

# Supplementary Data

For

## Structural insights into DNA degradation by human mitochondrial nuclease MGME1

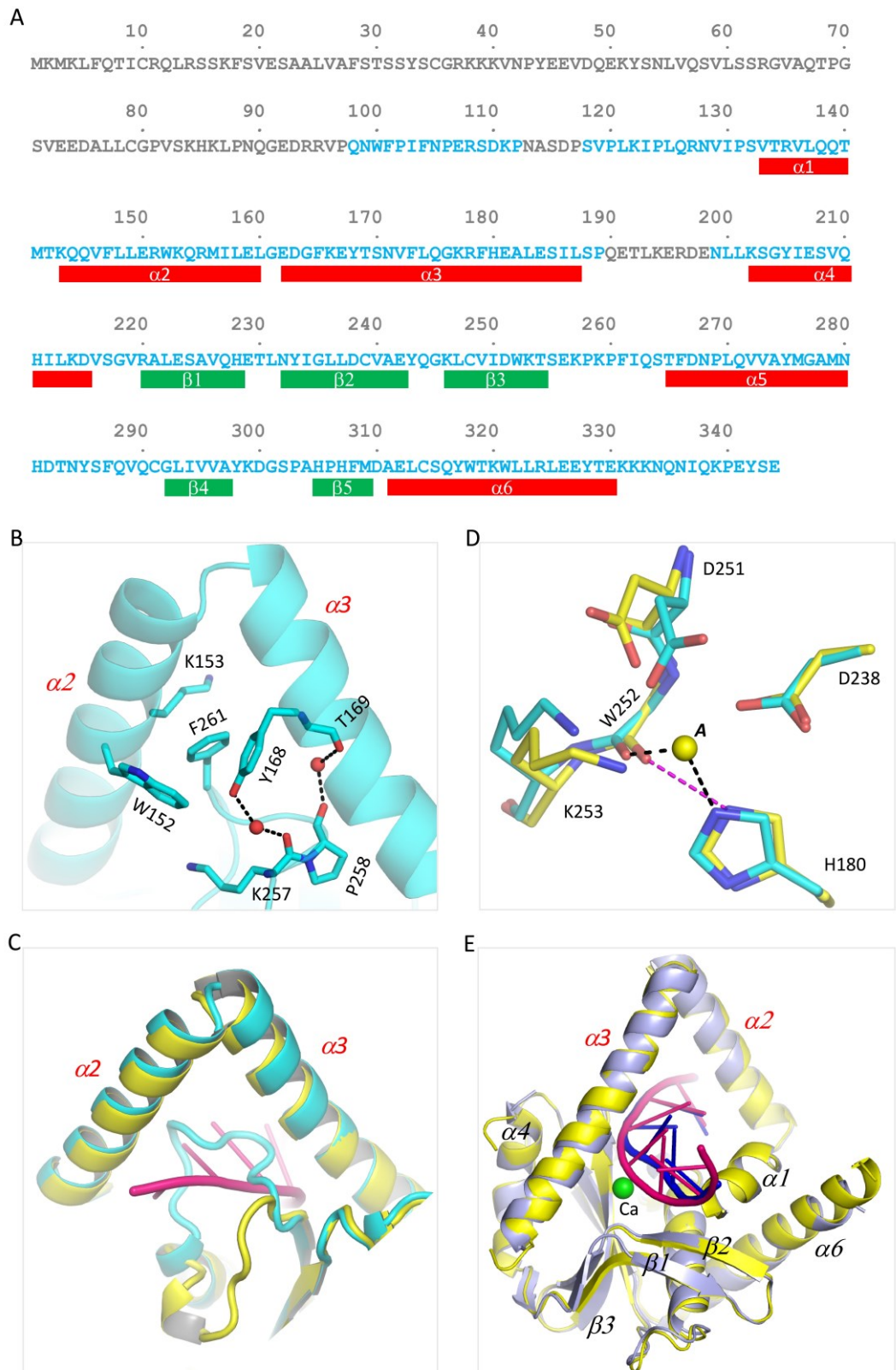
