

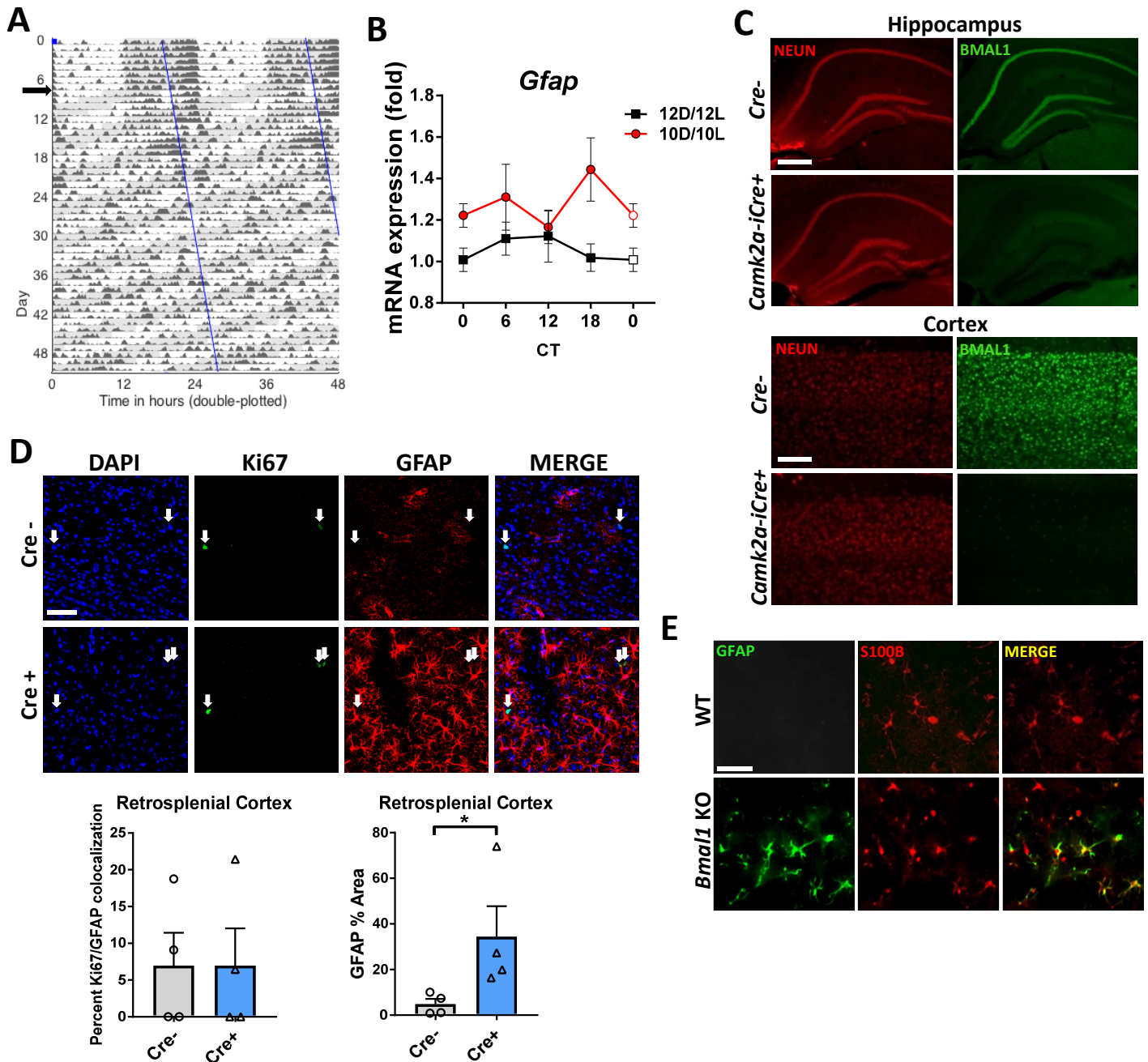
**Cell Reports, Volume 25**

## **Supplemental Information**

### **Cell-Autonomous Regulation of Astrocyte**

#### **Activation by the Circadian Clock Protein BMAL1**

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**Figure S1. Related to Figure 1; Characterization of mouse models with astrogliosis induced by loss of *Bmal1*.**

A. Representative actigraph of a WT mouse housed in 12h:12h light:dark, then switched to 10h:10h light:dark at black arrow, leading to behavioral arrhythmicity.

