Oost, B. A., Brunner, H. G., Reeders, S. T., Smeets, H. J. (1994). Hum Mol Genet 3, 1269-1273.

³ Mochizuki, T., Lemmink, H. H., Mariyama, M., Antignac, C., Gubler, M. C., Pirson, Y., Verellen-Dumoulin, C., Chan, B., Schröder, C. H., Smeets, H. J., Reeders, S. T. (1994). Nat Genet 8, 77-81.

⁴ Tazón Vega, B., Badenas, C., Ars, E., Lens, X., Milà, M., Darnell, A., Torra, R. (2003). Am J Kidney Dis 42, 952-959.

⁵ Heidet, L., Arrondel, C., Forestier, L., Cohen-Solal, L., Mollet, G., Gutierrez, B., Stavrou, C., Gubler, M. C., Antignac, C. (2001). J Am Soc Nephrol 12, 97-106.

⁶ Voskarides, K., Damianou, L., Neocleous, V., Zouvani, I., Christodoulidou, S., Hadjiconstantinou, V., Ioannou, K., Athanasiou, Y., Patsias, C., Alexopoulos, E., Pierides, A., Kyriacou, K., Deltas, C. (2007). J Am Soc Nephrol 18, 3004-3016

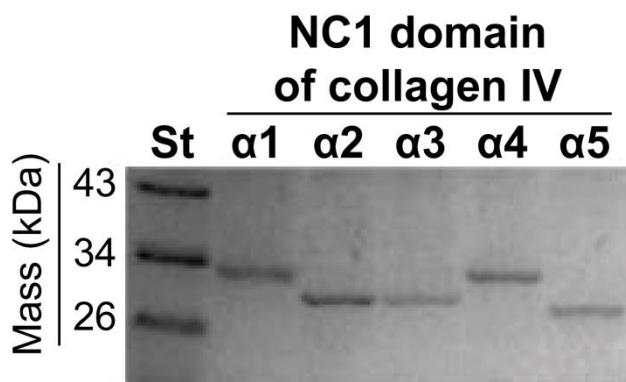


Figure S1 Purified human α 1-5NC1 domains produced by recombinant techniques. Coomassie-stained SDS-PAGE (12% polyacrylamide gel). St, protein standards

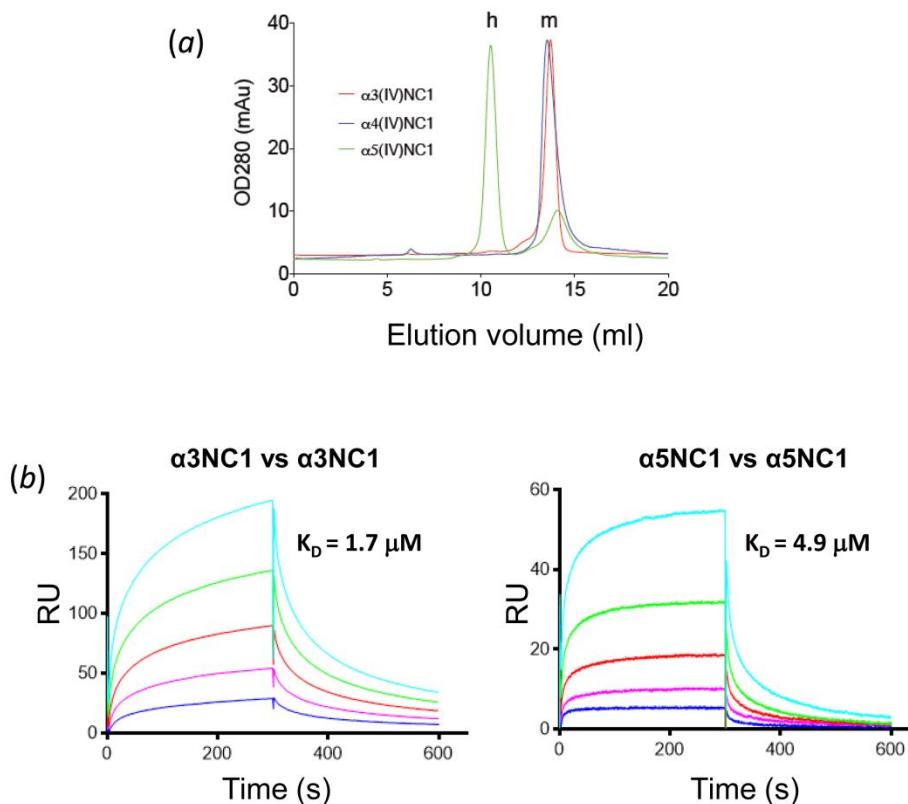


Figure S2 *In vitro* evidence of self-association capacity of α 3NC1 and α 5NC1. (a) Gel filtration chromatography of α 3NC1, α 4NC1 and α 5NC1. "h" and "m" stand for hexamer and monomer, respectively. (b) Surface Plasmon Resonance assays. α 3NC1 (left) or α 5NC1 (right) covalently bound to the sensorchip were used in these assays; the ligands were the α 3NC1 (left) or α 5NC1 (right) chains in solution at concentrations (profiles from bottom to top) of 0.25, 0.5, 1, 2 and 4 μ M. The figure depicts examples of sensorchip traces after subtraction of blanks obtained simultaneously with protein-free solutions. K_D values were estimated from the obtained signal level, (average of two assays at least). For further details, see Materials and Methods.

