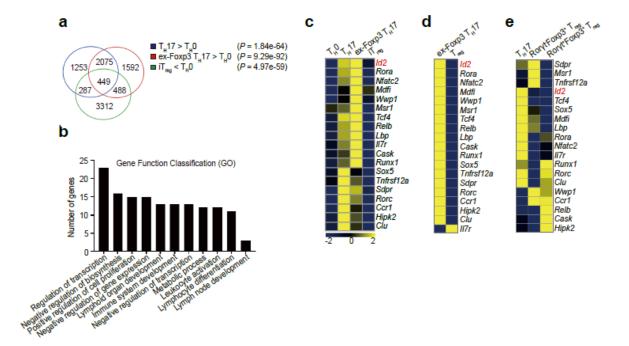
Supplementary information

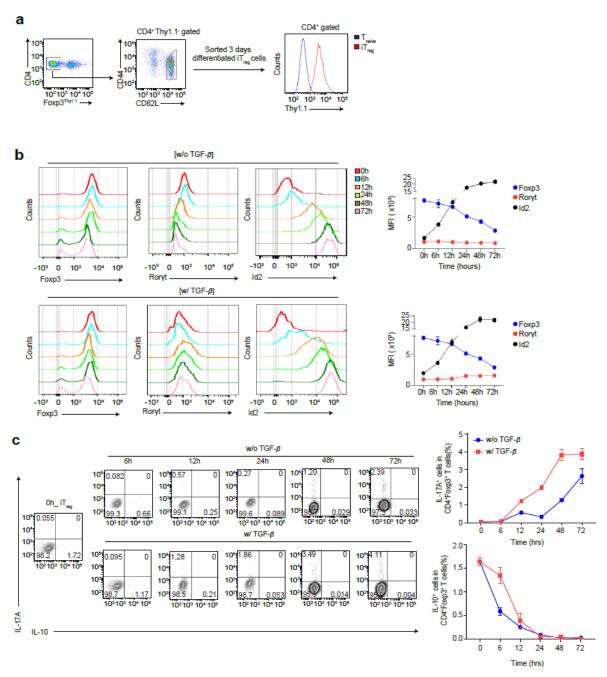
# Inflammation-induced Id2 promotes plasticity in regulatory T cells

Hwang et al.



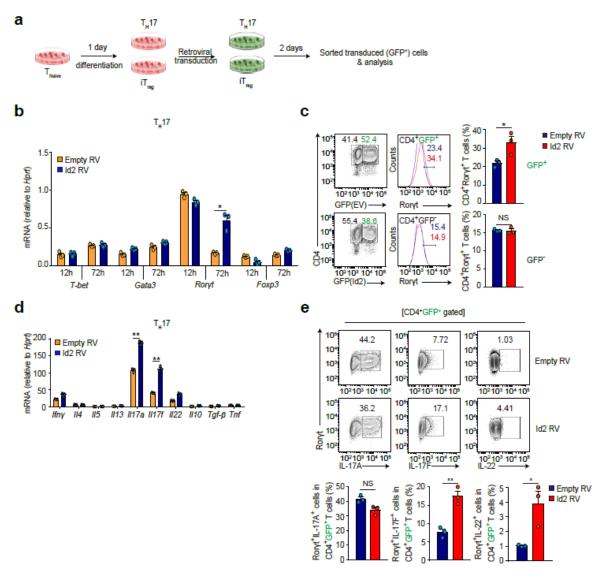
# Id2 is highly expressed in ex-Foxp3 $T_{\rm H}$ 17 cells.

**a** Microarray analysis (GEO accession code GSE48428) representing overlap of differentially expressed genes more in ex-Foxp3  $T_H17$  cells and  $T_H17$  cells and less in i $T_{reg}$  cells than  $T_H0$  cells (2-fold). **b** Gene Ontology (GO) analysis on 11 significant GO terms associated with 449 genes in which were normally suppressed by Foxp3 expression in i $T_{reg}$  cells while induced as absence of Foxp3 expression in ex-Foxp3  $T_H17$  cells and  $T_H17$  cells. **c** Heat map of selected 19 target genes related to immune system and /or are involved in regulation of transcription from (**b**). **d**,**e** Validation of selected 19 target genes expression from two other independent studies (GEO accession code GSE80804;**d**, GSE60059;**e**).



