Supporting Information For

Quasi-Resonance Signal Amplification By Reversible Exchange

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Table of Contents

1.	Computations of ¹⁵ N signal enhancement and polarization levels	S-:	3
2.	Details of shaped SLIC RF pulse preparation	S-4	4
3.	References Used in Supporting Information	S-(6

1. Computations of ¹⁵N signal enhancement and polarization levels

Computation of signal enhancements was very challenging at 0.05 T, because even highly concentrated sample would not provide any signal. Therefore, the signal enhancements were computed using lower limit estimate as the following.

<u>Metronidazole-¹⁵N₂-¹³C₂</u>: The highest SNR achieved was 1,270 at 20 mM concentration (Figure 4c). The concentration of ¹⁵N spins in signal reference sample (imidazole-¹⁵N₂ prepared as described previously¹) was ~8 M in D₂O. The reference sample was placed in conventional standard-wall 5 mm NMR tube. Note the reference sample contain two labeled ¹⁵N sites. SNR of at least 2 is required to see any signal, but no signal was observed with 16 scans (square root of 16 is 4 for SNR purpose calculation). 1.85 is a factor related to different effective cross-section of the NMR tubes in HP and thermally polarized experiments.²

Assuming that both ¹⁵N sites were polarized by OUASR-SABRE in metronidazole-¹⁵N₂-¹³C₂: we arrive to

 $\varepsilon_{15N} = ((\sqrt{\#} \text{ of scans reference})/(\sqrt{\#} \text{ of scans HP sample}))*((HP SNR)/(SNR))$

Reference))*1.85*([reference]/[hyperpolarized sample])*((# of ¹⁵N sites per molecule in reference sample)/(# of ¹⁵N sites per molecule in HP sample)) = $(\sqrt{16}/\sqrt{8})^{*}(1,700/2)^{*}1.85^{*}[8/0.02]^{*}[2/2] = 9.0^{*}10^{5}$

Note this is the lower limit of signal enhancements, because we do not know how low thermal reference truly is. 15 N thermal polarization at 49 mT and 298 K is $1.7*10^{-6}$ %.

Therefore, $\sqrt[6]{P_{15N}} = \sqrt[6]{P_{thermal}} \epsilon_{15N} = 1.7 \times 10^{-6} \times 9.0 \times 10^{5} = 1.5\%$

However, if only one ¹⁵N site was polarized, the number below would be effectively doubled!

The integral signal value in Figure 4a (SABRE-SHEATH) is **0.162**. The integral signal value in display Figure 4c (OUASR-SABRE) is 0.394.

Next, using integral values we arrive to the following numbers for SABRE-SHEATH:

 $\varepsilon_{15N} = 9.0*10^{5}*(0.162/0.394) = 3.7*10^{5}$ with % $P_{15N} \sim 0.6\%$

One again, these are the lower limit numbers.

Note, under similar conditions, the average ¹⁵N polarization using high-field NMR spectroscopy for SABRE-SHEATH on metronidazole-¹⁵N₂-¹³C₂ was 1.4%.³ These numbers are somewhat similar, which is a good news.

The efficiency defined as S(QUASR-SABRE)/S(SABRE-SHEATH) = 2.43!

Note if we are polarizing only one site via OUASR-SABRE versus two ¹⁵N sites in SABRE-SHEATH, the

efficiency is doubled to 4.86!

<u>Pyridine-</u>¹⁵N: Using the numbers above we can also compute lower-limit values for ¹⁵N signal enhancement and polarization values for pyridine-¹⁵N for both SABRE-SHEATH and QUASR-SABRE (since the integral numbers are similar: 0.143 and 0.145 for Figure 2a and Figure 2c respectively):

 $\varepsilon_{15N} = ([metronidazole^{-15}N_2^{-13}C_2]/[pyridine^{-15}N])*((\# of^{-15}N sites in metronidazole^{-15}N_2^{-13}C_2)/(\# of^{-15}N sites in metronidazole^{-15}N sites$ in pyridine-¹⁵N))*((integral signal value of pyridine-¹⁵N)/(integral signal value of metronidazole-¹⁵N₂-¹³C₂))* ε_{15N} (metronidazole-¹⁵N₂-¹³C₂) = (20/20)*(2/1)*(0.145/0.394)*9*10⁵ = 6.6*10⁵ with %P_{15N} ~ 1.1%. Again, this is not far from what is expected in this concentration range.⁴⁻⁵

<u>Acetonitrile-¹⁵N</u>: Using the numbers above we can also compute low-limit values for ¹⁵N signal enhancement and polarization values for acetonitrile-¹⁵N for both SABRE-SHEATH (Figure 3a, signal integral value of **0.0525**) in a manner similar to that for pyridine-¹⁵N calculation detailed above:

 $\varepsilon_{15N} = (20/40)^*(2/1)^* (0.0525/0.394)^*9.0^*10^5 = 1.2^*10^5$ with % $P_{15N} \sim 0.2\%$.

