

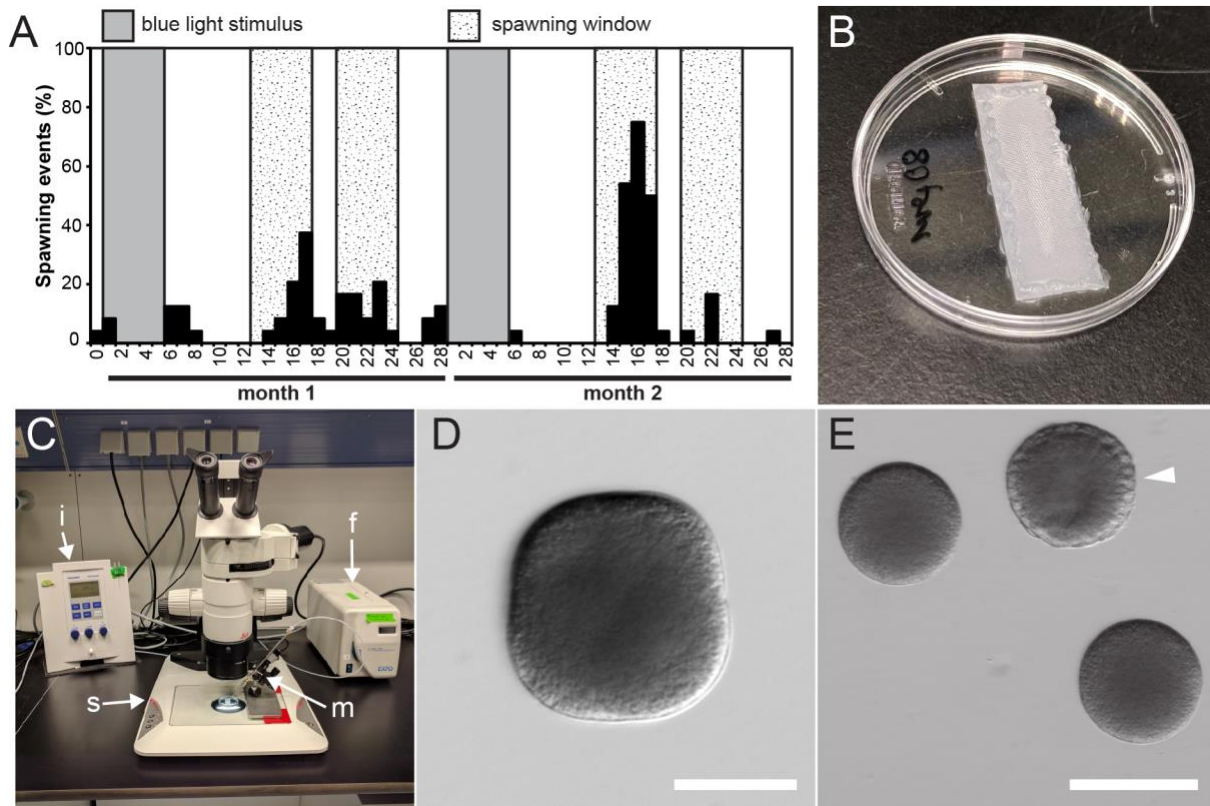
Jones et al.
Supplementary Information

Supplementary Information

Microinjection to deliver protein, mRNA, and DNA into zygotes of the cnidarian endosymbiosis model *Aiptasia* sp.