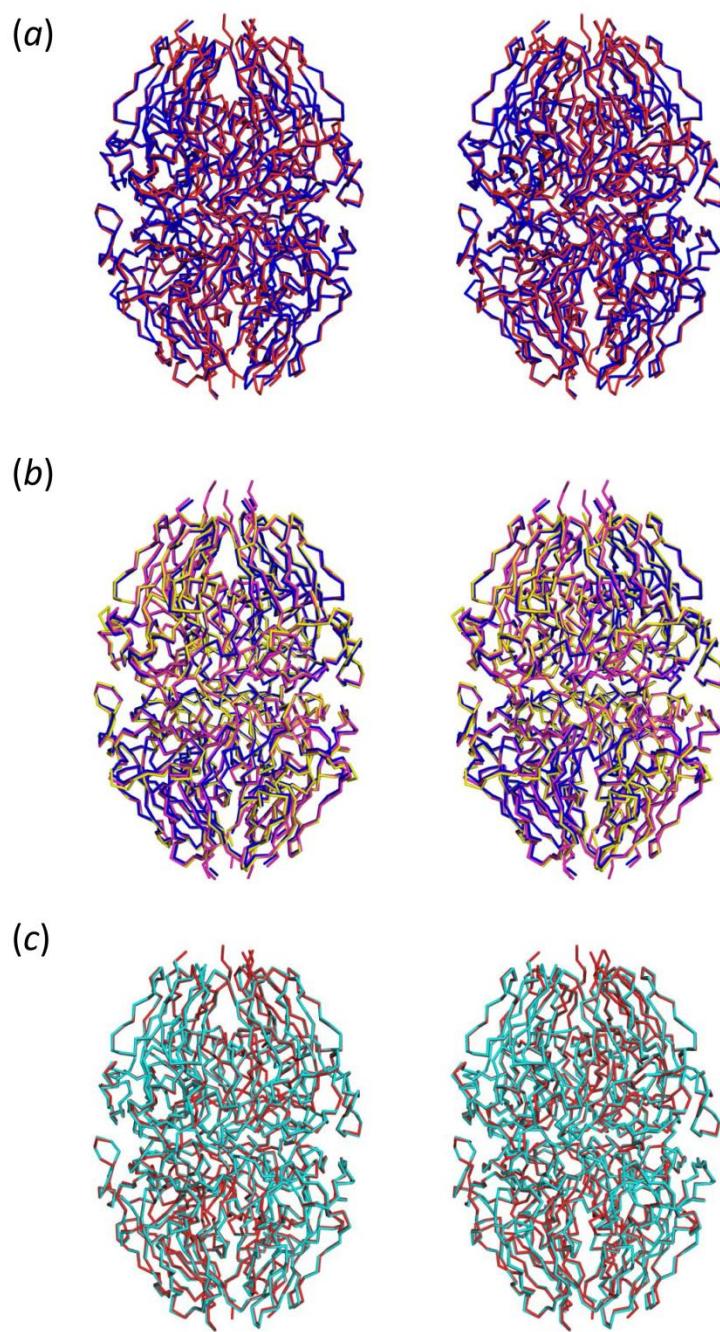


Figure S3 Stereo views of the superimpositions of the backbone of NC1 structures. (a) $\alpha 121\text{NC}1$ hexamer obtained from human placenta (PDB file 1LI1) (red) superimposed over $\alpha 5\text{NC}1_{\text{homo}}$. (b) $\alpha 1\text{NC}1_{\text{homo}}$ (magenta), $\alpha 3\text{NC}1_{\text{homo}}$ (yellow) and $\alpha 5\text{NC}1_{\text{homo}}$ (blue) hexamers. (c) $\alpha 121\text{NC}1$ hexamers obtained from placenta (red) or produced by recombinant techniques in this study (cyan).