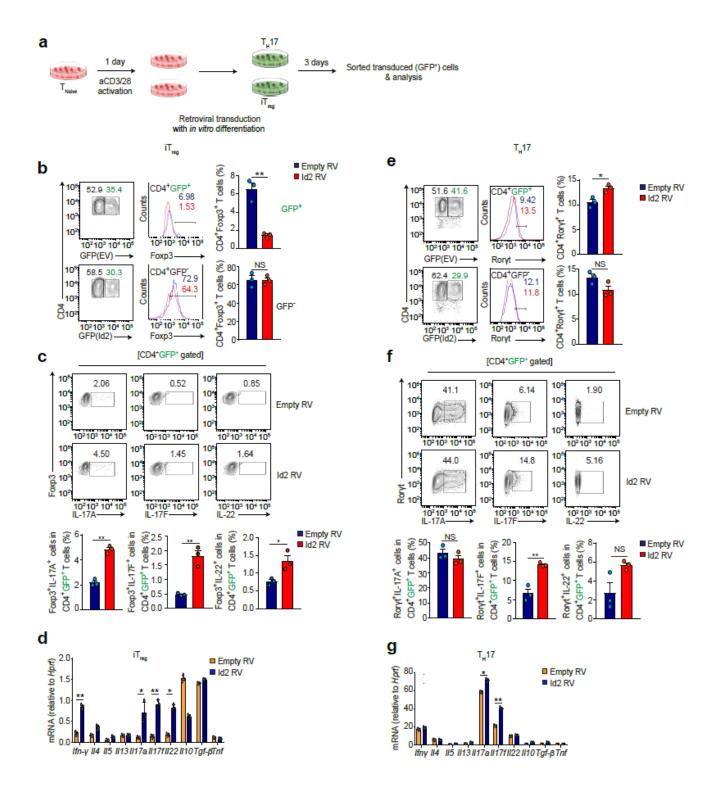
### Kinetics of iT<sub>reg</sub> - ex-Foxp3 T<sub>H</sub>17 conversion.

**a** Strategy for sorting naïve CD4<sup>+</sup> T cells and differentiated  $iT_{reg}$  cells from Foxp3<sup>Thy1.1</sup> reporter mice. **b,c** Sorted CD4<sup>+</sup>CD62L<sup>hi</sup>CD44<sup>lo</sup>Thy1.1<sup>-</sup> naïve T cells were activated *in vitro* under  $iT_{reg}$  cell differentiation conditions for 3 days. After 3 days,  $iT_{reg}$  cells were restimulated *in vitro* in ex-Foxp3 T<sub>H</sub>17 polarizing conditions with(w/) or without(w/o) exogenous TGF- $\beta$  for additional 3 days. Representative FACS plot showing expression of Foxp3, Roryt, Id2 (**b**), IL-10 and IL-17A (**c**) positive populations in CD4<sup>+</sup>Foxp3<sup>+</sup> T cells after harvesting cells at indicated time points (Data is summary of three experiments).



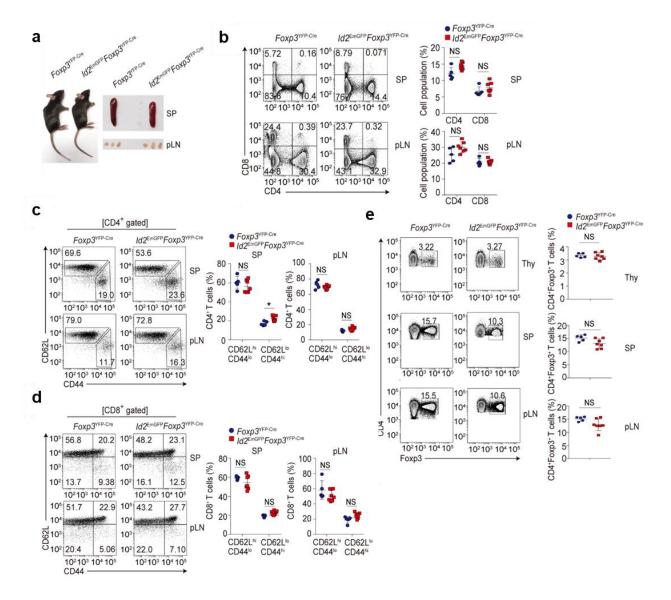
### Effect of Id2 overexpression during T<sub>H</sub>17 and iT<sub>reg</sub> differentiation conditions.

