CUL4A^{DET1-COP1} and OTUD7B Regulate Sox2 Proteostasis and Govern Neural Progenitor Cell Differentiation

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Supplementary Figure 1. CUL4A contributes to Sox2 ubiquitylation during neuronal differentiation.

(a) mRNA level of Sox2 and WWP2 was measured during NPCs differentiation by Q-PCR. NPCs (neural stem/progenitor cells derived from human PSCs, WA09) were cultured in NIM (neural induction medium) to undergo cellular differentiation for the indicated times. Results are shown as mean ± sd. Each error bar shows the standard deviation of the value from 3 independent experiment. *, *P*<0.05, **, *P*<0.01 compared with Day 0, Student's t-test. (b) WWP2 protein level was detected by immunoblotting in NPCs and HEK293T (as positive control). (c) WWP2, Set7 and Ube2S protein level was detected by immunoblotting in ESC, NPCs and neurons. (d, e) Increasing amounts of CUL4A (d) or CUL4B (e) were transfected into HEK293T cells and ectopic Flag-Sox2 expression was detected. The representative images are shown from 3 independent experiments. Unprocessed original scans of blots are shown in Supplementary Fig. 9.



Supplementary Figure 2. The CUL4A^{DET1-COP1} interacts with Sox2 and regulates its stability.

(a) The lysates of HEK293T transfected with indicated constructs were subjected to immunoprecipitation with anti-Flag antibody. The immunoprecipitates were then blotted. (b) The purified His-COP1 were incubated respectively with purified GST or GST-Sox2. The mixtures were subjected to GST pulldown and blotted. (c) Diagram of two COP1-recognized VP motif of Sox2. (d) Flag-WWP2 or Flag-COP1 were co-transfected into HEK293T cells with empty vector, Myc-Sox2 WT or Myc-Sox2 AA mutant, respectively. Cell lysates were subjected to immunoprecipation with anti-Flag antibody and immunoblot. The representative images are shown from 3 independent experiments. Unprocessed original scans of blots are shown in Supplementary Fig. 9.



Supplementary Figure 3. COP1 interacts with Sox2 directly.

(a) Cells were transfected with indicated siRNA and western blot was performed to measure expression of Sox2. (b) HEK293T cells were transfected with indicated constructs, and the lysates were immunoblotted with anti-Myc, anti-Flag or anti-His. The representative images are shown from 3 independent experiments. Unprocessed original scans of blots are shown in Supplementary Fig. 9.



Supplementary Figure 4. CUL4A^{DET1-COP1} ubiquitylates Sox2.

(a) HEK293 cells transfected with indicated constructs were treated with MG132 before collection. Myc-Sox2 were immunoprecipated with anti-Myc and immunoblotted with anti-HA. (b) NPCs with stable knockdown of COP1, WWP2 or both of them were generated by lentiviral infection. Cell were treated with 10 µg per ml CHX for indicated time. Then Sox2 was immunoblot with anti-Sox2. (c) NPCs with stable knockdown of COP1, WWP2 or both of them were generated by lentiviral infection. Cells were treated with MG132 for 8h before collection. Cell lysates were immunoprecitated by anti-Sox2 antibody and then blotted by anti-Ub antibody. (d, e) Dual luciferase reporter assay was used to examine the effects of CUL4ADETI-COP1 on the activity of Sox2 in regulating FGF4 transcription. Cells were transfected with indicated constructs. Forty-eight hours after transfection, cell lysates were subjected to dual luciferase reporter assay. The data are normalized to the activity of the cells transfected with empty vectors and presented as the mean \pm sd. (##: P<0.01 compared to the cells transfected only with FGF4-luc; **: P<0.01 compared to the cells transfected with Flag-Sox2 and FGF4-luc). The expression levels of CUL4A, COP1, DET1 and Sox2 were monitored by western blot with antibodies against Flag and HA, respectively. The representative images are shown from 3 independent experiments. Unprocessed original scans of blots are shown in Supplementary Fig. 9.



Supplementary Figure 5. COP1 promotes NPCs differentiation in a Sox2-dependent manner. COP1 (red) levels were detected with immunostaining in NPCs with stable COP1 knockdown (a),

combined knockdown of COP1 and Sox2 (b), as well as combined overexpression of COP1 and Sox2-AA (c). Nuclei were counterstained with HO (blue). Scale bar, 50 μ m. The representative images are shown from 3 independent experiments.



