# Science Advances

advances.sciencemag.org/cgi/content/full/4/11/eaau2104/DC1

### Supplementary Materials for

#### Dynamic assembly of liquid crystalline graphene oxide gel fibers for ion transport

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Published 2 November 2018, *Sci. Adv.* **4**, eaau2104 (2018) DOI: 10.1126/sciadv.aau2104

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Table S5. Comparison of ionic conductivity of GO gel fibers with various nanosheet films. References (39–50)

#### Other Supplementary Material for this manuscript includes the following:

(available at advances.sciencemag.org/cgi/content/full/4/11/eaau2104/DC1)

Movie S1 (.mp4 format). In situ observation of dynamic assembly of GO gel fiber at  $v_2 = v_1$ . Movie S2 (.mp4 format). In situ observation of dynamic assembly of GO gel fiber at  $v_2 = 3v_1$ .



**Fig. S1. Characterization of GOLC dispersion.** (**a**) A typical SEM image of GO sheets. (**b**) Size distribution of the GO sheets. (**c**) A typical atomic force microscopy (AFM) image of GO sheet. The scale bar (I) denotes 1.6 nm of height. From the uniform thickness and average lateral size (1.7  $\mu$ m), the aspect ratio of GO sheet was calculated to be  $1.07 \times 10^3$  and is in the range of previously reported values (700-2600) of GO with high liquid crystallinity (*14*, *39*). (**d**) A digital photo of typical GOLC dispersion (0.5 wt%) used for the gel-fiber assembly. (**e**-**f**) POM images of GOLC dispersion with 0.5 and 1.0 wt%, respectively.



**Fig. S2. Analysis of velocity profile.** (a) Expanded POM image of 1<sup>st</sup> one in **Fig. 1c** showing two dotted lines; *A*-*A*' (inside the nozzle) and *B*-*B*' (in the ejected fluid). (b) Experimental and fitted flow velocity of GOLC fluid inside the nozzle as a function of distance showing a parabolic velocity profile. (c) Experimental velocity profile of ejected fluid at  $v_2 = v_1$ .

**Note:** The parabolic velocity profile in (b) was fitted with the following equation:  $v_0(r) = v_{0, \text{ max}}$  $(1 - r^2/R^2)$ , where  $v_0(r)$  is the velocity of the GOLC fluid depending on radial location (r) in the nozzle. *R* and  $v_{0, \text{ max}}$  are the radius of the nozzle (200 µm) and the maximum velocity (32 mm s<sup>-1</sup>) of the fluid at r = 0.



Fig. S3. Estimation of time scale for  $Ca^{2+}$  diffusion. (a) The estimated concentration of  $Ca^{2+}$ (*C*) as a function of radial distance ( $\rho$ ) for the diffusion into the ejected fluid at  $v_2 = v_1$ . (b) The estimated concentration at the core region ( $0 \ \mu m \le \rho \le 50 \ \mu m$ ). The critical gelation concentration (CGC) of  $Ca^{2+}$  (9 mM) is highlighted with a red dotted line. (c) 1<sup>st</sup> POM image in Fig. 1c showing the elapsed time after ejection at each axial position (*z*).

**Note**: The diffusion time scale of  $Ca^{2+}(t)$  was estimated as solving the diffusion equation in cylindrical coordinates

$$\frac{\partial C}{\partial t} = \frac{1}{\rho} \frac{\partial}{\partial \rho} \left( \rho D \frac{\partial C}{\partial \rho} \right) \tag{1}$$

where *D*, *C*, and  $\rho$  are the diffusion coefficient of Ca<sup>2+</sup> (0.79 × 10<sup>-5</sup> cm<sup>2</sup> s<sup>-1</sup>), the concentration of Ca<sup>2+</sup>, and radial distance from the center of ejected fluid, respectively. In our system, the cations (Ca<sup>2+</sup>) diffuse from near the surface ( $\rho_{surface} = 230 \mu m$ ;  $C_{surface} = 450 mM$ ) to the center ( $\rho_{center} = 0 \mu m$ ;  $C_{center} = 0 M$ ) by the large concentration gradient (1<sup>st</sup> image in **Fig. 1c**). Thus, the boundary conditions are as follows

$$C(\rho_{\text{surface}}, t) = C_{\text{surface}} \tag{2}$$

$$\frac{\partial C}{\partial \rho} \left( \rho_{center}, t \right) = 0 \tag{3}$$

We plotted the estimated *C* as a function of  $\rho$  (**Fig. S3a** and **b**). The critical concentration of Ca<sup>2+</sup> for the gelation of GO was chosen to 9 mM based on a previous report (40). As shown in **fig. S3b**, the time required for fully gelling the ejected fluid was estimated to be ~1.5 s.



