SUPPORTING INFORMATION

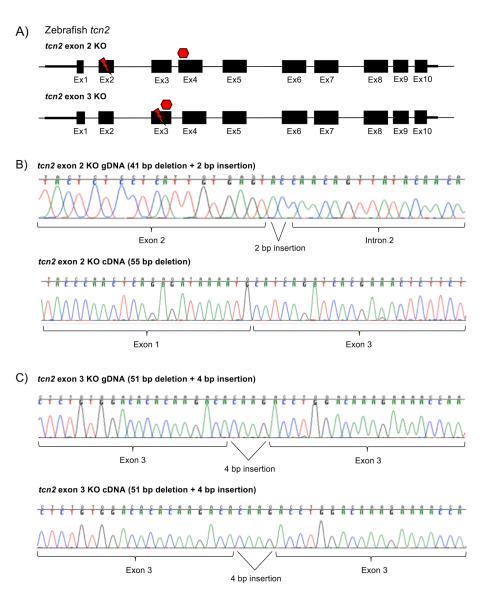


Figure S1. Design and characterization of CRISPR/Cas9 mutagenized $tcn2^{-t}$ **zebrafish lines.** (A) Two sgRNAs were designed to target either the second or the third exon out of ten in zebrafish tcn2. Exons and introns not to scale. Red hexagon demonstrates approximate location of early stop codon in the resulting transcript. (B) Chromatograms of gDNA and cDNA sequencing results for tcn2 exon 2 KO line. gDNA sequencing demonstrates the alternative splicing removal of exon 2 in the cDNA transcript, causing the resulting transcript to be out of frame. (C) Chromatograms of gDNA and cDNA sequencing results for tcn2 exon 3 KO line. gDNA and cDNA sequencing confirm a 51-bp deletion/4-bp insertion in exon 3, causing the resulting transcript to be out of frame.