And for and QUASR-SABRE (Figure 3c, signal integral value of **0.0230**):

 $\varepsilon_{15N} = (20/40)^*(2/1)^* (0.023/0.394)^*9.0^*10^5 = 5.3^*10^4$ with % $P_{15N} \sim 0.09\%$.

2. Details of shaped SLIC RF pulse preparation

The rectangular ¹⁵N RF pulse was calibrated on a hyperpolarized sample prepared via SABRE-SHEATH approach at 210.0 kHz resonance frequency. The calibration yielded a value of $t_{90^\circ} = 260 \ \mu s$ at -33 db of power setting. This value corresponded to ~ 960 Hz of B₁ power or ($\omega_1/2\pi$). The TOMCO RF amplifier was deemed to be linear all the way to -48 db (Figure S4c). As a result, the power setting employed (-40 db for all experiments) had a B₁ power of (960/2.24) = 430 Hz.

The SLIC pulse was designed in 100 equally spaced steps with the amplitude starting from 1.00, 0.99, 0.98,..., 0.01 as a table. The shape of the RF pulse was tested on the oscilloscope. We note a minor droop in power at the end of the shaped pulse related to the RF amplifier non-linearity at very low power settings.



Figure S1. The photograph of two shaped pulse on the digital oscilloscope.



Figure S2. Supplemental pyridine-¹⁵N data. The experimental conditions were the same as those using to obtain the data shown in Figure 2. a) ¹⁵N QUASR-SABRE signal dependence on the duration of the shaped pulse; g) ¹⁵N QUASR-SABRE signal dependence on the duration of the delay. Note the individual spectra employed for figures in displays a and b were auto-phased, and the data is presented in the magnitude mode.



Figure S3. Supplemental acetonitrile-¹⁵N data. The experimental conditions were the same as those using to obtain the data shown in Figure 3. a) ¹⁵N QUASR-SABRE signal dependence on the duration of the shaped pulse; b) ¹⁵N QUASR-SABRE signal dependence on the duration of the shaped pulse; c) ¹⁵N QUASR-SABRE signal dependence on the duration in displays a and b.



Figure S4. Supplemental metronidazole- ${}^{15}N_2 - {}^{13}C_2$ data. The experimental conditions were the same as those using to obtain the data shown in Figure 4. a) ${}^{15}N$ QUASR-SABRE signal dependence on the duration of the shaped pulse; b) ${}^{15}N$ QUASR-SABRE signal dependence on the duration of the delay; c) ${}^{15}N$ QUASR-SABRE signal dependence on the applied radio frequency power. Note the individual spectra employed for figures in all three displays were auto-phased, and the data is presented in the magnitude mode.

3. References Used in Supporting Information

1. Shchepin, R. V.; Barskiy, D. A.; Coffey, A. M.; Feldman, M. A.; Kovtunova, L. M.; Bukhtiyarov, V. I.; Kovtunov, K. V.; Goodson, B. M.; Koptyug, I. V.; Chekmenev, E. Y. Robust Imidazole-¹⁵N₂ Synthesis for High-Resolution Low-Field (0.05 T) ¹⁵N hyperpolarized Nmr Spectroscopy. *ChemistrySelect* **2017**, *2*, 4478–4483.

2. Truong, M. L.; Shi, F.; He, P.; Yuan, B.; Plunkett, K. N.; Coffey, A. M.; Shchepin, R. V.; Barskiy, D. A.; Kovtunov, K. V.; Koptyug, I. V., et al. Irreversible Catalyst Activation Enables Hyperpolarization and Water Solubility for NMR Signal Amplification by Reversible Exchange. *J. Phys. Chem. B* **2014**, *18* 13882–13889.

3. Shchepin, R. V.; Jaigirdar, L.; Chekmenev, E. Y. Spin-Lattice Relaxation of Hyperpolarized Metronidazole in Signal Amplification by Reversible Exchange in Micro-Tesla Fields. *J. Phys. Chem. C* **2018**, *122*, 4984–4996.

4. Theis, T.; Truong, M. L.; Coffey, A. M.; Shchepin, R. V.; Waddell, K. W.; Shi, F.; Goodson, B. M.; Warren, W. S.; Chekmenev, E. Y. Microtesla Sabre Enables 10% Nitrogen-15 Nuclear Spin Polarization. *J. Am. Chem. Soc.* **2015**, *137*, 1404-1407.

5. Truong, M. L.; Theis, T.; Coffey, A. M.; Shchepin, R. V.; Waddell, K. W.; Shi, F.; Goodson, B. M.; Warren, W. S.; Chekmenev, E. Y. ¹⁵N Hyperpolarization by Reversible Exchange Using SABRE-SHEATH. *J. Phys. Chem. C* **2015**, *119*, 8786–8797.