A complete *Leishmania donovani* reference genome identifies novel genetic variations associated with virulence.

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Chromosome	Start	Stop	Strand	Gene ID	Internal Stop	Frameshift	L. Major origin
LdCL_06	518294	520041	+	LdCL_060018900		XXXX	LmjF.22.0010
LdCL_08	550785	551865	+	LdCL_080018700		XXXXX	LmjF.08.1265
LdCL_10	79199	79729	+	LdCL_100006900	XX	XX	LmjF.10.0185
LdCL_12	603220	604085	+	LdCL_120018300		XXXXXX	LmjF.12.0880
LdCL_16	316885	317791	-	LdCL_160014100	XXXX	XXXX	LmjF.16.0880
LdCL_26	580729	582519	+	LdCL_260021400	XXXX	XXX	LmjF.26.1590
LdCL_27	527810	529301	-	LdCL_270017800	XXXX	XXXXXX	LmjF.14.0180
LdCL_27	732393	732680	-	LdCL_270023400*	Х		LmjF.27.1740
LdCL_31	1179486	1180414	-	LdCL_310031700	XXXXX	XXX	LmjF.31.2310
LdCL_31	1334630	1335706	-	LdCL_310035400	XXX	XX	LmjF.29.1570
LdCL_32	784327	785612	-	LdCL_320026000	XX	Х	LmjF.32.1950
LdCL_33	702441	703568	+	LdCL_330023800	XXXXX	XXXXX	LmjF.32.2640
LdCL_35	1479577	1480153	-	LdCL_350044600	XX	XX	LmjF.35.3910

Supplementary Table S1. List of *L. donovani* pseudogenes derived from *L. major* 

13 pseudogene annotations in *L. donovani* originated from 13 intact functional genes from *L. major* in the current assembly. The 13 pseudogenes were common across all 3 isolates sequences in this study. The locations of the pseudogenes are given in chromosomal position from start to stop. Strand indicates direction of transcription. Each stop codon and frameshift are marked with an X *L. major* origin denotes the equivalent functional gene ID in *L. major*.

\*One gene contains only one stop codon; this gene however is also missing a start codon.

## Supplementary Figure S1. Coverage graphs of the new *L. donovani* assembly

























Position (bp)



Read coverage of each chromosome in the new assembly. Graphs were generated from the moving average of the raw coverage at each position with a window size of 450bp, equivalent to the average Illumina read insert size. Illumina data plotted in blue, PacBio data plotted in orange.

## **Supplementary Figure S2**. Complete amino acid sequences of A2 genes from the attenuated cutaneous *L. donovani* strain from Sri Lanka

#1	#2	#3	#4
MKIRSVRPLVVLLVCVAAVLALSASAEPHKAAVD	$\tt MKIRSVRPLVVLLVCVAAVLALSASAEPHKAAVD$	MKIRSVRPLVVLLVCVAAVLALSASAEPHKAA VD	MKIRSVRPLVVLLVCVAAVLALSASAEPHKAA VD
VGPLSVD	VGPLS <b>VD</b>	VGPLSVD	VGPLS <b>VD</b>
VGPLSVGPQS VGPLS <b>VD</b>	VGPLSVGPQS VGPLSVD	VGPLSVGPQS VGPLS <b>VD</b>	VGPLSVGPQS VGPLS <b>VD</b>
VGPQAVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQAVGPLS	VGPLSVGPQA VGPLS <b>VD</b>	VGPLSVGPQA VGPLS <b>VD</b>
VGPQSVGPLS VD	VGPQSVGPLS VD	VGPQAVGPLS VGPQSVGPLS	VGPQAVGPLS VD
VGPQAVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQAVGPLS	VGPQSVGPLS VGPLSVGPLS	VGPQAVGPLS VGPQSVGPLS
VGPQAVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQAVGPLS	VGPQSVGPLS VGPQAVGPLS	VGPQSVGPLS VGPLSVGPLS
VGPQAVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQAVGPLS	VD	VGPQSVGPLS VGPQSVGPLS
VGPQAVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQSVGPLS	VGPQAVGPLS VGPQSVGPLS	VD
VGPQSVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPQAVGPLS VGPQSVGPIS
VGPQSVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQSVGPLS	VD	VGPQAVGPLS VGPQAVGPLS
VGPQAVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQSVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQSVGPLS
VGPQSVGPLS VGPQSVGPLS	VD	VD	VGPQSVGPLS VGPQSVGPLS
VGPQSVGPLS VD	VGPQAVGPLS VGPQAVGPLS	VGPQSVGPLS VGPQSVGPLS	VGPLSVGPLS VGPLSVGPQS
VGPQAVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPQSVGPLS VD	VGPLSVGPQS VGPLSVGPQS
VGPQAVGPLS VGPQAVGPLS	VGPQSVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS
VGPQSVGPLS VGPQSVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQS VGPLSVGPQS
VGPQAVGPLS VGPQAVGPLS	VGPQSVGPLS VGPQSVGPLS	VGPLS <b>VD</b>	VGPLSVGPQS VGPLSVGPQS
VGPQAVGPLS VGPQAVGPLS	VGPQSVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS
VGPQSVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQAVGPLS	VGPLSVGPQS VGPLSVGPQS
VGPQSVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQSVGPLS	VGPQAVGPLS VGPLSVGPLS	VGPLSVGPQS VGPLSVGPQS
VGPQAVGPLS VGPQSVGPLS	VGPQSVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQA VGPLSVGPQS
VGPQSVGPLS VGPQSVGPLS	VD	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQS VGPLSVGPQS
VGPQSVGPLS VGPQSVGPLS	VGPQAVGPLS VGPQAVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQS VGPLSVGPQS
VGPQSVGPLS VGPQAVGPLS	VGPQAVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQA VGPLSVGPQS
VGPQSVGPLS VGPLSVGPQS	VGPQSVGPLS VGPQAVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQS VGPLSVGPQS
VGPLSVGPQS VGPLSVGPQS	VGPQAVGPLS VGPQAVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQS VGPLSVGPQA
	VGPQAVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPLSVGPQS VGPLSVGPQA
	VGPQSVGPLS VGPQSVGPLS	VGPLSVGPQA VGPLSVGPQS	VGPLSVD
	VGPQSVGPLS VGPQAVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPQAVGPLS VGPQSVGPLS
	VGPQSVGPLS VGPQSVGPLS	VGPLSVGPQS VGPLSVGPQS	VGPQSVGPLS VGPQAVGPLS
	VGPQSVGPLS VGPQAVGPLS	VGPLSVGPQA VGPLSVGPQS	VGPQSVGPLS VGPLSVGPQS
	VGPQSVGPLS VGPQSVGPLS	VGPLSVGPQA VGPLS <b>VD</b>	VGPLSVGPQS
	VGPQAVGPLS VGPQSVGPLS	VGPQAVGPLS VGPQSVGPLS	
	VGPLSVGPQS VGPLSVGPQS	VGPQSVGPLS VGPQAVGPLS	
		VGPQSVGPLS VGPLSVGPQS	
		VGPLSVGPQS	
VDVSPVS	VDVSPVS	VDVSPVS	VDVSPVS