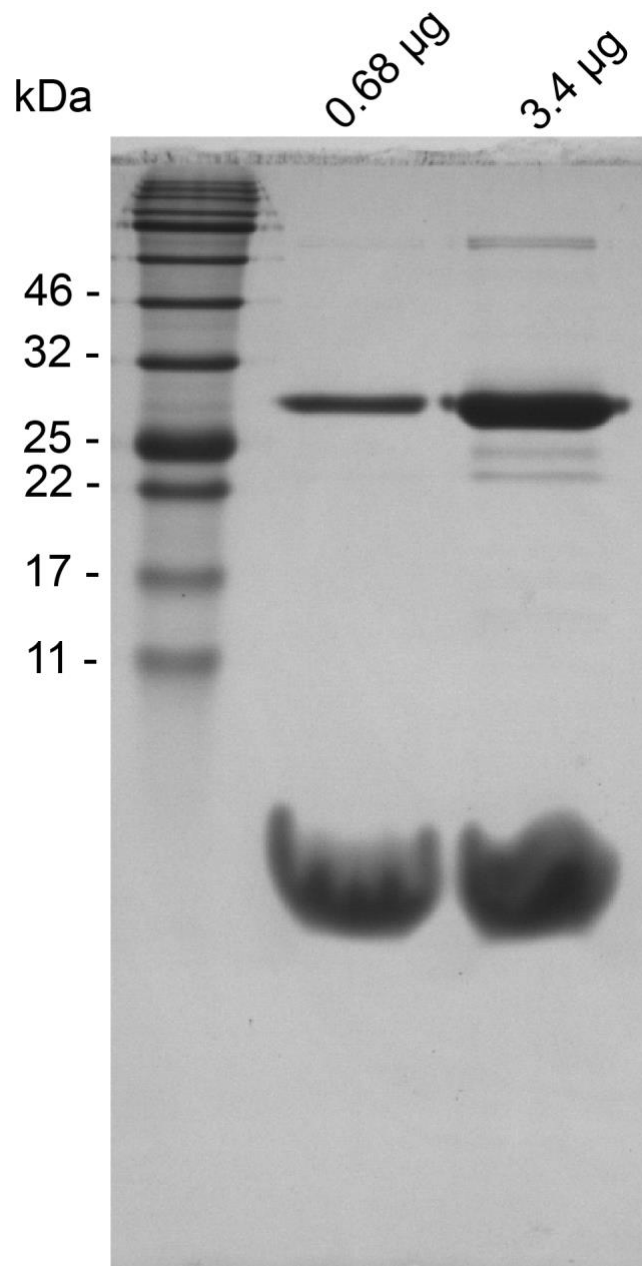
Victor A. S. Jones, Madeline Bucher, Elizabeth A. Hambleton, Annika Guse

Figure S1



(A) Spawning induction efficiency is demonstrated by the percentage of female tanks containing spawned eggs on each day of two consecutive artificial lunar cycles. Duration of the blue-light stimulus (spawning induction cue) is indicated in grey; the expected spawning period is indicated with a speckled background (total of 24 tanks). **(B)** Microinjection dishes are prepared by affixing a strip of 80 x 80 μm nylon mesh on the bottom of a small petri dish lid using silicon grease around the edges. **(C)** Microinjection set-up with stereomicroscope (s), fluorescence lamp (f), micromanipulator (m), injection pedal (not shown), and FemtoJet 4i injector (i). **(D)** The injection session proceeds until most zygotes assume a box-like shape, indicating imminent cleavage to 4-cell stage. Scale = 100 μm . **(E)** Developing embryos (arrowhead) and unfertilized eggs can be clearly differentiated 4-5 h post-fertilization (hpf). Scale = 50 μm .