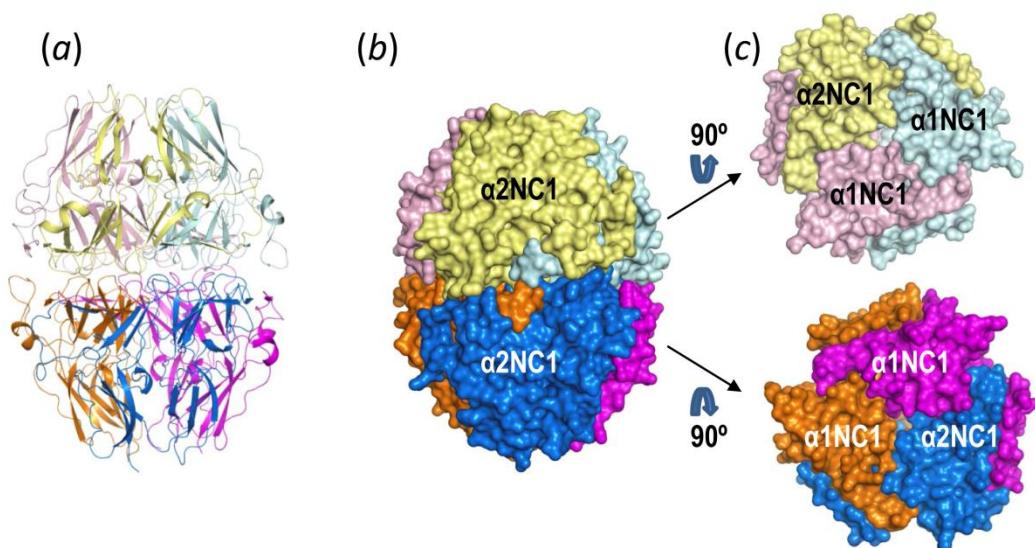


Figure S4 Structure of recombinant α 121NC1 found in the asymmetric unit of its crystal: (a) cartoon representation or (b) surface representation, with each subunit showed with a different colour. In (c) the hexamer is deconstructed in its two protomers, viewed from their equatorial surfaces through which both trimers interact.

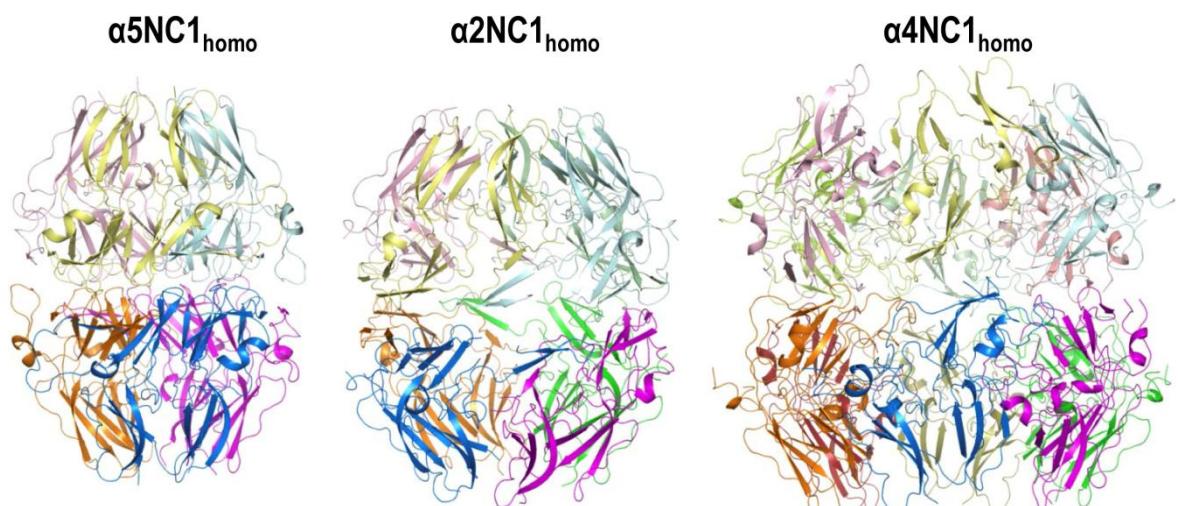


Figure S5

Cartoon representation of the oligomer quaternary structures for the indicated NC1 domains.