a A Schematic of retroviral transduction of naïve CD4<sup>+</sup> T cells. Naïve CD4<sup>+</sup> T cells that were activated *in vitro* were transduced with Empty-GFP RV or Id2-GFP RV on day 1 under each T helper (T<sub>H</sub>17 and iT<sub>reg</sub>) cell-differentiation conditions. The following day 2 after culture, GFP<sup>+</sup> cells were sorted and analyzed. **b** RT-qPCR analysis of *T-bet*, *Gata3*, *Roryt* and *Foxp3* mRNA expression between Empty RV and Id2 RV transduced T cells on day 3 post-infection during T<sub>H</sub>17 cell-differentiation conditions (n=3-5, per group). **c** Flow cytometry analysis of Roryt expression in Empty RV or Id2 RV transduced (GFP<sup>+</sup>) and nontransduced (GFP<sup>-</sup>) T<sub>H</sub>17 cells. **d** RT-qPCR analysis of the indicated cytokine encoding mRNA expression in Empty RV and Id2 RV transduced T cells after day 3 post-infection during iT<sub>reg</sub> cell-differentiation conditions. **e** Flow cytometry analysis of IL-17A, IL-17F and IL-22 from Empty RV or Id2 RV transduced CD4<sup>+</sup>GFP<sup>+</sup> T<sub>H</sub>17 cells. NS=Non-Significant, \* *P* < 0.05, \*\* *P* < 0.005 (Student's *t*-test). Data are representative two to three independent expression the probability of the solution of the s



### Effect of Id2 overexpression concomitant to iTreg or TH17 differentiation.

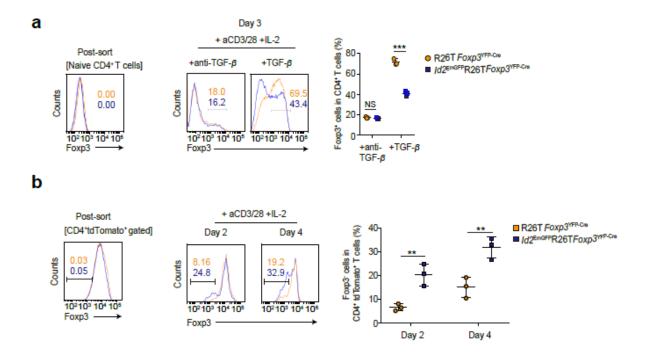
**a** Naïve CD4<sup>+</sup> T cells were sorted from wild-type C57BL/6 (B6) mice, and transduced with control vector (Empty RV) or vector encoding Id2 cDNA (Id2 RV) after 1 day activated by plate bound anti-CD3/28. Cells were then differentiated under iT<sub>reg</sub> or T<sub>H</sub>17 differentiation conditions. After 3 days cells were harvested and Id2 expression was measured by RT-qPCR. **b,c** Flow cytometry analysis of Foxp3 (**b**) expression and IL-17A, IL-17F and IL-22 (**c**) between Empty RV or Id2 RV transduced (GFP<sup>+</sup>) and non-transduced (GFP<sup>-</sup>) iT<sub>reg</sub> cells. **d**, RT-qPCR analysis of mRNA expression for the indicated cytokines in Empty RV and Id2 RV transduced T cells sorted after 3 days post-infection followed by iT<sub>reg</sub> cell-differentiation. **e,f** Flow cytometry analysis of Roryt (**e**) expression and IL-17A, IL-17F and IL-22 (**f**) in Empty RV or Id2 RV transduced (GFP<sup>+</sup>) and non-transduced (GFP<sup>-</sup>) T<sub>H</sub>17 cells. **g** RT-qPCR analysis of the expressions of indicated cytokines in Empty RV and Id2 RV transduced T cells sorted after 3 days post-infection conditions. NS=Non-Significant, \* P < 0.05, \*\* P < 0.005 (Student's *t*-test). Data are representative of two independent experiments (error bars, s.d.).



**Supplementary Figure 5** 

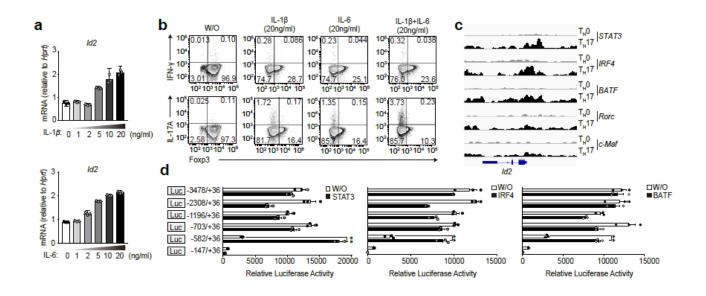
# Analysis of 6-8 weeks of *Id2*<sup>EmGFP</sup>*Foxp3*<sup>YFP-Cre</sup> mice.