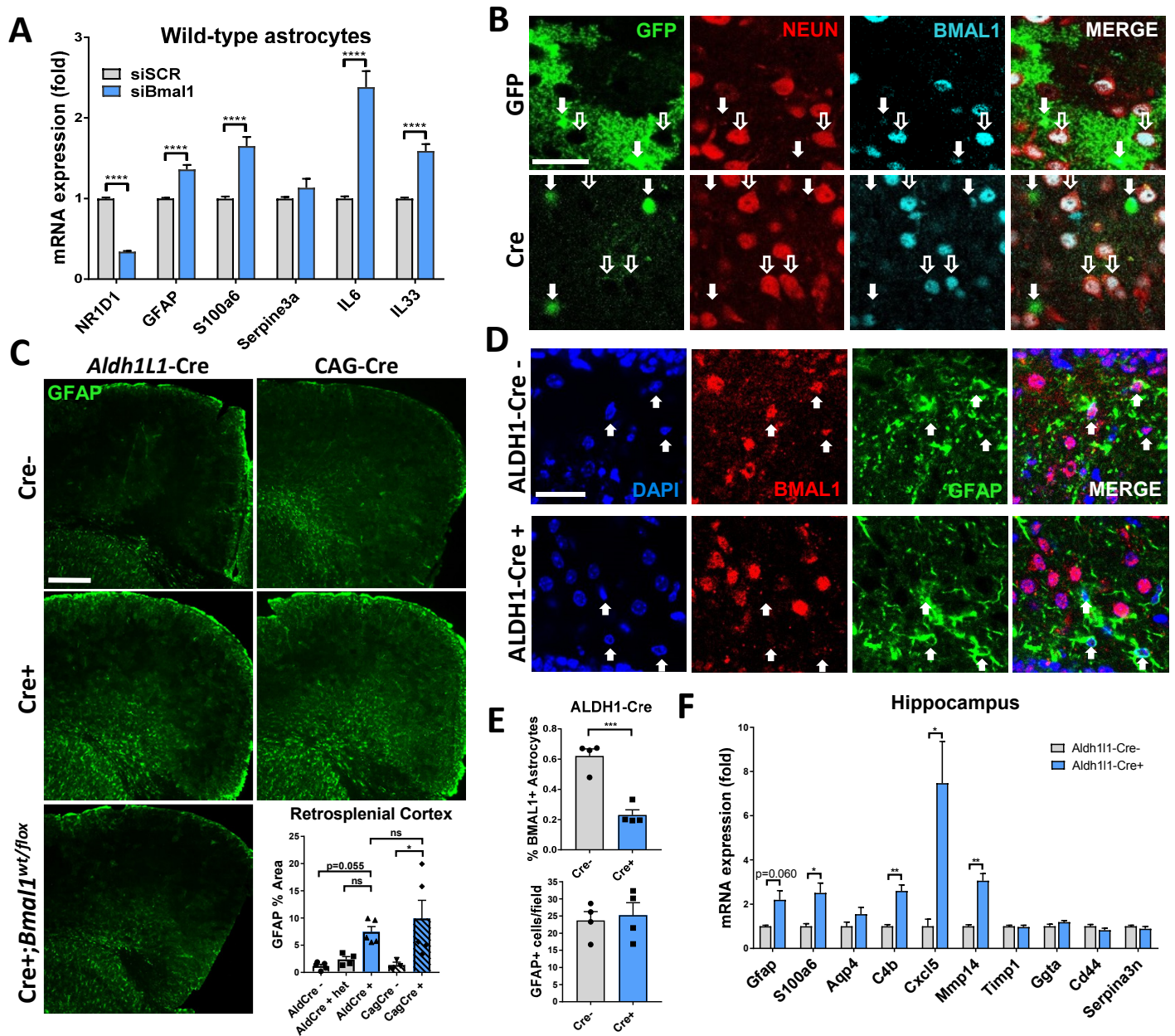
B. Cortical qPCR showing mRNA levels from 10h:10h light:dark WT mice from Fig. 1A, S1A reveals increased *Gfap* expression at most timepoints throughout the circadian day.

C. Hippocampal (top, scale bars = 150 $\mu$ m) and cortical (bottom, scale bars = 50 $\mu$ m) staining of NEUN (red) and BMAL1 (green) from 4mo *Camk2a-iCre+;Bmal1<sup>ff</sup>* mice and Cre- controls shows complete loss of neuronal BMAL1 in Cre+ animals.

D. Representative images (top) and quantification (bottom) illustrating minimal localization of the cell division marker Ki67 staining with GFAP+ astrocytes in cerebral cortex from 4 mo *Nestin-Cre+;Bmal1<sup>ff</sup>* (NBmal1 KO) mice (left graph), despite large increases in GFAP+ cells as compared to Cre- controls (right graph). Scale bar= 50 $\mu$ m.