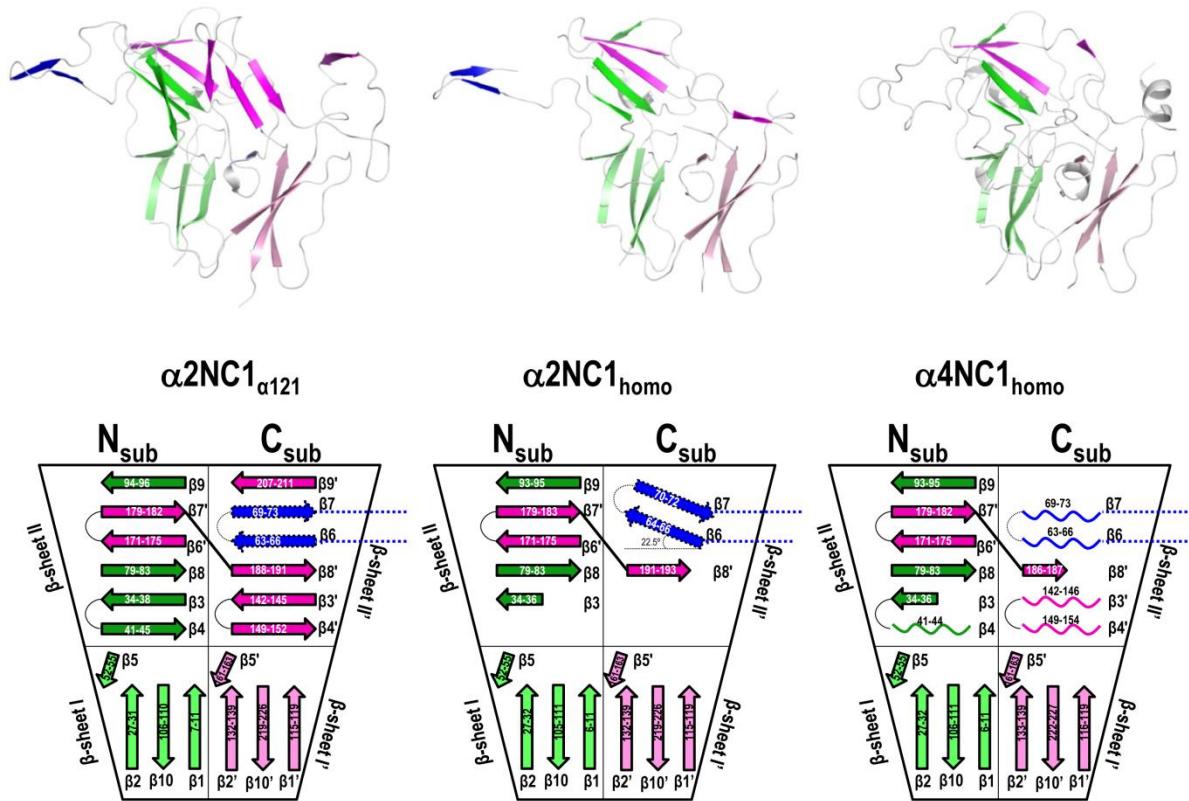


Figure S6 Cartoon representation of the $\alpha 2\text{NC1}$ subunit in the $\alpha 121\text{NC1}$ hexamer ($\alpha 2\text{NC1}_{\alpha 121}$) and the $\alpha 2\text{NC1}$ and $\alpha 4\text{NC1}$ domains in the corresponding crystalline homo-oligomers. Below, schemes of the corresponding folds can be seen with the same colour code as in the cartoon representations and labelling of β strands as in Fig 1.