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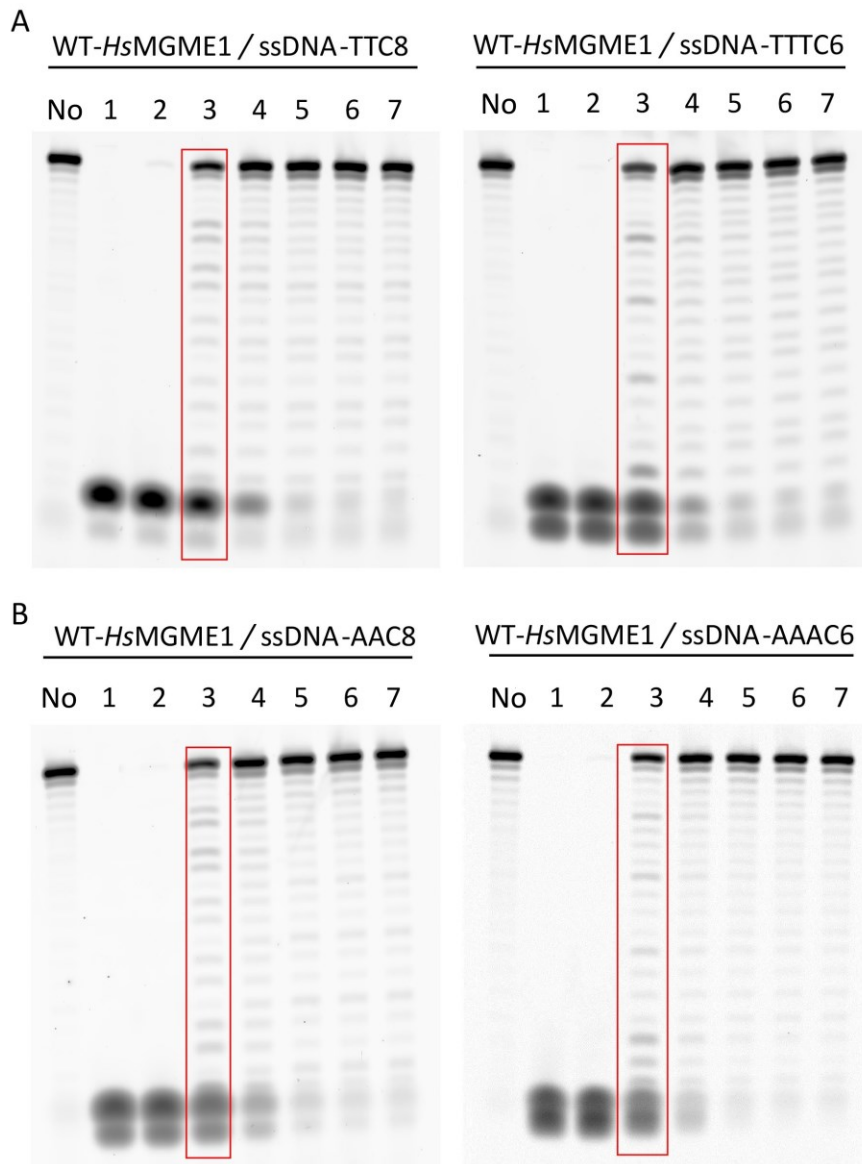
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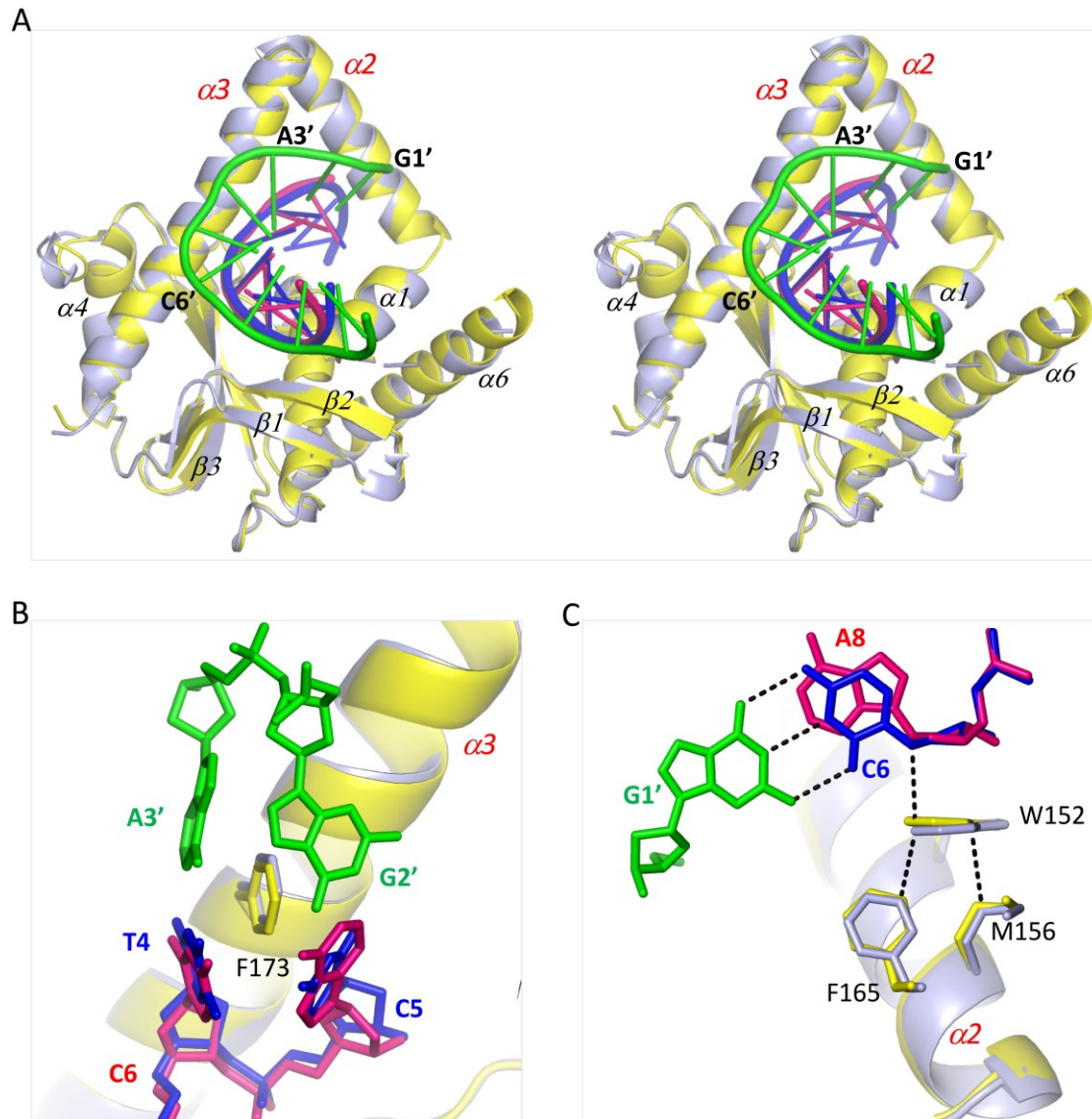