E. NBmal1 KO mice have increased GFAP+ activated astrocytes in the cerebral cortex without major changes in number of total (S100B+) astrocytes. Scale bar = 50 $\mu$ m.

All data represent mean+SEM. \* $p < 0.05$  by 2-tailed T-test.



**Figure S2. Related to Figure 2; Characterization of astrogliosis seen with loss of *Bmal1* *in vitro* and *in vivo*.**

A. qPCR showing mRNA levels of several astrocyte activation and inflammatory genes 7 days after treatment of WT astrocytes with non-targeting siRNA (siSCR) or siRNA targeting *Bmal1* (siBmal1). N=12-15 experiments.

B. Representative cortical images from mice in Fig. 2F-H. *Top row*. AAV8-GFAP-GFP-infected cells (solid arrows) show whole-cell GFP expression, persistent colocalized nuclear BMAL1, and fail to colocalize with neuronal nuclei (hollow arrows). *Bottom row*. AAV8-GFAP-Cre infected cells show only nuclear GFP expression (solid arrows, Cre<sup>eGFP</sup> fusion protein), loss of nuclear BMAL1 and fail to colocalize with neuronal nuclei (hollow arrows). Scale bar= 30µm.

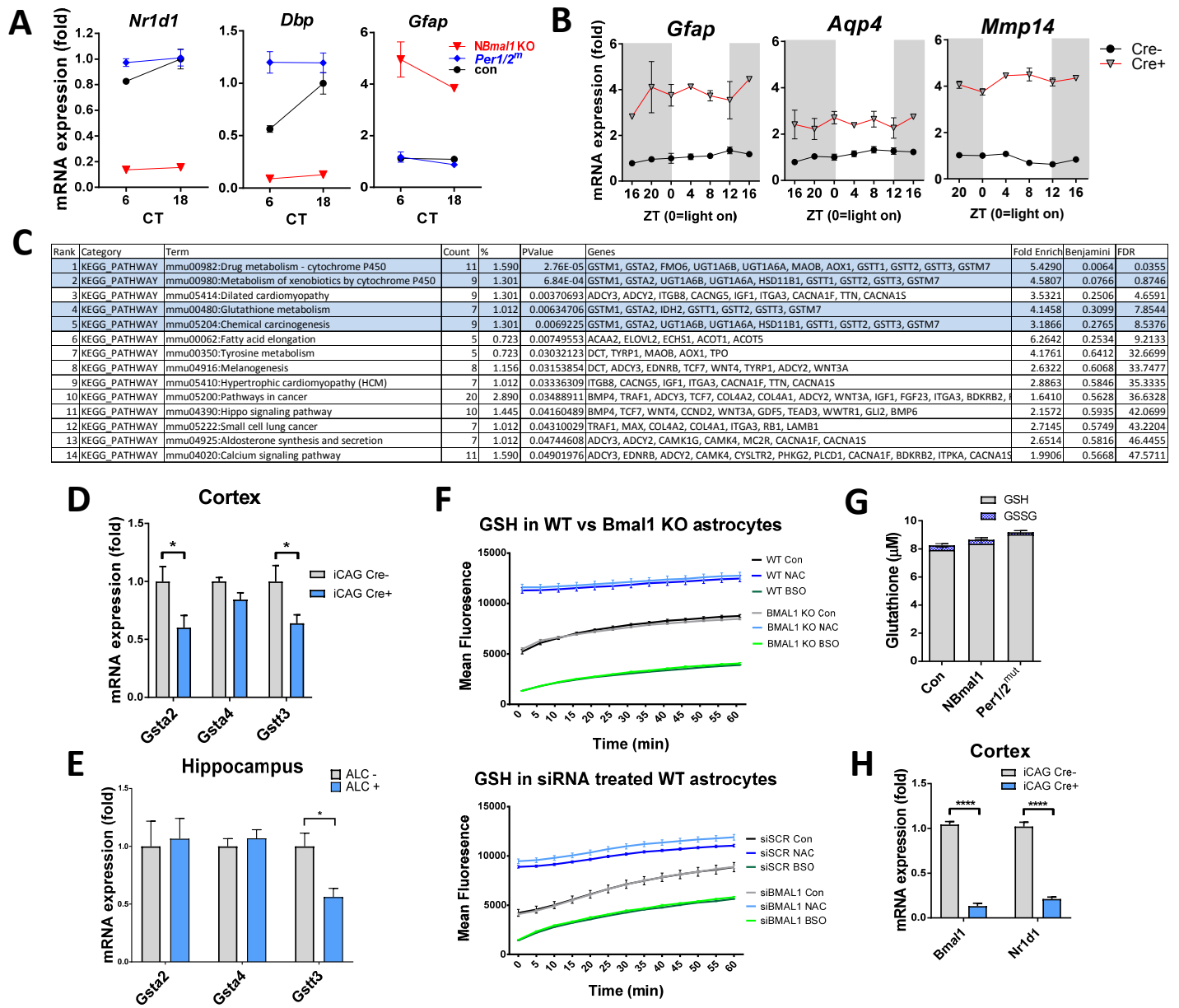
C. Representative images, quantification of %GFAP area in retrosplenial cortex of mice in Fig. 2H. Scale bar = 100µm.