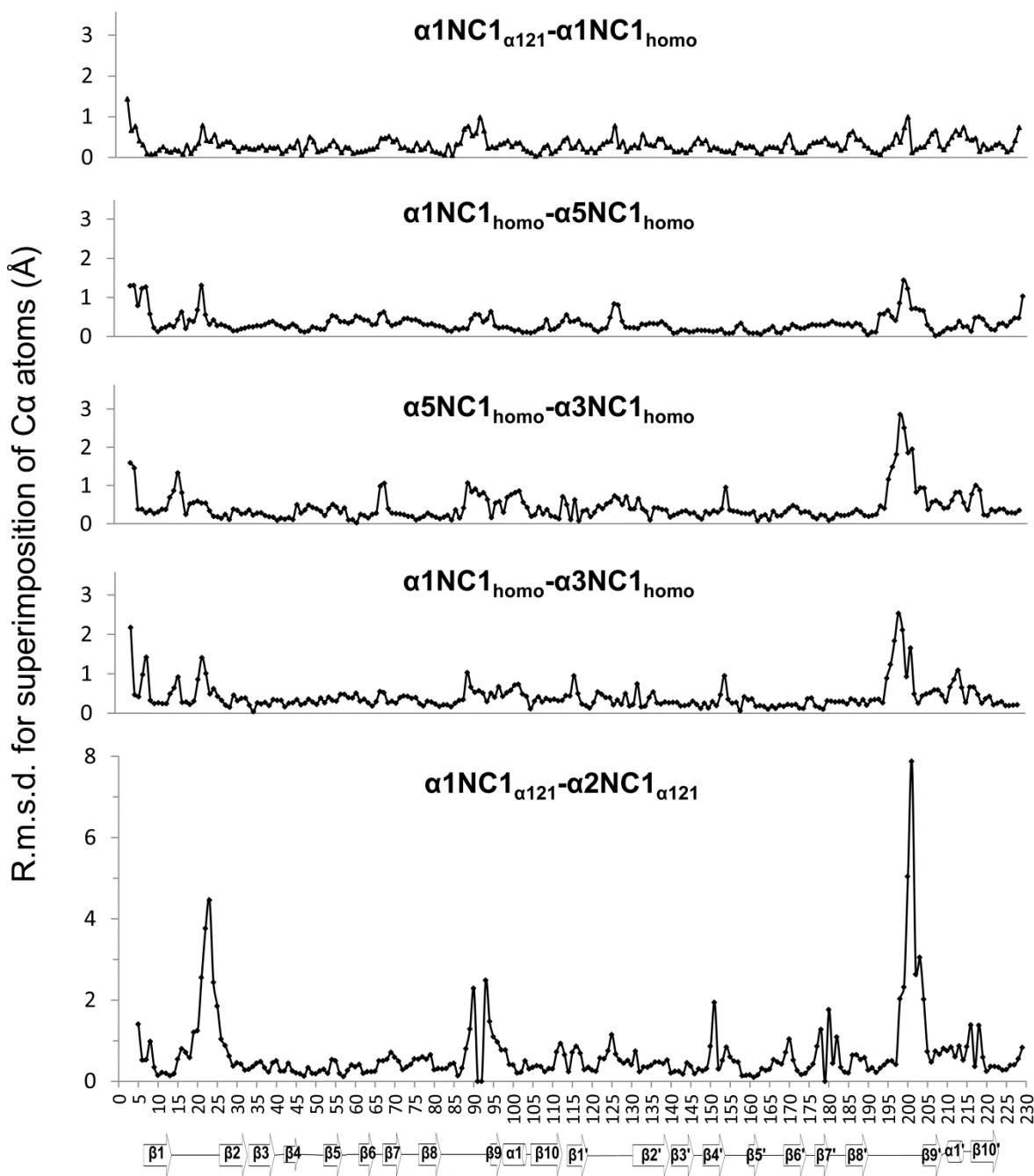


Figure S7 RMSD value for each C α atom after superposition of the α NC1chains. The residue number and secondary structure of the chains are shown in the X-axis at the bottom.