**Supplementary Figure S1. Conformational changes of *Hs*MGME1.** (A) Sequence and secondary structure of *Hs*MGME1. Residues disordered in *Hs*MGME1-Mn<sup>2+</sup> complex are colored in gray. (B) Detailed conformations of the helical arch and  $\beta$ 3- $\alpha$ 5

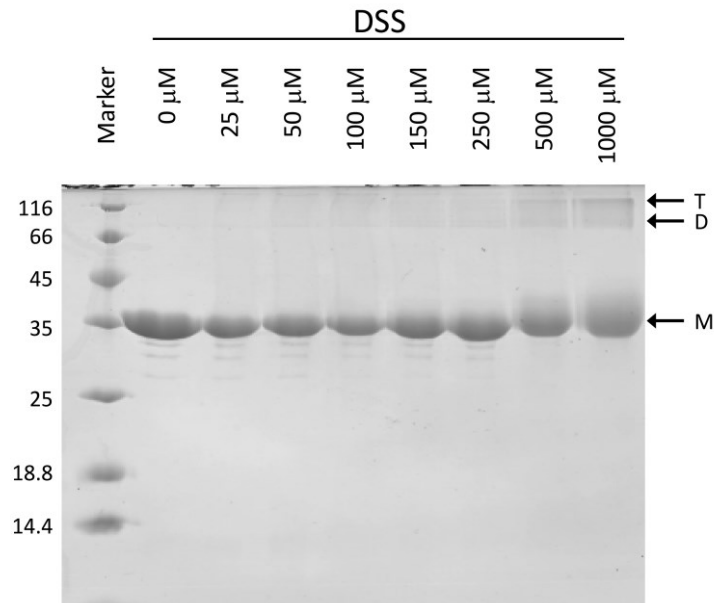
connecting loop observed in the *HsMGME1*-Mn<sup>2+</sup> complex. **(C)** Superposition of *HsMGME1*-Mn<sup>2+</sup> and *HsMGME1*-ssDNA2 complexes showing the conformational change of the  $\beta$ 3- $\alpha$ 5 connecting loops. **(D)** Comparison of the catalytic site residues of *HsMGME1*-Mn<sup>2+</sup> and *HsMGME1*-ssDNA2. **(E)** Superposition showing the local conformational difference of the DNAs, which are shown as magenta and blue cartoons in *HsMGME1*-ssDNA2 and H180Q-ssDNA2-Ca<sup>2+</sup> complexes, respectively. Water molecules, Mn<sup>2+</sup> and Ca<sup>2+</sup> are shown as red, yellow, and green spheres, respectively.