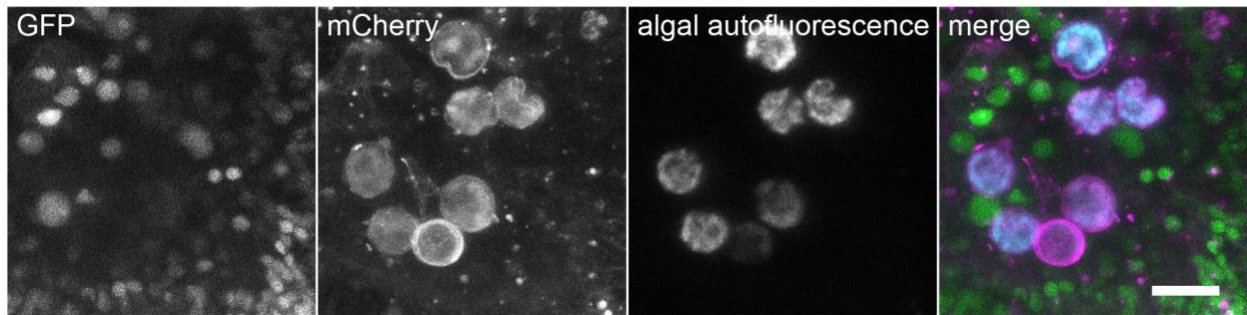
Figure S2



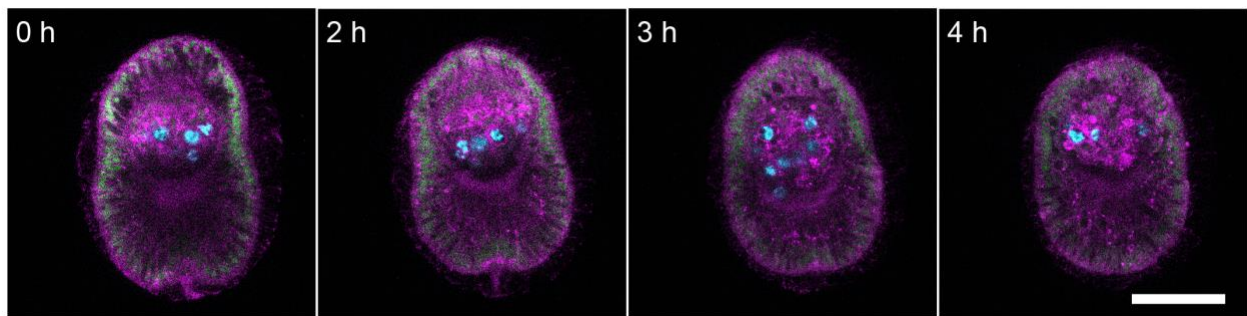
Recombinant Lifect-eGFP protein (30.6 kDa) resolved with SDS-PAGE and stained with Coomassie Brilliant Blue to visualize. Protein was injected at 3.4 mg/ml into zygotes. Left lane, ladder.

Figure S3

A



B



(A) Within a single *Aiptasia* larva, the strength of symbiosome labeling by farnesylated mCherry (magenta) and algal far-red autofluorescence (cyan) are variable. Larva was injected with *NLS-eGFP-2A-mCherry-CaaX* mRNA and incubated for 2 days with *Symbiodinium* strain SSB01 at 10,000 cells/ml, before fixation at 4 dpf. Scale = 10 μ m. (B) Agarose-embedded larvae can be imaged live for several hours. Larva was injected with *NLS-eGFP-2A-mCherry-CaaX* mRNA and incubated for 1 day with *Symbiodinium* strain SSB01 at 10,000 cells/ml, then at 3 dpf embedded in low-melt agarose and imaged at the shown times. Z-stacks were taken with confocal laser-scanning microscopy once every 30 min. Scale = 50 μ m.

Movie S1

Live imaging of larvae after incubation with *Symbiodinium* strain SSB01 at 100,000 cells/ml for 2 days. Larva was embedded in low-melt agarose and imaged at 4 dpf with DIC and fluorescence microscopy to monitor intracellular symbionts via their natural autofluorescence using a TexasRed filter set. False color representation with 5 ramps LUT. Scale = 100 μ m.

Movie S2

Live imaging of the endoderm of a larva injected with *NLS-eGFP-2A-mCherry-CaaX* mRNA and incubated for 1 day with *Symbiodinium* strain SSB01 at 100,000 cells/ml. Larva was embedded in low-melt agarose and imaged at 3 dpf with confocal laser-scanning microscopy, at a rate of one frame per 2.5 sec. mCherry channel left, grayscale; merge of mCherry (magenta), eGFP (green) and algal red autofluorescence (cyan), right. Scale = 10 μ m.

Supplementary Table S1

Actin gene accession #	Primer direction	Primer sequence 5' – 3'	Amplicon (promoter) length, bp
<i>XM_021049442.1</i>	Forward	CCTCTGGCAACGTAACACCAAC	1479
<i>XM_021049442.1</i>	Reverse	TTTGTGAGTAGTTTTGAATTGAGA	1479
<i>XM_021045858.1</i>	Forward	GGCATCCGTCTCGTCACGATAG	1504
<i>XM_021045858.1</i>	Reverse	TTTAATATATTTTAGAAAGGTCAAACACC	1504
<i>XM_021060755.1</i>	Forward	CTGACATCAGCCGTACTCTAGAAC	1463
<i>XM_021060755.1</i>	Reverse	TGTAAGTTTGAAGTATGAAGTGAA	1463
<i>KXJ15097.1</i>	Forward	TATATTGACGATCCAAGTGTCACA	998
<i>KXJ15097.1</i>	Reverse	GTTGGTTYTAAAATTTACTTACAACGAACA	998