a A representative picture of 6-8 week-old  $Foxp3^{\text{YFP-Cre}}$  and  $Id2^{\text{EmGFP}}Foxp3^{\text{YFP-Cre}}$  mice (*left*) and spleen (SP) and peripheral lymph nodes (pLN) from those mice (*right*). b Percentages of CD4<sup>+</sup> and CD8<sup>+</sup> T cells from SP and pLN ( $Foxp3^{\text{YFP-Cre}}$ ; n=5,  $Id2^{\text{EmGFP}}$ ; n=7). c,d Percentages of CD62L<sup>hi</sup>CD44<sup>lo</sup> (naïve) and CD62L<sup>lo</sup>CD44<sup>hi</sup> (effector/memory) cells among CD4<sup>+</sup> and CD8<sup>+</sup> T cells in SP and pLN ( $Foxp3^{\text{YFP-Cre}}$ ; n=5,  $Id2^{\text{EmGFP}}$ ; n=7). e Foxp3<sup>+</sup> cell percentages among CD4<sup>+</sup> T cells in thymus, SP and pLN from 6-8 week-old mice ( $Foxp3^{\text{YFP-Cre}}$ ; n=5,  $Id2^{\text{EmGFP}}$ ; n=7). NS=Non-Significant, \* P < 0.05. All data are representative three independent experiments (error bars, s.d.).



 $T_{reg}$ -specific ectopic expression of Id2 results in enhanced loss of Foxp3 expression from  $T_{reg}$  cells, and not enhanced stability of promiscuous Foxp3 expressing naïve T cells.

**a** Sorted naïve CD4<sup>+</sup> T cells from R26T*Foxp3*<sup>YFP-Cre</sup> and *Id2*<sup>EmGFP</sup>R26T*Foxp3*<sup>YFP-Cre</sup> mice were activated with anti-CD3, anti-CD28 and IL-2 in the presence of TGF- $\beta$  blocking antibody (anti-TGF- $\beta$ ) or TGF- $\beta$  for 3 days. After 3 days, Foxp3-expression was measured by flow cytometry. **b** Sorted T<sub>reg</sub> cells (CD4<sup>+</sup>tdtmomato<sup>+</sup>YFP<sup>+</sup>) from R26T*Foxp3*<sup>YFP-Cre</sup> and *Id2*<sup>EmGFP</sup>R26T*Foxp3*<sup>YFP-Cre</sup> mice were activated with anti-CD3, anti-CD28 and IL-2 *in vitro*. Expression of Foxp3 was measured by flow cytometry at indicated time points. NS=Non-Significant, \*\* *P* < 0.01, \*\*\* *P* < 0.001, \*\*\*\* *P* < 0.0001. Data are representative of three independent experiments (error bars, s.d.).

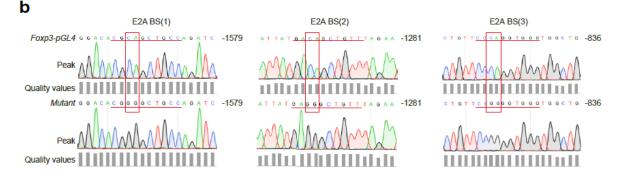


# Id2 is induced by IL-1 $\beta$ and IL-6 signaling-mediated STAT3, IRF4 and BATF in T<sub>reg</sub> cells.

**a** RT-qPCR analysis of *Id2* mRNA expression given at different dose of IL-1 $\beta$  (upper) and IL-6 (lower) in iT<sub>reg</sub> cells. **b** Intracellular staining of IFN- $\gamma$ , IL-17A and Foxp3 expression in W/O, IL-1 $\beta$  (20ng/ml), IL-6 (20ng/ml) and IL-1 $\beta$ + IL-6 (20ng/ml) conditions for 2 days in iT<sub>reg</sub> cells. **c** ChIP-seq results for STAT3, IRF4 and BATF at the Id2 locus in T<sub>H</sub>0 and T<sub>H</sub>17 cells (GEO accession code GSM1004793). **d** Analysis of luciferase activity from six serial-deletion of Id2 promoter-luciferase constructs transfected into HEK 293T cells with STAT3, IRF4 and BATF expressing vectors, respectively. (W/O; control). Luciferase activities were measured with the reporter activities normalized to renilla luciferase activity. \*\* *P* < 0.005 (Student's *t*-test). All data are representative three independent experiments with similar results (error bars, s.d.).