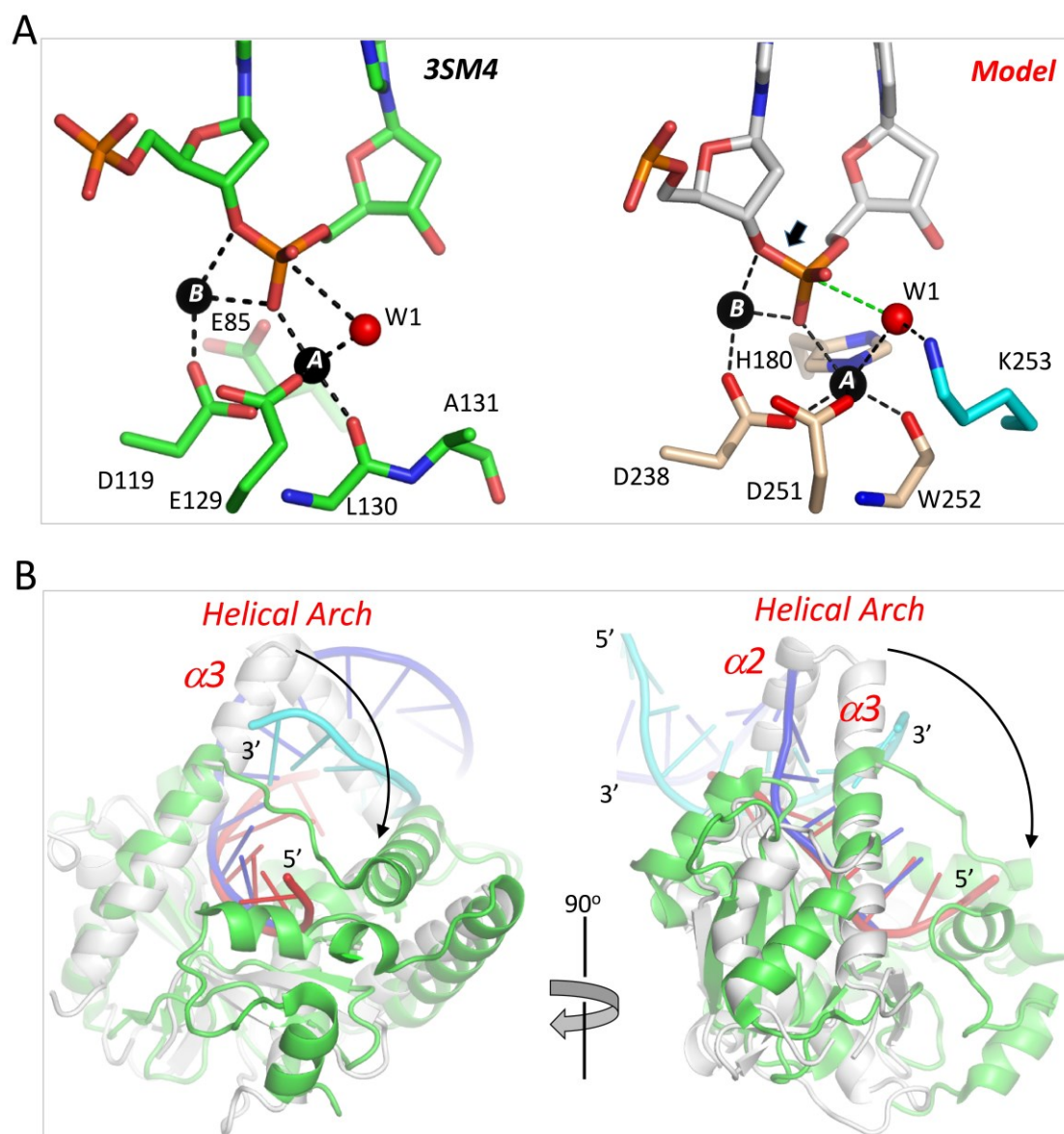
**Supplementary Figure S2. The weak cleavage site preference of *HsMGME1*.** (A) *in vitro* ssDNA-TTC8 and ssDNA-TTTC6 cleavage by WT *HsMGME1*. (B) *in vitro* ssDNA-AAC8 and ssDNA-AAAC6 cleavage by WT *HsMGME1*. DNA concentrations are 0.8  $\mu$ M. Protein concentrations are 0.8  $\mu$ M, 0.4  $\mu$ M, 0.2  $\mu$ M, 0.1  $\mu$ M, 0.05  $\mu$ M, 0.025  $\mu$ M, and 0.0125  $\mu$ M in lanes 1-7, respectively. *HsMGME1* is absent in the lane labelled with No.