CLUSTAL O(1.2.4) multiple sequence alignment

human TC	EMCEIPEMDSHLVEKLGQHLLPWMDRLSLEHLNPSIYVGLRLSSLQAG	48
human IF	TSTQTQSSCSVPSAQEPLVNGIQVLMENSVTSSAYPNPSILIAMNLAGAYNL	52
human_HC	EICEVSEENYIRLKPLLNTMIQSNYNRGTSAVNVVLSLKLVGIQIQ	46
zebrafish_Tcn2	KPCASDHETLLQSLNKQLLRSVDT-QDNLPNPSVHIALRLSTQHNL	45
zebrafish_Tcnba		0
zebrafish_Tcnbb		0
1		
human_TC	TKEDLYLHSLKLGYQQCLLGSAFSEDDGDCQGKPSMGQLALYLLALRANCEFVRGH	104 95
human_IF human HC	KAQKLLTYQLMSSDNNDLTIGQLGLTIMALTSSCRDPGDKVSI-	95 91
zebrafish Tcn2	TLMQKMIQQIKYNVKSRLSDVSSGELALIILAL-GVCRNAEENLIY- DKENQYLNRLKKEFHEDIEKSLRNGELVVGRLALYILALRSSCHDLSLHLNHN	98
zebrafish Tcnba		0
zebrafish_Tcnbb		0
human_TC	ee f -KGDRLVSQLKWFLEDEKRAIGHDHKGHPHTSYYQYGLGILALCLHQKRVHDSVVDKLLY	163
human_IF	LQRQMENWAPSSPNAEASAFYGPSLAILALCQKNSEATLPIAVR-F-	140
human HC	DYHLIDKLENKFQAEIENME-AHNGTPLTNYYQLSLDVLALCLFNGNYSTAEVVNHF-	147
zebrafish Tcn2	EKNEFLLTHLKKEMEEEKQNIAFSHRPKTNYYQYSLGILALCVSGVRVSTHVSHKLIH	156
zebrafish_Tcnba		0
zebrafish_Tcnbb		0
	a+g	
human_TC	AVEPFHQGHHSVDTAAMAGLAFTCLKRSNFNPGRRQRITMAIRTVREEILK	214
human_IF	AKTLLANSSPFNVDTGAMATLALTCMYNKIPVGSEEGYRSLFGQVLKDIVEKISM	195
human_HC	TPENKNYYFGSQFSVDTGAMAVLALTCVKKSLINGQIKADEGSLKNISIYTKSLVEKILS	207
zebrafish_Tcn2	AVEHGQIKHGESLCIDSHAMAGMALQCLKNEGISVKDEAELDKALATIKQKLVD	210
zebrafish_Tcnba		0
zebrafish_Tcnbb		0
	ag	
human_TC	AQTPEGHFGNVYSTPLALQFLMTSPMRGAELGTACLKARVALLASLQDGAFQNALMISQL	274
human_IF	KIKDNGIIGDIYSTGLAMQALSVTPE-PSKKEWNCKKTTDMILNEIKQGKFHNPMSIAQI	254
human_HC	EKKENGLIGNTFSTGEAMQALFVSSDYYNENDWNCQQTLNTVLTEISQGAFSNPNAAAQV	267
zebrafish_Tcn2	SKRADGHMGNEFSTGLAVQALLAMGVEMEECGTAIEALRGDIRKGTYHNPMAASQV	266 0
zebrafish_Tcnba zebrafish_Tcnbb		0
human_TC	LPVLNHKTYIDLIFPDCLAPRVMLEPAAETIPQTQEIISVTLQVLSLLPP	324
human_IF	LPSLKGKTYLDVPQVTCSPDHEVQPTLPSNPGPGPTSASNITVIYTINNQLRGVEL	310
human_HC	LPALMGKTFLDINKDSSCVSASGNFNISADEPITV-TPPDSQSYISVNYSVRINET	322
zebrafish_Tcn2	LPALYQQSYLHLKSKECRSEDDTLTADVESASEVLPSLGQV-AVQVEVIKSNGEA	320 25
zebrafish_Tcnba	EIPVKVTIVNDFTN-EQ GQVSINVVVTNKFAN-EL	25
zebrafish_Tcnbb	GQV5INVVVINAAN-EL : :	20
	d	
human_TC	YRQSISVLAGSTVE-DVLKKAHEL-GGFTYET-QASLSGPYLTSVMGKAAGEREF	376
human_IF	LFNETINVSVKSGSVLLVVLEEAQRK-NPMFKFET-TMTSWGLVVSSINNIAENVNHKTY	368
human_HC	YFTNVTVLNGSVFLSVMEKAQKMNDTIFGFTM-EERSWGPYITCIQGLCANNNDRTY	378
zebrafish_Tcn2	SVFPINVPKGSSLFEALNLLQDKQ-TGFTFKT-EDSLWGAFLSVLNDEQARQTDRRY	375
zebrafish_Tcnba	$\texttt{L}{=}{-}\texttt{SYSTTVIQEGLMFGVLNQLMESN}{-}\texttt{ADFKFSYTIHHTFGIY}\texttt{LESVNGLAGSDEDQTY}$	81
zebrafish_Tcnbb	NTYPVTAPKGMPIFGVLNQLQDSNQLNFTYSISKSYGI ^F LESVNGLAGSTENKT ^Y : : : * :::	83
human_TC	d c cb WQLLRDPNTPLLQGIADYRPKDGETIELRLVSW 409	
human_IF	WQFLSGVTPLNEGVADYIPFNHEHITANFTQY 400	
human_HC	WELLSGGEPLSQGAGSYVVRNGENLEVRWSKY 410	
zebrafish_Tcn2	WHVSSDGTSLTQGIKDYKIDSAQRITIKNTGY 407	
zebrafish_Tcnba	WELLSEKSGVVTRLEVGIGCYQVQRDENLILRFTTWATKK 121	
zebrafish_Tcnbb	WELLSKRERKTTRLNVGIGCYQPERNENFIMNFTTWA 120	
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Figure S2. Multiple sequence alignment of known and potential cobalamin carrier proteins in humans and zebrafish. A multiple sequence alignment for human TC, IF, and HC and zebrafish Tcn2, Tcnba and Tcnbb was performed using Clustal-Omega software. Specific annotations were added according to findings in (1). Blue lettering indicates residues involved in hydrophobic interactions with cobalamin, while letters above the alignment indicate residues involved in hydrogen bonds to a specific side chain of cobalamin (side chain indicated by the letter shown). Dots below alignment indicate levels of local residue conservation, while asterisks signify complete residue conservation within this set of sequences. Red bars below alignment indicate regions targeted for site-directed mutagenesis.