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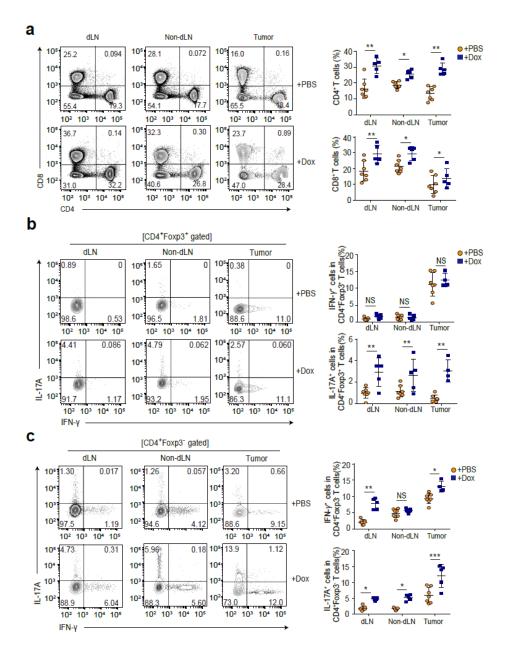
Foxp3 promoter sequences -1702 to +174 TSS E2A Binding Site (BS) 1 E2A Binding Site (BS) 3 Foxp3 locus 1593 to -1584 -1295 to -1286 -837 to -829 E2A Binding Site (BS) 2 : exon BS: Binding site TG<u>GATACCTGGAACTCTTAGCTC</u>TCTGCAGGATGCCAGGGCCACCAAAGGCTGGAAGCCTTAGCCGTGCCTTGTCAGGAAAAAC -1659 ChIP primer (F1) TCTGTGGAGGCTCGTCTGTAGTAAACAGTGGTTACAGGGAGCCGGTCTGTGCCAAATCGAGGACA<u>CGCAGCTGCC</u>AGATCTTG -1576 E2A BS (1) AATACAAACCTTAAAACCTCACAAACATCAAGTTCCAGAGGAGTCTCCAAGTCCTAGAACTTCTATGACACTGTTGGCTTCAGGA -1491 -1406 -1322 AGTTAAATGTTGGCTACCAAAATTAT<u>GACAGCTGTTTAGAATCCTAAACCTTTGC</u>AAACGGGAGTGTTTCTTTCCTTTGTTTGTG -1236 -1148 CTTAAACTCTTGATTCTTCTGCCTCAGTTTCTGGGGTGCTGGGATTACTGGTATGTGATACTGGATGAAACTGGAACTTTTCAGA -1063 GTAGACTGTTACAAAGTTTAGAATCATCAGGCTATGGCTATATTGTTCCTGACAGGACTAGGACCCTGGGCCGCTATGTGTATG -979 GTTTTTTGTTTGTTTGTTTGTTTAACAACCCAGAGCCTTGTGCGTGTTAAACAAGCACTCTGGCACTGAGCTGCAATGGCCAGCCTT -894 -817 -733 -652 -569 GAGGTTTGGAGCAGAAGGAAGCCTCTGGAGACAGAGCACTACCCCACCTCTCCCCTGGCTGCTTCCCATTCACATGGCAGGC -487 TTCAGATCCCTTCTTCTGTTCAACCCAGCGATCCTCCAACGTCTCACAAACACAATGCTGTCTCTACCTGCCTCGG -403 TGTGATTTGACTTATTTTCCCCTCAGTTTTTTTTCTGACTCTACACACTTTTGTTTAAGAAATTGTGGTTTCTCATGAGCCCTGTT -316 ATCTCATTGATACCTTTTACCTCTGTGGTGAGGGGAAGAAATCATATTTTCAGATGACTTGTAAAGGGCAAAGAAAAAACCCAAA -231 -146 -61 TTTTCTTCTCGGTATAAAAGCAAAGTTGTTTTTGATAATGTGGCAGTTTCCCACAAGCCAGG +1



### **Supplementary Figure 8**

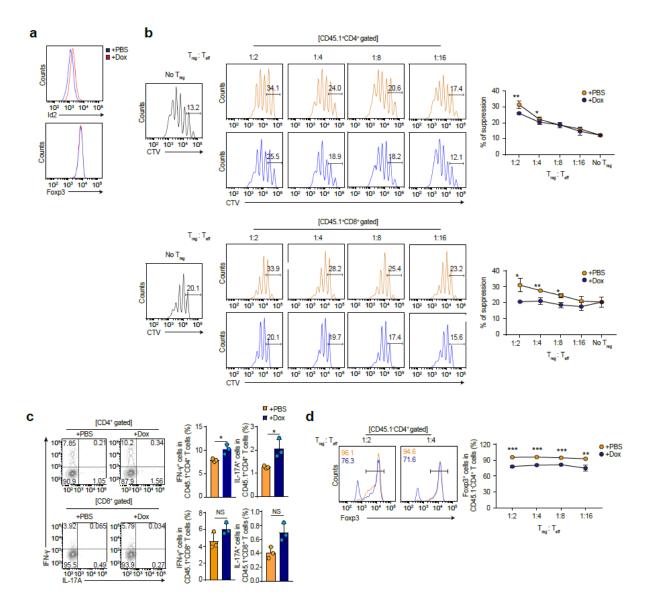
### Nucleotide sequences of the Foxp3 promoter and E2A binding sites.