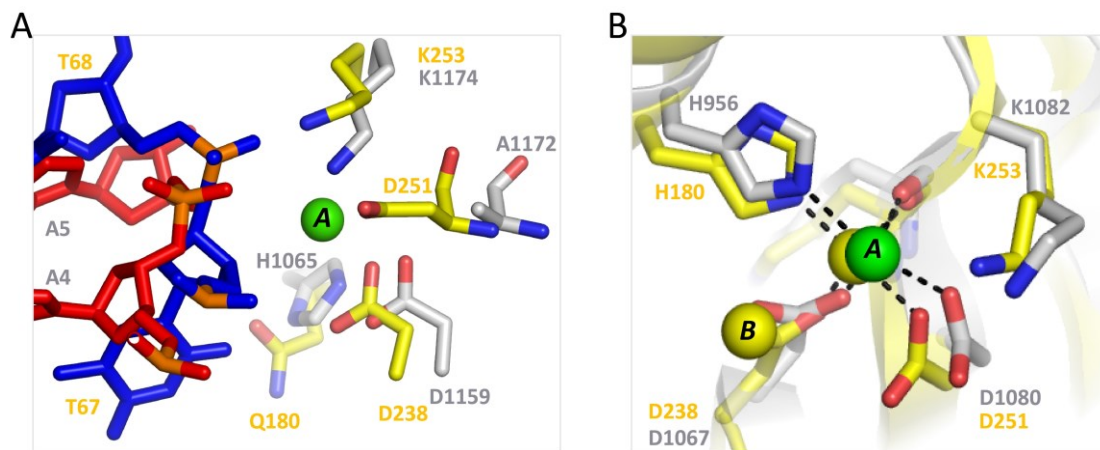
**Supplementary Figure S3. Comparison of single-stranded and duplex DNA bound *HsMGME1*.** (A) Stereoview showing the comparison of *HsMGME1*-ssDNA2 and *HsMGME1*-DNA3 complexes. (B) and (C) Conformational comparison of the pin residues and the surrounding nucleotides. For *HsMGME1*-DNA2, the protein and DNA are colored in yellow and magenta. For *HsMGME1*-DNA3, the protein and the two DNA strands are colored in light-blue, green, and blue, respectively. The sequences of ssDNA2 (5'-AACACAACAAC-3') and DNA3 (5'-GGATCCTTCTTCTTCTC -3') have no coincidence.



**Supplementary Figure S4. DSS crosslinking analysis of *HsMGME1*.** Protein and DNA3 concentrations are 50  $\mu\text{M}$  and 50  $\mu\text{M}$ , respectively. DSS concentrations are indicated on the gel. The bands corresponding to the monomer, dimer, and tetramer of *HsMGME1* are indicated by arrows and labelled as M, D, and T, respectively.



**Supplementary Figure S5. Structural comparison between *HsMGME1* and phage lambda nuclease.** (A) Comparison of the active site structure of the K131A mutant of phage lambda nuclease (PDB\_ID: 3SM4) and the catalytic MGME1 model we proposed. Cations and the catalytic water molecule are shown as spheres in black and red, respectively. (B) Structural superposition showing the flexibility of the regions corresponding to the helical arch of *HsMGME1*. *HsMGME1* and the phage lambda nuclease are colored in white and green, respectively. DNA is colored in red in *HsMGME1*-DNA2 structure, whereas it is colored in blue and cyan in the phage lambda nuclease structure.