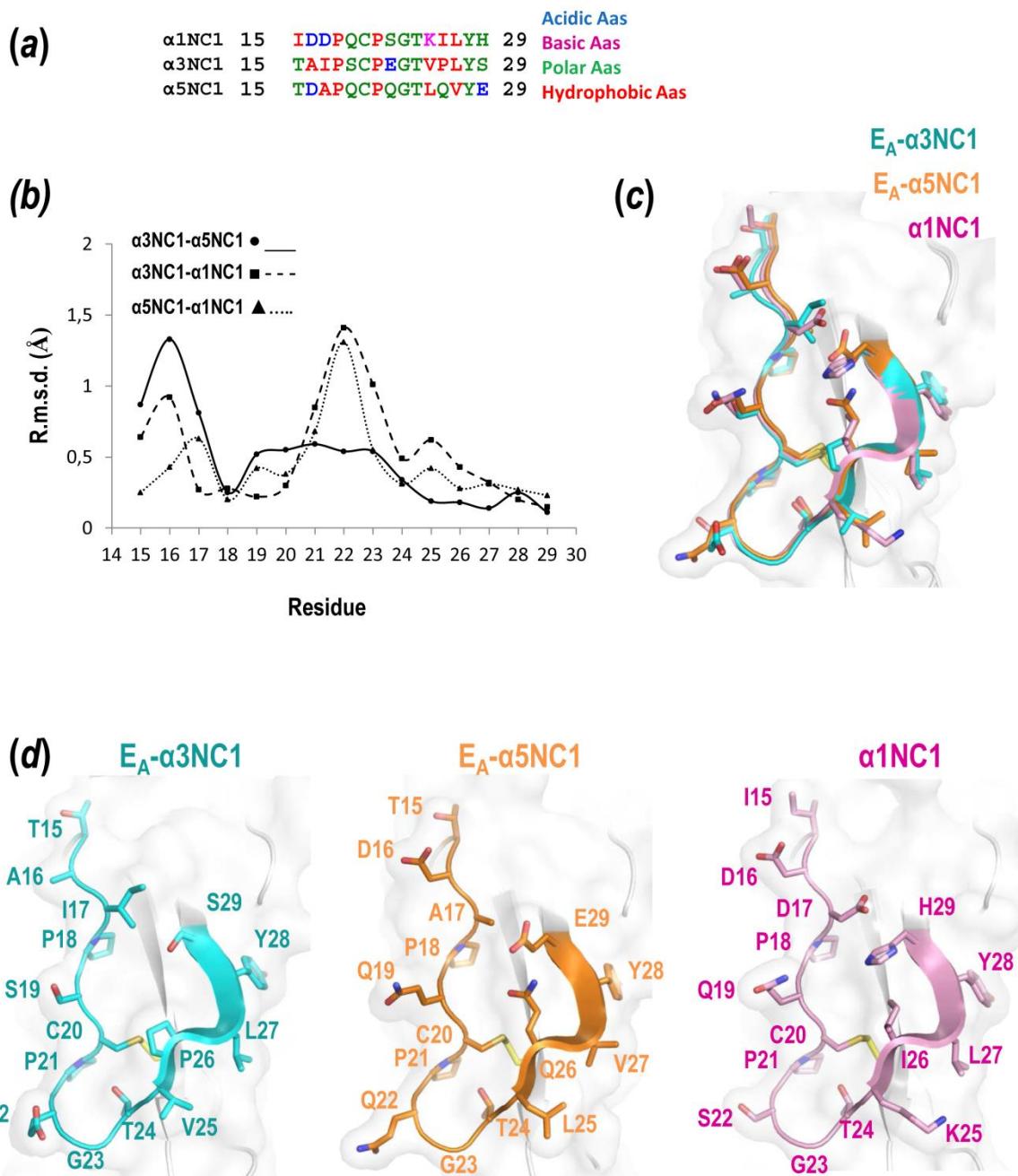


Figure S8 The Goodpasture's E_A epitope. (a) Sequence alignment of $\alpha 3NC1$ and $\alpha 5NC1$ (residues 15–29 of the present structures) E_A epitopes with the corresponding residues of $\alpha 1NC1$ (b) the RMSD values for the superimpositions of the structures corresponding to the sequences aligned in a (all from NC1 domains in homohexamers). (c) and (d) Cartoon representation of the structural superposition and their individual structures, respectively.