**a** Sequences of *Foxp3* promoter from -1741 to +1. Highly-putative E2A binding sites (score>10) are marked in blue, red and yellow (BS1, BS2 and BS3, respectively). Each ChIP-primers (F/R) were indicated, respectively. **b** Sequencing results of mutation of E2A binding sites (Mutants; BS1, BS2 and BS3) in the corresponding control (Foxp3-pGL4). The rectangle indicates the nucleotides comprising a major E2A binding sites (E-box site).



### Increased T cell infiltration and IL-17A expression in doxycycline-treated TetR-*Id2*<sup>EmGFP</sup> mice.

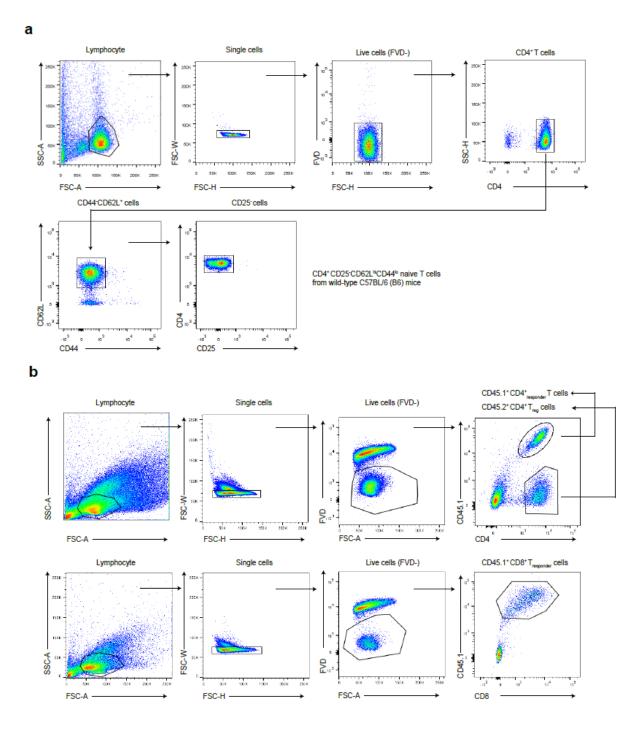
a Flow cytometry analysis of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in tumor-draining lymph nodes (dLN), non-draining lymph nodes(non-dLN) and tumor (PBS; n=7, Dox; n=5). **b,c** Intracellular staining of cytokines in CD4<sup>+</sup>Foxp3<sup>+</sup> and CD4<sup>+</sup>Foxp3<sup>-</sup> T cells isolated from dLN, non-dLN and tumor (PBS; n=7, Dox; n=5), stimulated with phorbol myristate acetate (PMA) and ionomycin for 6 h. NS=Non-Significant, \* P < 0.05, \*\* P < 0.005, \*\*\* P < 0.001 (Student's *t*-test). All data are representative of three independent experiments (error bars, s.d.).



**Supplementary Figure 10** 

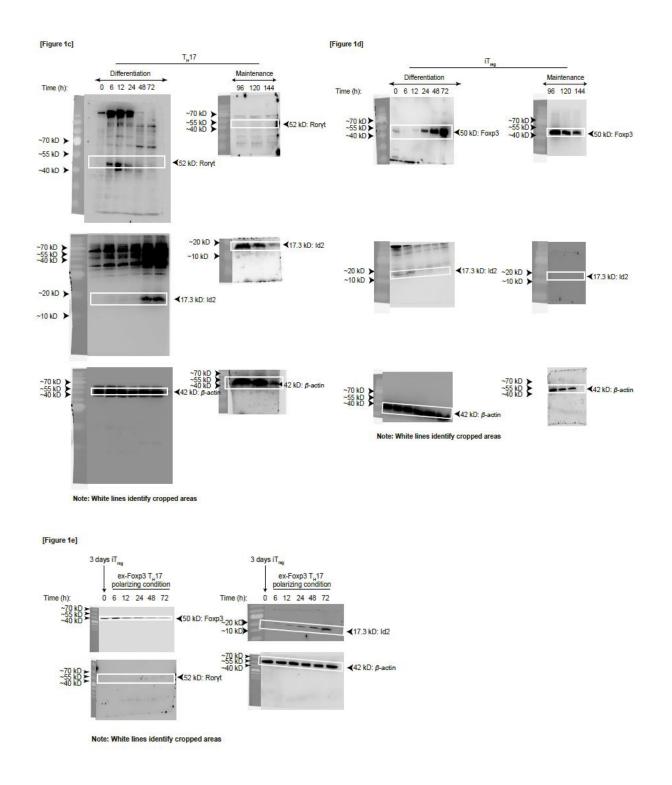
# Dox-treated Id2 induced Treg cells display reduced suppressive capacity in vitro.