**Supplementary Figure S6. Structural comparison between *HsMGME1* and RecB-type nucleases.** (A) Structural superposition showing the different conformations of DNAs near the cleavage sites of H180Q-ssDNA2-Ca<sup>2+</sup> and AddAB-DNA complexes. Ca<sup>2+</sup> is shown as green sphere in H180Q-ssDNA2-Ca<sup>2+</sup>. The DNA C-atoms are colored in red and blue in the H180Q-ssDNA2-Ca<sup>2+</sup> and the AddAB-DNA complexes, respectively. *HsMGME1* H180Q mutant and RecB are shown as sticks with their C-atoms colored in yellow and white, respectively. (B) Structural superposition showing the similar coordination of cations bound in the active sites of the *HsMGME1*-Mn<sup>2+</sup> and the RecBCD-DNA complexes. Mn<sup>2+</sup> of *HsMGME1*-Mn<sup>2+</sup> and Ca<sup>2+</sup> of RecBCD-DNA are shown as spheres in yellow and green, respectively. *HsMGME1* and D1172A mutant of AddA are all shown as sticks with their C-atoms colored in yellow and white, respectively.



**Supplementary Table S1.** Sequences of the primers used for WT and mutant *HsMGME1* construction

Name	Sequence <sup>a</sup>
Full-length MGME1-F	CGC <u>GGATCC</u> GGTGGTGGTATGAAGATGAAGTTATTTTCAGACC ATTTGCAGGCAG
WT-F	CGC <u>GGATCC</u> GAATCAGCTGCCCTTGTGGC
delN90-F	CGC <u>GGATCC</u> GGTGAGGACAGACGAGTGCCAC
delN130-F	CGC <u>GGATCC</u> GGTGGTGGTATACCAAGTGTGACCCGAGTCCTT CAGC
WT-R	AA <u>ACTCGAG</u> CTATTCTGAATATTCTGGTTTCTGAATATTC
WT-delC-R	AA <u>ACTCGAG</u> CTATTTCTTTCCGTATATTCTTCTAGTC
T134A-F	GTGATACCAAGTGTGGCCCGAGTCCTTCAGCAG
T134A-R	CTGCTGAAGGACTCGGGCCACACTTGGTATCAC
Q145A-F	CCATGACAAAACAAGCTGTTTTCTTGTGGAGAGG
Q145A-R	CCTCTCCAACAAGAAAACAGCTTGTGTTGTCATGG
W152A-F	GTTTTCTTGTGGAGAGGGCTAAACAGCGGATG
W152A-R	CATCCGCTGTTTAGCCCTCTCCAACAAGAAAAC
F173A-F	GAATACACTTCAAACGTCGCTTTACAAGGGAAACGGTTC
F173A-R	GAACCGTTTCCCTTGTAAGCGACGTTTGAAGTGTATTC
H180Q-F	GGGAAACGGTTCCAAGAAGCCTTGGAAAG
H180Q-R	CTTTCCAAGGCTTCTTGAACCGTTTCCC
E184Q-F	CCACGAAGCCTTGCAAAGCATACTTTCACCCC
E184Q-R	GGGGTGAAAGTATGCTTTGCAAGGCTTCGTGG
E223Q-F	GGAGTGCGAGCTCTTCAAAGTGCTGTTCAACATG
E223Q-R	CATGTTGAACAGCACTTTGAAGAGCTCGCACTCC
D238N-F	CTATATAGGTCTGCTGAACTGTGTGGCTGAGTATC
D238N-R	GATACTCAGCCACACAGTTCAGCAGACCTATATAG
D251N-F	GCAAGCTCTGTGTGATTAATTGGAAGACATCAGAG
D251N-R	CTCTGATGTCTTCCAATTAATCACACAGAGCTTGC
T254A-F	GTGTGATTGATTGGAAGGCTTCAGAGAAACCAAAG
T254A-R	CTTTGGTTTCTCTGAAGCCTTCCAATCAATCACAC
F266A-F	GCCTTTTATTCAAAGTACAGCTGACAACCCACTGC
F266A-R	GCAGTGGGTTGTGACAGCTGTACTTTGAATAAAAGGC
Q271A-F	GACAACCCACTGGCTGTTGTGGCATAACATGGG
Q271A-R	CCCATGTATGCCACAACAGCCAGTGGGTTG TC
Y275A-F	CTGCAAGTTGTGGCAGCTATGGGTGCCATGAAC
Y275A-R	GTTTCATGGCACCCATAGCTGCCACAACCTTGCAG

<sup>a</sup>GGATCC and CTCGAG highlighted with underline are BamHI and XhoI recognition sequence.