D. Representative confocal images of *Aldh111-Cre+;Bmal1<sup>fl/fl</sup>* mice and Cre- controls. DAPI+, GFAP+ (solid arrows) astrocytes colocalize with BMAL1 in Cre- mice, but not in Cre+ mice. Scale bar= 30µm.

E. Quantification of DAPI/GFAP/BMAL1+ cells (top) or GFAP+ cells per field of view (bottom) in *Aldh111-Cre+;Bmal1<sup>fl/fl</sup>* mice vs Cre- controls in hippocampus. Each data point = 1 mouse. 6 fields/mouse with an average of 24.5 GFAP+ cells/field, totaling 1,176 cells counted. Note that there is not an increase in GFAP+ cells in hippocampus, just the size and shape of these cells.

F. mRNA levels of astrocyte activation markers in hippocampus from *Aldh111-Cre+;Bmal1<sup>fl/fl</sup>* mice vs Cre- controls.

All data represent mean+SEM. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001 by 2-tailed T-tests (A), (E), and (F) or 1-way ANOVA (D) with Hom-Sidak correction for multiple comparisons when applicable.



**Figure S3. Related to Figure 3; Pathway analysis, GSTx, and GSH with *Bmal1* deficiency.**

A. Expression of *BMAL1* transcriptional targets and *Gfap* from microarray in Fig. 3A at CT 6 and 18.

B. Expression of astrocyte activation transcripts *Gfap*, *Aqp4*, and *Mmp14* in cortex of *NBmal1* KO (*Cre+*, red) and *Cre*-control mice (black) at 4 hour timepoints across the circadian cycle. No clear circadian oscillations were observed in *Cre-* or *Cre+* mice.  $N=3-4$  mice/genotype/timepoint.

C. KEGG pathway analysis of *NBmal1* KO microarray data. Blue = pathways related to glutathione homeostasis.

D. qPCR of mRNA expression of glutathione transferases in the cortex of *iCAG-Cre+;Bmal1<sup>fl/fl</sup>* mice (Fig. 3F), 9 days after tamoxifen treatment, normalized to *Cre-*, *Bmal1<sup>fl/fl</sup>* controls.

