

Figure S1, Related to Figure 1. Immune cell populations in the spleen from pups born to ZIKV-immune mothers and naïve mothers. Spleens were harvested from 4- to 5-week-old *LysMCre+Ifnar1*^{fl/fl} mice born to mothers infected with ZIKV strain SD001 (10⁶ FFU via retro-orbital route) for 8 months (*n*=2) or naïve mothers (*n*=1). Splenocytes were analyzed for CD3, CD4, CD8, CD19, CD138, and IgD expression via flow cytometry. (**A**) Total number of live splenocytes, (**B**) Number of CD3⁺ splenocytes, (**C**) Frequency of CD3⁺ splenocytes, (**D**) Number of CD4⁺ T cells, (**E**) Frequency of CD4⁺ T cells (**F**) Number of CD8⁺ T cells, (**G**) Frequency of CD8⁺ T cells, (**H**) Number of CD19⁺ B cells, (**I**) Frequency of CD19⁺ B cells, (**J**) Number of CD19⁺CD138⁺IgD⁻ plasma cells, (**K**) Frequency of CD19⁺CD138⁺IgD⁻ plasma cells. An unpaired student's t test was performed and no statistically significant differences were observed.



Figure S2, Related to Figures 1. Decreased ZIKV tissue burden and ZIKV-induced clinical disease manifestations in mice born to ZIKV-immune mothers but not DENV-immune mothers. Four- to 5-week-old *LysMCre*⁺*Ifnar1*^{*fl/fl*} mice born to mothers previously infected with ZIKV strain SD001 (10^6 FFU via retro-orbital route), mothers previously infected with DENV2 strain S221 (5×10^5 FFU via tail vein), or naïve mothers were challenged via retro-orbital route with 10^5 FFU of ZIKV strain SD001. (**A**) Mice were sacrificed 3 days p.i. and serum, spleen, liver, brain, and eyes were harvested. Samples were titrated for infectious ZIKV via focus forming assay. Open squares represent pups

(*n*=7) from naïve mothers (*n*=2), closed circles represent pups (*n*=7) born to ZIKV-immune mothers infected for 2 months (*n*=2), and closed triangles represent pups (*n*=7) born to DENV-immune mothers infected for 2-3 months (*n*=2). Dotted line indicates limit of detection. (**B**) Clinical scores of pups (*n*=7) from naïve mothers (*n*=2), (**C**) Clinical scores of pups (*n*=6) born to ZIKV-immune mothers infected for 6-7 months (*n*=2), (**D**) Clinical scores of pups (*n*=7) from naïve mothers (*n*=2), (**E**) Clinical scores of pups (*n*=8) born to DENV2-immune mothers infected for 2-3 months (*n*=2), (**F**) Weight loss of pups born to ZIKV-immune mothers, and (**G**) Weight loss of pups born to DENV-immune mothers. Data represent pooled data from two independent experiments and are expressed as mean ± standard error of mean. One-way ANOVA was used for all comparisons in (**A**). Unpaired Student's t test of groups for each day in (**F**) and (**G**). **P* < 0.05, ** *P* < 0.01, **** *P* < 0.001, **** *P* < 0.0001.



Figure S3, Related to Figures 1 and 2. Mice born to DENV-immune mothers have reduced DENV disease compared to mice born to naïve mothers, and sera from mice born to DENV-immune mothers do not bind ZIKV. Four- to 5-week-old *LysMCre*⁺*Ifnar1*^{#/#} mice born to mothers previously infected with DENV2 strain S221 ($5x10^5$ FFU via tail vein) or to naïve mothers were challenged with 10^6 FFU of DENV2 strain S221 via tail vein injection. (**A**) Clinical scores of pups (*n*=6) from naïve mothers (*n*=2), Clinical scores of pups (*n*=12) born to DENV2-immune mothers infected for 8-11 months (*n*=3), and (**B**) Weight loss of pups born to DENV2-immune mothers. Serum

samples were collected from 4- to 5-week-old *LysMCre+Ifnar1*^{fl/fl} mice born to mothers previously infected with DENV2 strain S221 (5x10⁵ FFU via tail vein) or naïve mothers. Open squares represent pups (*n*=10) from naïve mothers (*n*=2) and closed triangles represent pups (*n*=12) born to DENV2-immune mothers infected for 8-11 months (*n*=3). (**C**) anti-ZIKV IgG and (**D**) anti-DENV IgG were detected via ELISA. (**E**) Neutralization capacity was assessed against DENV2 S221 via U937-DC-SIGN cells and a flow cytometry-based assay (Wen et al., 2017). Dotted line indicates limit of detection for ELISA and 0% neutralization for the neutralization assay. Data represent pooled data from two independent experiments and are expressed as mean ± standard error of mean. * *P* < 0.05, ** *P* < 0.01, *** *P* < 0.001, **** *P* < 0.0001. Unpaired Student's t test of groups for each day in (**B**).



Figure S4, **Related to Figure 3**. **IgG isolated from pups born to ZIKV-immune mothers bind both ZIKV and DENV2 but neutralize ZIKV and not DENV2**. IgG isolated and pooled from 23 mice born to ZIKV-immune mothers or 23 mice born to naïve mothers (group 1) and 13 mice born to ZIKV-immune mothers and 9 mice born to naïve mothers (group 2) for the IgG preparations used in passive transfer experiments in Figure 3 was assessed for binding and neutralization capabilities. **(A)** anti-ZIKV IgG and **(B)** anti-DENV IgG were detected via ELISA. Neutralization capacity was assessed against **(C)** ZIKV

SD001 and (**D**) DENV S221 via U937-DC-SIGN cells and a flow cytometry-based assay (Wen et al., 2017). Dotted line indicates limit of detection for ELISA and 50% or 0% neutralization for the neutralization assay. Data are expressed as mean +/- standard error of mean. (**E**) Day 3 p.i. serum samples from mice that received 145 µg of IgG and then challenged with 2 x 10⁵ FFU of DENV2 strain S221 via tail vein injection (same mice used in Figure 3) were assessed for TNF levels using a R&D systems Quantikine ELISA kit for mouse TNF. Data are pooled from 2 independent experiments with *n*=6-8 mice group, and are expressed as mean +/- standard error of mean. Mann-Whitney test, ***P* < 0.01.