Fig. S4. Diameter and extension rate of ejected fluid at  $v_2 = 5.5v_1$  as a function of distance.



**Fig. S5. POM images and birefringence intensity profiles of gel fibers.** (a-b) POM images of a (a)  $3v_1$ -GO gel fiber and (b)  $4v_1$ -GO gel fiber, respectively. (c) Birefringence intensity profile along the red lines in the glass nozzle for (a) and (b). (d) Normalized birefringence intensity profile along the yellow lines for (a) and (b). The dashed lines in (d) denote mean values.



**Fig. S6. Sol-gel transition and gel behavior of GO solution.** (a) Series of photographs showing the sol-gel transition of GO dispersion started from the membrane interface and propagated normal to the membrane plane. Finally, GO dispersion was fully converted to a gel after 12 h. (b) Casson plots of GO gels prepared with different amount of the added NH<sub>4</sub>OH in 5 wt% CaCl<sub>2</sub> coagulation solution.



**Fig. S7. Zeta potential of GO as a function of pH in aqueous dispersion at a concentration of 0.05 mg ml**<sup>-1</sup>**.** The red and blue arrows denote the pH values of CaCl<sub>2</sub>-only (9.5) and modified (11) coagulation solution.



**Fig. S8. XPS spectra of GO gels.** High resolution C 1*s* spectra in XPS of cylindrical gels and gel fibers coagulated in (**a**) modified and (**b**) CaCl<sub>2</sub>-only coagulation solution, respectively.



Fig. S9. Raman spectra of GO gels.



Fig. S10. Determination of the degree of orientation in gel fibers. (a-c) SAXS patterns of  $4v_1$ and  $v_1$ -GO gel fibers, respectively. (b-d) Corresponding azimuthal scans used to determine the degree of orientation in the gel fibers.



Fig. S11. Stress-strain curves of dried GO fibers under 10% min<sup>-1</sup> of tensile strain.



**Fig. S12. Determination of the degree of orientation in dried fibers.** (a-b) 2D SAXS patterns of (a)  $4v_1$ - and (b)  $v_1$ -GO fibers, respectively. *f* denotes Herman's orientation function.



Fig. S13. *I-V* curves of NICs of different lengths.



**Fig. S14. Experimental setup for measuring the ionic conductivity of NICs.** (a) A digital photo of a typical PDMS cell, consisting of both source and drain reservoirs with a 10 mm-length gap between them. (b) A magnified photo of the black box in (a) showing a GO gel fiber bridging the two reservoirs. (c-d) The measurement cell was sealed in a glass container and connected to a potentiostat using Ag/AgCl electrodes. Photo credit: Ki Hyun Lee, Hanyang University, Dept. of Organic and Nano Engineering.



**Fig. S15. Removal of Ca<sup>2+</sup> by ion exchange in GO gel fibers.** (a-b) High resolution Ca 2p and K 2p curves of GO gel fibers in XPS before and after KCl treatment, respectively, suggesting removal of Ca<sup>2+</sup> and exchange for K<sup>+</sup>.



Fig. S16. Modeling of GO gel fiber nanochannels using the configuration of GO films. (a) A typical SEM image of the cross-sectional view of a dried GO gel fiber. (b) Magnified image of the white box in (a). Dotted lines represent some curved 2D nanochannels in the GO gel fiber. (c) XRD patterns of GO gel fibers, indicating an interlayer spacing of 1.38 nm after rehydration in KCl solution. (d) Schematic illustration of re-assembly of nanochannels confined in a cylindrical fiber with radius (r); this was converted into a rectangular parallelepiped as is performed for GO films.