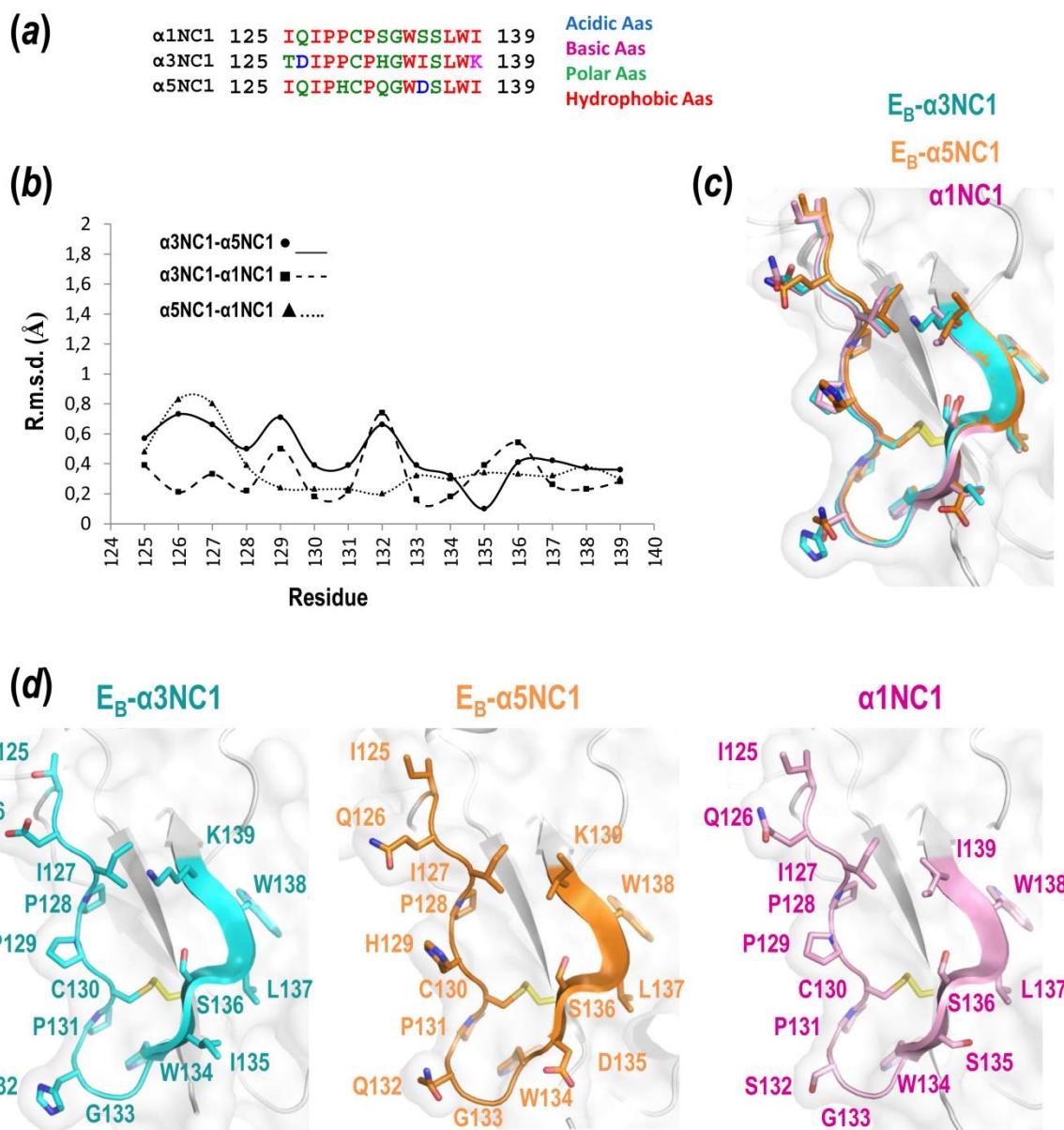


Figure S9 The Goodpasture's E_B epitope. (a) Sequence alignment of $\alpha 3NC1$ and $\alpha 5NC1$ (residues 125-139 of the present structures) E_B epitopes, with the corresponding residues of $\alpha 1NC1$ the RMSD values for the superimpositions of the structures corresponding to the sequences aligned in a (all from NC1 domains in homohexamers). (c) and (d) Cartoon representation of the structural superposition and their individual structures, respectively.

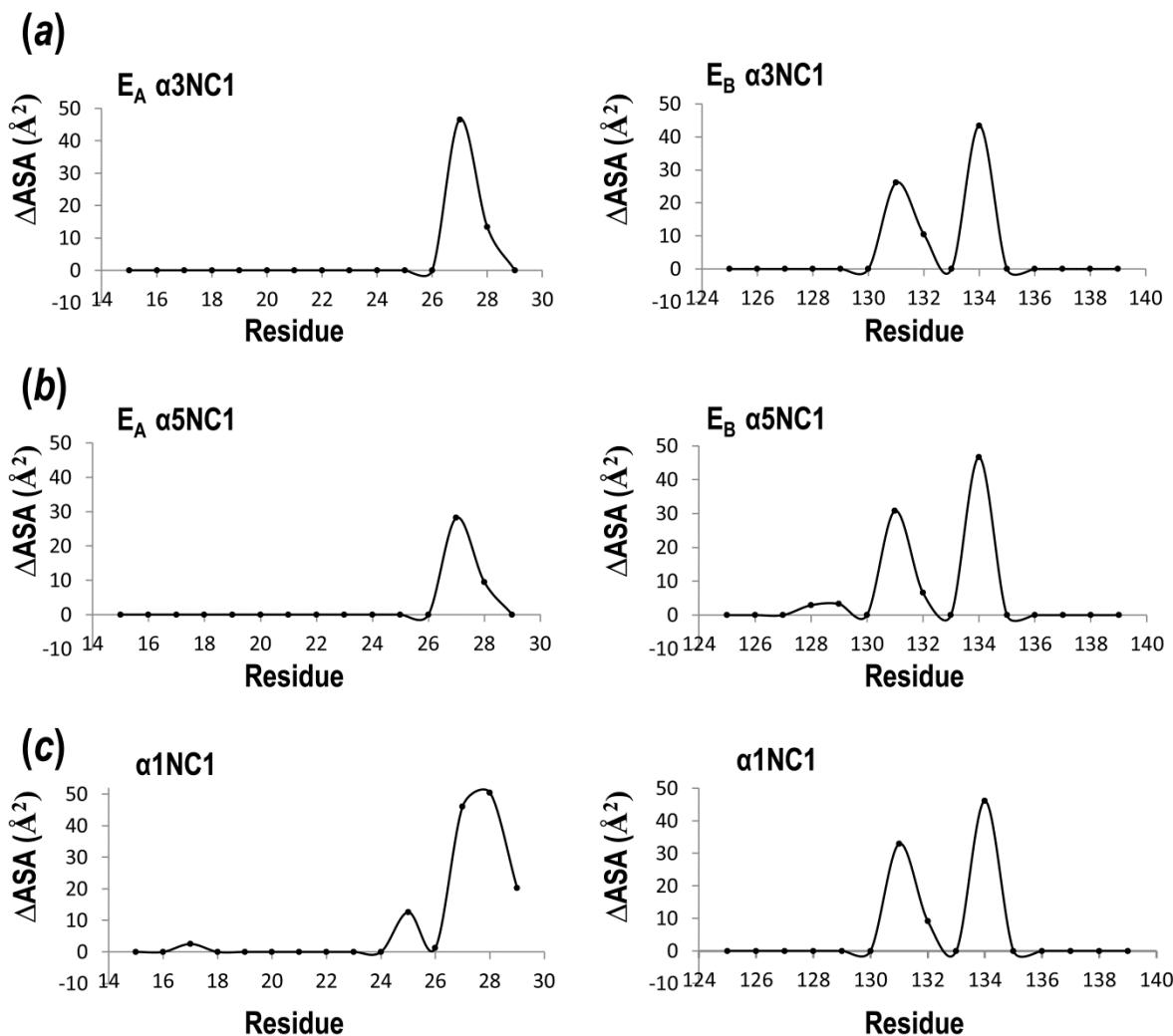


Figure S10 Accessible surface area difference between monomer and homohexamer (ΔASA) for the E_A (left) and E_B (right) epitopes of (a) α 3NC1 and (b) α 5NC1 and, for comparison, the corresponding region (c) of α 1NC1.