**Supplementary Table S2.** Data collection and refinement statistics <sup>a</sup>

Structure (PDB ID)	MGME1-Mn <sup>2+</sup> 5ZYW	MGME1-DNA2 5ZYU	H180Q-DNA2-Ca <sup>2+</sup> 5ZYV	MGME1-DNA3 5ZYT	Se-MGME1- DNA3
<b>Data collection<sup>a</sup></b>					
Space group	P4 <sub>3</sub> 2 <sub>1</sub> 2	P3 <sub>2</sub>	P2 <sub>1</sub> 2 <sub>1</sub> 2	C2	C2
Cell parameter:					
a (Å)	68.0	80.7	81.3	146.5	146.3
b (Å)	68.0	80.7	106.2	77.5	78.2
c (Å)	114.8	79.8	33.4	164.7	165.7
β (°)	90	90.0	90.0	112.4	112.4
Wavelength (Å)	0.9793	0.9793	0.9793	0.9793	0.9793
Resolution (Å)	30.0-2.20	30.0-1.75	30.0-2.70	30.0-2.70	30.0-3.0
Last shell (Å)	2.28-2.20	1.81-1.75	2.80-2.70	2.80-2.70	3.05-3.0
Completeness (%)	100.0(100.0)	100.0(100.0)	97.6(86.8)	94.6(90.6)	99.9(100.0)
Redundancy	21.6(14.8)	9.6(8.0)	7.5(3.4)	5.6(5.1)	6.6(6.5)
I/σ(I)	31.0(2.5)	16.4(2.3)	17.3(2.7)	15.4(2.2)	16.0(2.1)
Rmerge (%)	10.5(48.0)	9.4(47.7)	9.5(35.6)	9.4(46.4)	12.7(69.5)
<b>Refinement</b>					
Resolution (Å)	29.9-2.20	28.4-1.75	28.3-2.72	30.0-2.70	
R <sub>work</sub> (%) / R <sub>free</sub> (%)	22.6/25.2	17.4/20.0	21.2/25.4	25.0/29.6	
No. of atoms					
Protein	1854	3541	1555	7164	
DNA		290	126	872	
Cations	3		2		
water	94	532	13		
R.m.s. deviations					
Bond length (Å)	0.007	0.011	0.004	0.010	
Bond angle (°)	1.113	1.036	0.613	1.250	
Ramachandran plot (%)					
Most favored	98.7	98.6	95.3	96.0	
Additional allowed	1.3	1.4	4.7	4.0	

<sup>a</sup>: Values in parentheses are for the last resolution shell.

**Supplementary Table S3.** Sequences of DNAs used in crystallization and *in vitro* cleavage assays

Name	Sequence (5'-3')
<b>DNAs for crystallization</b>	
ssDNA2	AACAACAACAAC
DNA3	<u>GGATCCTTCTTCTTCTTC</u>
<b>DNAs for <i>in vitro</i> cleavage assay <sup>a</sup></b>	
ssDNA1	TTCTTCTTCTTC
ssDNA2	AACAACAACAAC
dsDNA4	<u>GGATGGGGGATCCCCCATCC</u>
5'-overhang DNA4	TTCTTCTTCTTC <u>CGGATGGGGGATCCCCCATCC</u>
3'-overhang DNA4	<u>GGATGGGGGATCCCCCATCC</u> TTCTTCTTCTTC
ssDNA-TTC8	TTCTTCTTCTTCTTCTTCTTC
ssDNA-TTTC6	TTTCTTTCTTTCTTTC
ssDNA-AAC8	AACAACAACAACAACAACAACAAC
ssDNA-AAAC6	AAACAAACAAACAAACAAACAAAC

<sup>a</sup>: Except 5'-overhang DNA4 that was FAM-labelled at the 3'-end, all other DNAs are FAM-labelled at their 5'-ends. The sequences that can form self-complementary duplexes are highlighted with underlines.