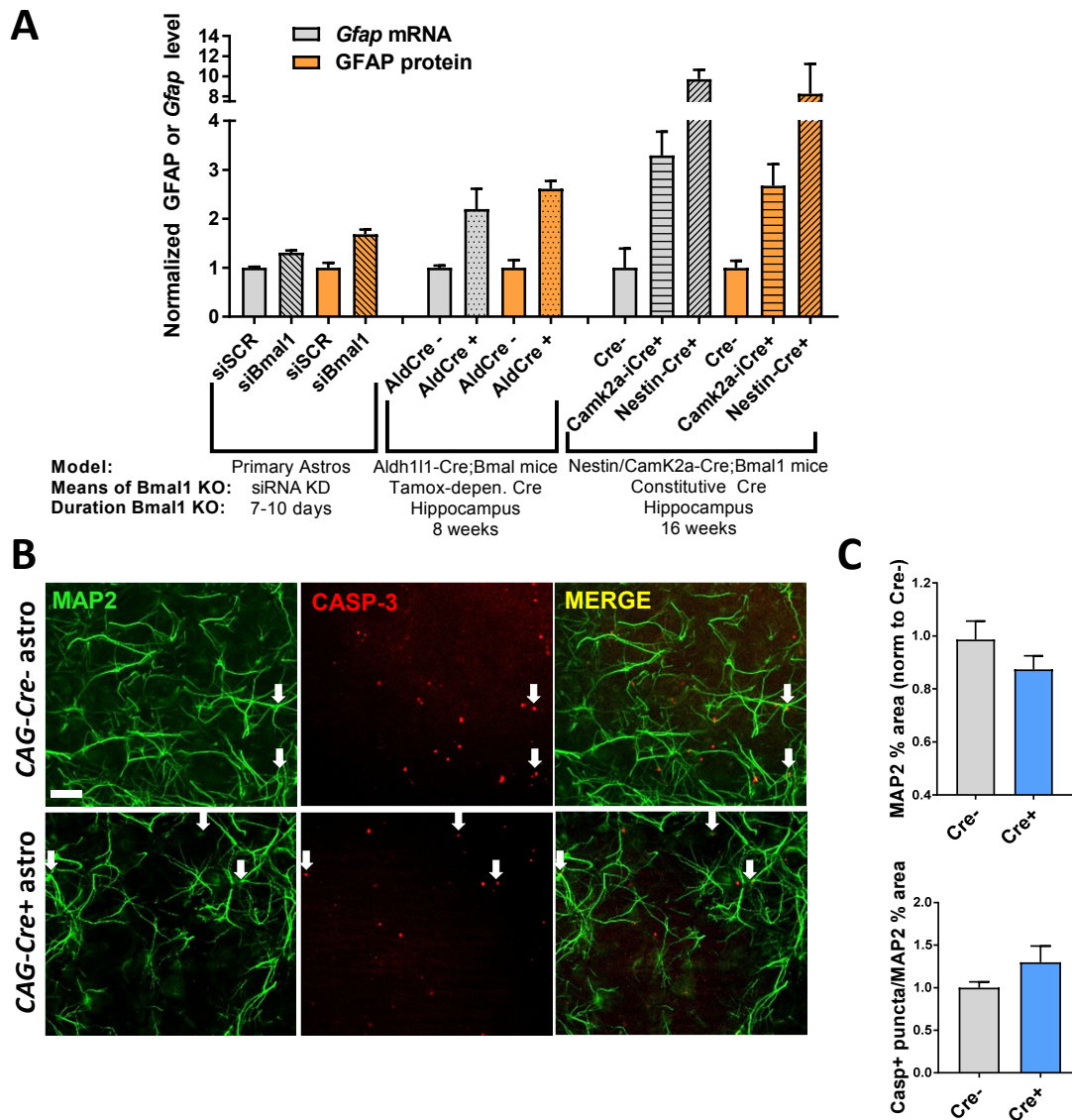
E. qPCR of mRNA expression of glutathione transferases in hippocampus of *Aldh1l1-Cre+;Bmal1<sup>fl/fl</sup>* mice (Fig. 2H).

F. GSH levels in *Bmal1* KO (top) and si*Bmal1* treated WT (bottom) astrocyte enriched cultures along with appropriate controls as measured by fluorometric intracellular glutathione detection assay.

G. Quantification of GSH and GSSG levels in cortical tissue (CT 10) from control, *NBmal1* KO, and *Per1/2<sup>mut</sup>* mice. None of the averages were significantly different.

H. mRNA expression of *Bmal1* and *Nr1d1* in the cortex of *iCAG-Cre+;Bmal1<sup>fl/fl</sup>* mice used in Fig. 3G, 9 days after tamoxifen treatment, normalized to *Cre-*, *Bmal1<sup>fl/fl</sup>* controls.

All data represent mean $\pm$ SEM. \* $p<0.05$ , \*\*\*\* $p<0.001$  by 2-tailed T-test with Holm-Sidak correction for multiple comparisons when applicable.



**Figure S4. Related to Figures 3 and 4; Comparison of *Gfap* mRNA and GFAP protein across experiments; Further characterization of neuronal support deficits observed with loss of *Bmal1*.**

A. Comparison of *Gfap* mRNA (quantified by qPCR) and GFAP protein (quantified by immunofluorescence) across experiments in this manuscript. Note that in both cultures cells and two distinct Cre-Lox mouse models, *Gfap* mRNA and GFAP protein are very closely related. Data is shown separately in Figs. 1 and 2.

B. Representative images showing MAP2+ (green) and cleaved-Caspase-3+ (CASP-3, red, arrows) WT neurons (grown as in Fig. 4A) at DIV 7. Scale bar = 100µm.

C. Quantification of MAP2 percent area (top) and cleaved-Caspase-3+ (bottom) neurons from (C) shows non-significant trends toward decreased MAP2 and increased cleaved-Caspase-3+ neurons when neurons are plated on BMAL1-deficient astrocytes (Cre+).

All data represent mean+SEM.