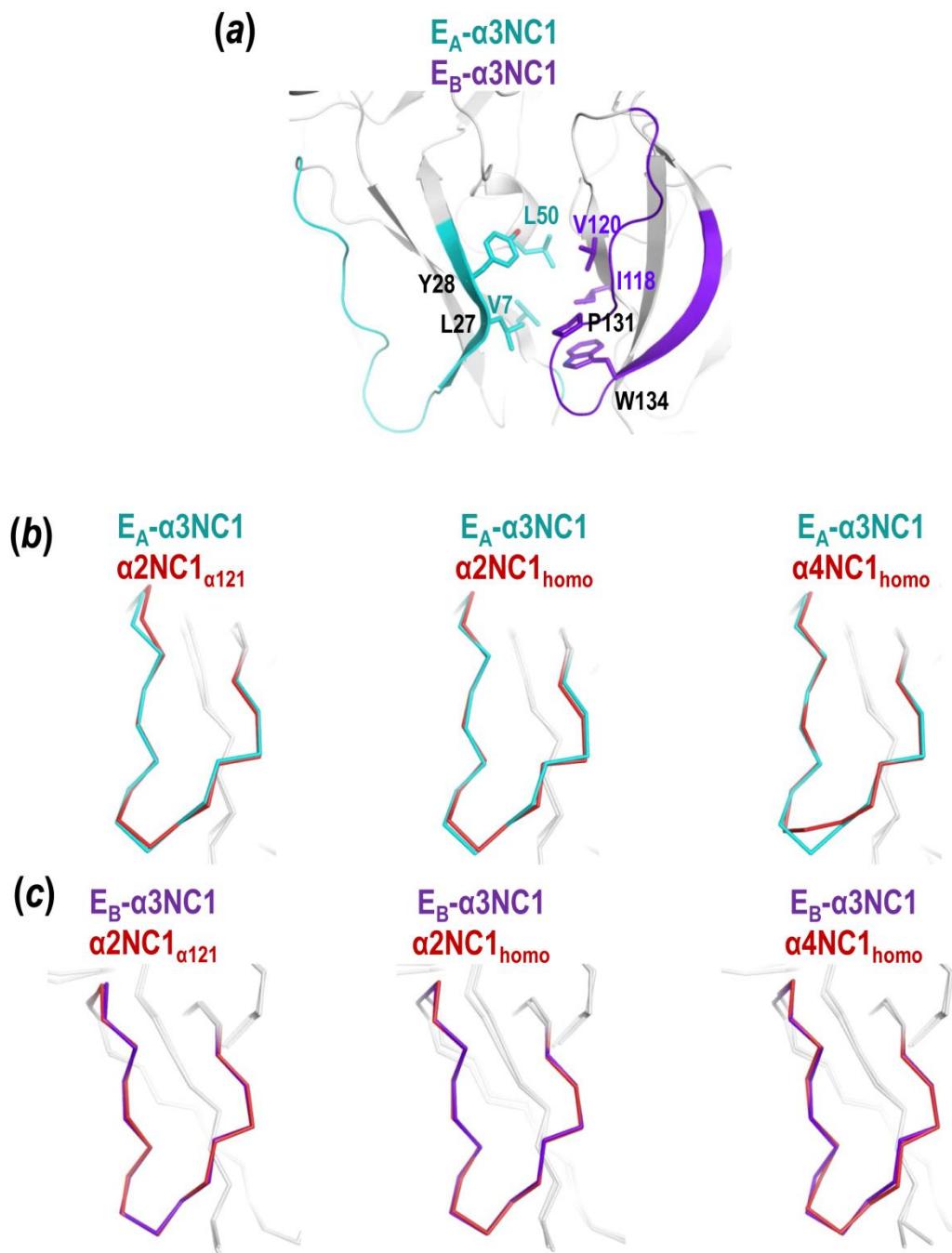


Figure S11 Hydrophobic patch gluing of E_A and E_B epitopes in the $\alpha 3\text{NC}1$ domain structure (a), and illustration of high structural similarity of the two epitopes of $\alpha 3\text{NC}1$ with the corresponding non-autoimmunogenic regions of $\alpha 2\text{NC}1_{\alpha 121}$, $\alpha 2\text{NC}1_{\text{homo}}$ and $\alpha 4\text{NC}1_{\text{homo}}$ (b, c).