a Flow cytometry analysis of Id2 and Foxp3 in CD4<sup>+</sup>Foxp3<sup>+</sup> T<sub>reg</sub> cells sorted from PBS- and TetR- $Id2^{\text{EmGFP}}Foxp3^{\text{YFP-Cre}}$  mice. **b** CTV labelled naïve responder Dox-treated CD45.1<sup>+</sup>CD4<sup>+</sup>Foxp3<sup>-</sup> (upper panel) and CD45.1<sup>+</sup>CD8<sup>+</sup>Foxp3<sup>-</sup> (lower panel) T cells were cocultured with sorted CD45.2<sup>+</sup>CD4<sup>+</sup>Foxp3<sup>+</sup> T<sub>reg</sub> cells from PBS- or Dox-treated TetR-Id2<sup>EmGFP</sup>Foxp3<sup>YFP-Cre</sup> mice, in the presence of APCs purified from T cell depleted splenocytes. Responder T cell proliferation was analyzed by flow cytometry. c Intracellular cytokines produced by CD45.1<sup>+</sup>CD4<sup>+</sup>Foxp3<sup>-</sup> staining of (upper panel) or CD45.1<sup>+</sup>CD8<sup>+</sup>Foxp3<sup>-</sup> T cells (lower panel) that were co-cultured with PBS- or Dox-treated T<sub>reg</sub> cells at a ratio of 1:4 under conditions similar to (b). d Flow cytometry analysis of residual Foxp3 expression in Treg cells derived from PBS- or Dox-treated TetR- $Id2^{EmGFP}Foxp3^{YFP-Cre}$  mice that were used for suppression assay in (b). Data represents three independent experiments. NS=Non-Significant, \* P < 0.05, \*\* P < 0.005, (Student's *t*-test).



## Gating strategy for cell sorting

**a** Gating strategy used in **Figure 1** and **Figure 2** for sorting of CD4<sup>+</sup>CD25<sup>-</sup>CD62L<sup>hi</sup>CD44<sup>lo</sup> naïve T cells from Wild-type C57BL/6 (B6) mice to generate  $T_H0$ ,  $T_H17$ ,  $iT_{reg}$  and ex-Foxp3  $T_H17$  cells *in vitro*. Gating strategies for other experiments were described in each figure. **b** Gating strategy used in **Supplementary Figure 10** for *in vitro*  $T_{reg}$  suppression assay.



# Original western blot panels with gel makers for Figures 1c,1d and 1e.

Complete scanned gels for western blots shown in Figure 1c,1d and 1e. Western images are overlaid onto image containing MW markers. White dashed line identifies cropped region shown in respective figure.