Supplementary Figure 6. OTUD7B deubiquitylates and stabilizes Sox2.

(a) The effect of increased OTUD7B or OTUD7A level on Sox2 expression in cells. Cells were transfected with indicated constructs and western blot was performed to measure expression of Flag-Sox2. (b) Half-life analysis of Sox2 in cells with OTUD7A knockdown. Cells transfected by indicated shRNA were treated with 10 µg per ml CHX, and collected at the indicated times for western blot. (c) Half-life analysis of Sox2 in cells with OTUD7A or OTUD7B overexpression. Cells transfected by indicated constructs were treated with 10 µg/ml CHX, and collected at the indicated times for western blot. Quantification of Sox2 levels relative to tubulin is shown. **, P<0.01, two-way ANOVA test. (d) Cells transfected with indicated constructs were lysed and subjected to western blot with anti-Flag, anti-Myc or anti-His antibody. (e) Myc-Sox2 was cotransfected into HEK293T cells with Flag-OTUD7B or the mutants, and the cell lysates were immunoprecipated with anti-Myc antibody for detection of the interaction of ectopic Sox2 (or Sox2 mutants) and OTUD7B in cells. (f) The interaction of ectopic Sox2 and OTUD7B were detected in HEK293T cells. The lystates of HEK293T cells co-transfected with Myc-Sox2 and Flag-OTUD7B were subjected to immunoprecipitation with anti-Myc antibody and immunoblotted. (g, h) Dual luciferase reporter assay was used to examine the effects of OTUD7B on the activity of Sox2 in regulating FGF4 transcription. Cells were transfected with indicated constructs. Forty-eight hours after transfection, cell lysates were subjected to dual luciferase reporter assay. The data are normalized to the activity of the cells transfected with empty vectors and presented as the mean \pm S.D. (##: P<0.01 compared to the group transfected only with FGF4luc; **: P<0.01 compared to the group transfected with Flag-Sox2 and FGF4-luc). The expression levels of OTUD7B and Sox2 were monitored by westernblot with antibodies against Flag and Myc respectively. The representative images are shown from 3 independent experiments. Unprocessed original scans of blots are shown in Supplementary Fig. 9.



Supplementary Figure 7. OTUD7B maintains NPCs stemness in a Sox2-dependent manner.

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OTUD7B (green) levels were detected with immunostaining in NPCs with stable OTUD7B knockdown (a), combined OTUD7B knockdown with Sox2-AA overexpression (b), as well as combined OTUD7B overexpression with Sox2 knockdown (c). Nuclei were counterstained with HO (blue). Scale bar, 50 µm. The representative images are shown from 3 independent experiments.



Supplementary Figure 8. COP1 and OTUD7B coordinate to control NPCs differentiation.

(a) Quantified data indicated the fraction of cells expressing Sox2, TUJ1, COP1 or OTUD7B during NPCs differentiation. Data are means \pm sd. Each error bar shows the standard deviation of numbers of positive cells in 5 fields of view. **, P<0.01, ***, P<0.001. Student's t-test. (b) Cell transfected with indicated constructs were treated with or without MG132 for 8 h. Sox2 level was measured by immunoblotting with anti-myc antibody. (c) NPCs were treated with DMSO, MLN4924 (13.3 μ M), PR-619 (25 μ M) and b-AP-15 (20 μ M) for 24h, and then the levels of Sox2 (green) and neuN (cherry) were determined with immunostaining. Nuclei were counterstained with HO (blue). Scale bar, 50 μ m. Each error bar shows the standard deviation of numbers of positive cells in 10 fields of view. *, *P*<0.05, **, *P*<0.01. Student's t-test. (d) ESCs, NPCs and neurons were treated with 10 μ g per ml CHX, and collected at the indicated time for western blot. Quantification of Sox2 levels relative to tubulin is shown. **: *P*<0.01, ***: *P*<0.001, two-way ANOVA test. (e) Sox2, DET1, CUL4A, COP1 and OTUD7B protein level was detected by immunoblotting in ESC, NPCs and neurons. The representative images are shown from 3 independent experiments. Unprocessed original scans of blots are shown in Supplementary Fig. 9.



Supplementary Figure 9. Uncropped scans of Western blots.



Supplementary Figure 9. Uncropped scans of Western Blots (Continuation 1).



Supplementary Figure 9. Uncropped scans of Western Blots (Continuation 2).



Supplementary Figure 9. Uncropped scans of Western lots (Continuation 3).