	Original GO gel	Modified GO gel				
	Original OO ger	0.05 M NH4OH	0.25 M NH4OH	1.00 M NH4OH		
Yield stress [Pa]	181	222	308	346		

#### Table S1. Yield stress values of GO gels prepared with different amounts of NH4OH.

Table S2. Oua	ntitative XPS	analyses	of cy	vlindrical	gels and	gel fibers.
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Sample	Coagulation solution	C [at%]	O [at%]	Ca [at%]	O/C ratio	Ca/C ratio
Cylindrical gel	CaCl <sub>2</sub> only	65.9	33.0	1.1	0.50	0.016
	CaCl <sub>2</sub> , NH <sub>4</sub> OH	68.1	29.6	2.3	0.43	0.034
Gel fiber	CaCl <sub>2</sub> only	65.2	33.9	0.9	0.52	0.014
	CaCl <sub>2</sub> , NH <sub>4</sub> OH	69.6	27.6	2.8	0.40	0.040

## Table S3. Comparison of mechanical properties of our GO fibers with previously reportedGO fibers and other nanocarbon-based fibers.

Sample	Young's modulus [GPa]	Tensile strength [MPa]	Elongation at break [%]	Draw ratio	Degree of orientation	Method	Note
$v_1$ -GO fiber	27.6 ± 7.3	$152.3 \pm 36.0$	$0.7 \pm 0.2$	1	0.82	Wet spinning	this
$4v_1$ -GO fiber	$38.3\pm1.7$	$301.4\pm52.9$	$0.8\pm0.1$	4	0.93	Wet spinning	work
GO fiber with large GO (18.5 µm of mean size)	6.3	364.4	6.8	1.3	-	Wet spinning	Ref. 42
GO fiber with large GO $(56 \pm 20 \ \mu m \text{ of mean size})$	12.1	239.3	2.7	1.2	-	Wet spinning	Ref. 43
Knittable GO fiber	7.9	135.8	5.9	-	-	Dry-jet wet spinning	Ref. 44
Double-walled carbon nanotube (DWCNT) fiber	8.3	299	5	-	-	Drawing-drying process	Ref. 45
Nanocelluose fiber	$21.3\pm5.5$	$296\pm71$	$2.8\pm0.5$	1	0.83	Wet spinning	Ref. 46
Nanocelluose fiber*	$17.6\pm0.7$	$490\pm86$	$6.4 \pm 1.6$	1.15	$0.50\pm0.01$	Hydrodynamic assembly	Ref. 18
Nanocelluose fiber	86	1570	~3	-	0.70	Hydrodynamic assembly	Ref. 47
Graphene nanoribbon fiber (annealed at 1300 °C)	$15.3\pm1.0$	$151.7\pm20.7$	$1.02\pm0.21$	-	0.53	Dry-jet wet spinning	Ref. 48

\*It should be noted that the diffusion time scale in Ref. 16 is 3.3 s.

Sample	Diameter [µm]	Linear density [Tex]	Apparent volume fraction [%]	Apparent porosity [%]	Apparent density [g cm <sup>-3</sup> ]
v <sub>1</sub> -GO fiber	26.8 ± 1.4	$0.70\pm0.07$	69	31	1.24
4v <sub>1</sub> -GO fiber	$14.3\pm0.5$	$0.23\pm0.01$	80	20	1.43

Table S4. Structural parameters of dried GO fibers.

**Note**: The apparent volume fraction, porosity, and density were calculated using previously reported density (1.8 g cm<sup>-3</sup>) of GO (41) by assuming the geometry of fiber is a cylinder.

Table S5.	Comparison	of ionic co	onductivity	of GO ge	el fibers	with	various	nanosheet fi	ilms.
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Sample	Building block	Electrolyte	Ionic conductivity [S cm <sup>-1</sup> ]	Note	
v <sub>1</sub> -GO gel fiber	GO	< 3 mM KCl	$1.7 imes10^{-4}$	this work	
$4v_1$ -GO gel fiber	GO	< 7 mM KCl	$4.2  imes 10^{-4}$	uns work	
GO film	GO	< 10 mM KCl	$\sim 1.0 \times 10^{-3}$	Ref. 7	
Boron nitride (BN) film	BN sheets	< 0.1 mM KCl	$\sim 1.0 \times 10^{-4}$	Ref. 49	
Vermiculite film	Vermiculite sheet	< 0.1 M HCl	$6.0 imes10^{-3}$	Ref. 50	