Table S1. **Cobalamin carrier protein homologs in vertebrate species.** Species name, prepended organism abbreviation, common name, lineage, and accession code are notated for each sequence included in Fig 2. Further divisions based on homolog classification are shown in alternate sheets.

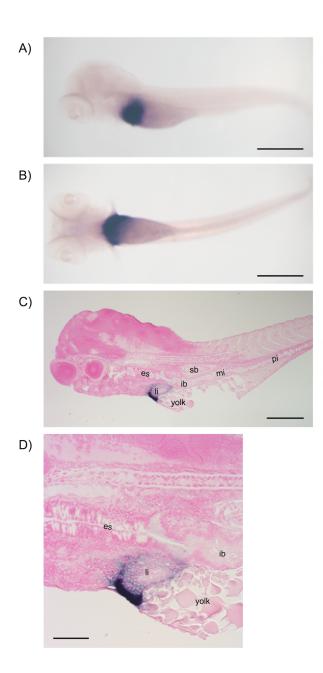


Figure S3. Whole-mount *in situ* hybridization and sectioning of liver-specific probe *fabp10a*. Wholemount *in situ* hybridizations were performed on zebrafish embryos at 5 dpf with a DIG-labelled RNA probe and subsequent treatment with an anti-DIG antibody conjugated to alkaline phosphatase (AP) and BMpurple staining for spatial comparison of the known liver-specific *fabp10a* with our genes of interest (A-B). Embryos were then sectioned and stained with Nuclear Fast Red (NFR) to visualize at higher resolution (C-D). (A) Lateral view of *fabp10a* expression. Scale bar: 400 µm. (B) Ventral view of *fabp10a* expression. Scale bar: 400 µm. (C) Sagittal section of *fabp10a*. Scale bar: 200 µm. (D) Zoom-in of the sagittal section of *fabp10a*. Scale bar: 50 µm. Abbreviations: es = esophagus, li = liver, ib = intestinal bulb, sb = swim bladder, mi = mid-intestine, pi = posterior intestine.

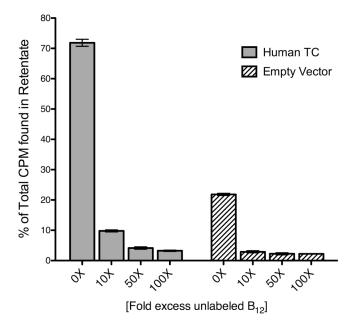


Figure S4. Competition assay demonstrates binding specificity of radiolabeled cobalamin. Specific activity (191 μ Ci/ μ g) of ⁵⁷Co-labelled vitamin B₁₂ solution was used to calculate input concentration of radiolabeled cobalamin used in binding assay experiments (145 pM). Media samples (Human TC and empty vector) were incubated with ⁵⁷Co-containing cobalamin and varying concentrations of excess unlabeled cobalamin competitor (0X, 10X, 50X, 100X). Samples were spun through 10 kDa filters to separate protein-bound and unbound radioactive cobalamin. Percent of total counts per minute (CPM) in the retentate (protein-bound fraction) are shown, ± SD (n=3).

REFERENCES

1. Wuerges, J., Geremia, S., and Randaccio, L. (2007) Structural study on ligand specificity of human vitamin B12 transporters. *Biochem J* **403**, 431-440