The DNA sequences obtained from the assembled A2 cluster were translated into their

corresponding amino acids. All four A2 proteins share a common leader and terminal sequence.

Deviations from the VGPLS VGPQS/A by the insertion of valine-aspartate are highlighted in

bold.

Supplementary Figure S3. Synteny map of the *L. major* and *L. donovani* chromosomes.



LmjF.01

LdCL 01



LdCL 02



LdCL 03





LdCL 04



LdCL 05





LdCL 06



LdCL 07



LdCL 08



LdCL 09



LdCL 10



LdCL 11





LdCL 12





LdCL 13



LdCL 14



LdCL 15





LdCL 16





LdCL 17





LdCL 18





LdCL 19





LdCL 20



LdCL 21





LdCL 22





LdCL 24





LdCL 25









LdCL 27





LdCL 29



700kb -

- 600kb

800kb

- 500kb





LdCL 31















LdCL 36

Annotations from both species were aligned to their respective genome and compared to each other. Each chromosome is represented as a half circle. ORFs are colored according to their coding strands. Black lines represent pairs of homologous genes between *L. major* (top) and attenuated cutaneous *L. donovani* (bottom).



Supplementary Figure S4. Confirmation of a 25kb deletion on chromosome 36

Primers were designed to amplify a fragment spanning across the deletion (span) and to amplify a region internal to the deletion (int). The IV strain only produced an amplicon from either end of the deletion indicating the 28kb region collapsed to 3kb but was unable to produce any amplicon internal to the deleted region, indicating the 25kb region was absent rather than translocated. The CL strain mainly produced the internal region as the entire 28kb fragment is too long to be efficiently resolved by PCR and a faint band also appeared in the span reaction indicating the presence of some cells within the population that also contained the deletion.

## **Supplementary methods S1**

```
Relaxed.spec:
merSize=14
asmOvlErrorRate=0.10
asmUtgErrorRate=0.10
asmCnsErrorRate=0.10
asmCgwErrorRate=0.10
asmOBT=1
asmObtErrorRate=0.08
asmObtErrorLimit=4.5
utgGraphErrorRate=0.05
utgMergeErrorRate=0.05
ovlHashBits=26
ovlHashLoad=0.80
sensitive = 1
maxCoverage=60
gridOptionsOverlap = -pe threads 16 -1 mem=2GB
gridOptionsConsensus = -pe threads 16
gridOptionsScript = -pe threads 5
merylThreads = 60
ovlThreads = 60
merylMemory = 12000
ovlStoreMemory = 12000
ovlConcurrency = 1
mbtThreads=60
merOverlapperThreads=60
frqCorrThreads=60
batThreads=60
```