# Supplementary Table 1: Primer sequences for RT-qPCR

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	
$ \begin{array}{ c c c c c c c } \hline R & GCTTTAATGTAATCCAGCAGGT \\ \hline T-bet (TBX21) & F & AGCAAGGACGGCGAATGTT \\ \hline R & GGGTGGACATATAAGCGGTTC \\ \hline Gata3 & F & CTCGGCCATTCGTACATGGAA \\ \hline R & GGATACCTCTGCACCGTAGC \\ \hline R & GGATACCTCTGCACCGTAGC \\ \hline R & AAGATCTGCAGCTTTCCATC \\ \hline R & AAGATCTGCAGCTTTTCCACA \\ \hline F & TTCCTTCCCAGAGTTCTTCC \\ \hline R & CTCAAATTCATCTACGGTCCA \\ \hline Id2 & F & TCATCGATTACATCTTGGACCT \\ \hline R & AATTCAGATGCCTGCAAGGA \\ \hline E2A & F & CATGCCCCGCCAG \\ \hline R & CAGTTCCTTGTCAGAGCCCA \\ \hline Ifn\gamma & F & GAGCCAGATTATCTTTTCACC \\ \hline R & GTTGTTGACCTCAAACTTGG \\ \end{array} $	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	
$ \begin{array}{ c c c c c } \hline Gata3 & R & GGATACCTCTGCACCGTAGC \\ \hline R & TTTGGAACTGGCTTTCCATC \\ \hline R & AAGATCTGCAGCTTTCCACA \\ \hline R & AAGATCTGCAGCTTTTCCACA \\ \hline F & TTCCTTCCCAGAGTTCTCC \\ \hline R & CTCAAATTCATCTACGGTCCA \\ \hline Id2 & F & TCATCGATTACATCTTGGACCT \\ \hline R & AATTCAGATGCCTGCAAGGA \\ \hline E2A & F & CATGCCCCGCCAG \\ \hline R & CAGTTCCTTGTCAGAGCCCA \\ \hline R & CAGTTCCTTGTCAGAGCCCA \\ \hline Ifny & F & GAGCCAGATTATCTCTTTCACC \\ \hline R & GTTGTTGACCTCAAACTTGG \\ \hline \end{array} $	
$ \begin{array}{ c c c c c c } \hline Roryt (Rorc) & R & AAGATCTGCAGCTTTTCCACA \\ \hline R & TTCCTTCCCAGAGTTCTTCC \\ \hline Foxp3 & F & TTCCTTCCAGAGTTCTACGGTCCA \\ \hline Id2 & F & TCATCGATTACATCTTGGACCT \\ \hline R & AATTCAGATGCCTGCAAGGA \\ \hline E2A & F & CATGCCCCGCCAG \\ \hline R & CAGTTCCTTGTCAGAGCCCA \\ \hline R & CAGTTCCTTGTCAGAGCCCA \\ \hline Ifny & F & GAGCCAGATTATCTCTTTCTACC \\ \hline R & GTTGTTGACCTCAAACTTGG \\ \hline \end{array} $	
$\frac{k}{F} = \frac{AAGAICIGCAGCITITCCACA}{F}$ $\frac{F}{R} = \frac{TTCCTTCCCAGAGTTCTTCC}{R}$ $\frac{Id2}{R} = \frac{F}{R} = \frac{TCATCGATTACATCTTGGACCT}{R}$ $\frac{F}{R} = \frac{CATGCCCCGCCAG}{R}$ $\frac{F}{R} = \frac{CATGCCCCGCCCAG}{R}$ $\frac{F}{R} = \frac{CAGTTCCTTGTCAGAGCCCA}{R}$ $\frac{F}{R} = \frac{GAGCCAGATTATCTCTTTCTACC}{R}$	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	
$ \begin{array}{c c c c c c } \hline R & CICAAAIICAICIACGGICCA \\ \hline Id2 & F & TCATCGATTACATCTTGGACCT \\ \hline R & AATTCAGATGCCTGCAAGGA \\ \hline E2A & F & CATGCCCCGGCCCAG \\ \hline R & CAGTTCCTTGTCAGAGCCCA \\ \hline Ifny & F & GAGCCAGATTATCTCTTTCTACC \\ \hline R & GTTGTTGACCTCAAACTTGG \\ \end{array} $	
Id2RAATTCAGATGCCTGCAAGGAE2AFCATGCCCCGCCCAGRCAGTTCCTTGTCAGAGCCCAIfnyFGAGCCAGATTATCTCTTTCTACCRGTTGTTGACCTCAAACTTGG	
$ \begin{array}{c c} E2A & R & CAGTTCCTTGTCAGAGCCCA \\ \hline Ifny & F & GAGCCAGATTATCTCTTTCTACC \\ R & GTTGTTGACCTCAAACTTGG \\ \end{array} $	
$\frac{R}{Ifn\gamma} = \frac{R}{R} = \frac{CAGTTCCTTGTCAGAGCCCA}{GAGCCAGATTATCTCTTTCTACC}$	
Ifny R GTTGTTGACCTCAAACTTGG	
R GIIGIIGACCICAAACIIGG	
F CAACGAAGAACACCACAGAG	
Il4 R GGACTTGGACTCATTCATGG	
F AGCCACTGGATTAGCTCAAGT	
<i>II5</i> R GGGGGTCACACATTTGCTTT	
F GCAACATCACAGGACCAGA	
Ill3 R GTCAGGGAATCCAGGGCTAC	
F TTCATCTGTGTCTCTGATGCT	
Ill7a R TTCATCTGTGTCTCTGATGCT	
F CCCATGGGATTACAACATCACTC	
Ill7f R CACTGGGCCTCAGCGATC	
F GTTGACACTTGTGCGATCTC	
II22 R CTGATCCTTAGCACTGACTCC	
F ATAACTGCACCCACTTCCCA	
Illo R TCATTTCCGATAAGGCTTGG	
F	
$Tgf-\beta$ R GCCTTAGTTTGGACAGGATCTG	
F CCCTCACACTCAGATCATCTTCT	
The R GCTACGACGTGGGCTACAG	