

Table 1. Changes in the microbiome after viral infection

Location of microbiome or colonization	Type of evaluation	Without viral infections		virus infections or stimulation	Changes in the bacterial burden, diversity, and host immune response after viral infections	Reference
		Predominant phyla	Predominant genera			
nostril	qPCR	—	<i>S. pneumoniae</i> (54%) <i>M. catarrharis</i> (33%) <i>H. influenzae</i> (26%) <i>M. catarrlaris</i> 30.0 - 51.7%	Mainly RV	↑ <i>S. pneumoniae</i> ↑ <i>M. catarrharis</i> ↑ <i>H. influenzae</i>	DeMuri et al., 2017
nasopharynx	Bacterial culture	—	<i>S. pneumoniae</i> 27.7 - 50.3% <i>S. aureus</i> 20.2 - 25.8% <i>H. influenzae</i> 12.8 - 41.8%	RV adenovirus	Positive association between: -RV and <i>S. pneumoniae</i> , <i>H. influenzae</i> , and <i>M. catarrharis</i> -adenovirus and <i>M. catarrharis</i>	Jacoby et al., 2007
nasopharynx	Bacterial culture	—	<i>M. catarrlaris</i> 66 - 80% <i>S. pneumoniae</i> 50 - 70% <i>H. influenzae</i> 34 - 60% <i>S. aureus</i> 5 - 16%	20 respiratory viruses	Positive association between: - <i>S. aureus</i> and influenza virus - <i>S. pneumoniae</i> and RV and enterovirus - <i>H. influenzae</i> and RV and RSV - <i>M. catarrharis</i> and coronavirus and adenovirus	van den Bergh et al., 2012
nasopharynx	qPCR	—	<i>S. aureus</i> 5% <i>S. pneumoniae</i> 11% <i>H. influenzae</i> 10%	Influenza A	<i>S. pneumoniae</i> 12% <i>S. pneumoniae</i> 24% <i>H. Influenzae</i> 32%	Safaeyan et al., 2015
nasopharynx	qPCR	—	<i>S. pneumoniae</i> <i>M. catarrharis</i> <i>H. influenzae</i> <i>S. aureus</i>	LAIV	LAIV increased the nasal colonization density of <i>S. pneumoniae</i> in subjects who harbored this bacterium at the time of vaccination, and transiently increased rates of colonization by <i>H. influenza</i>	Thors et al., 2016
nasopharynx	16S rRNA sequence	—	—	RV RSV	Significant differences in the nasopharyngeal microbiome between patients with RV and RSV ↓ <i>S. aureus</i> ↑ <i>Prevotella</i> ↑ <i>Streptobacillus</i> ↑ <i>Porphyromonas</i>	Rosas-Salazar et al., 2016
nasopharynx	16S rRNA sequence	—	—	Influenza A and B	↑ <i>Granulicatella</i> ↑ <i>Veillonella</i> ↑ <i>Fusobacterium</i> ↑ <i>Haemophilus</i> -Increased bacterial diversity is associated with influenza severity	Langevin e al., 2017

Abbreviations: RV, rhinovirus; RSV, respiratory syncytial virus; LAIV, live attenuated influenza vaccine

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nasopharynx	16S rRNA sequence	Actinobacteria Firmicutes Proteobacteria	<i>Corynebacterium</i> <i>Staphylococcus</i> <i>Moraxella</i>	LAIV	<p>↑<i>Staphylococcus</i> ↑<i>Bacteroides</i> -Up-regulation of type I and II IFN-stimulated genes and activation of tpe I IFN-mediated antiviral response may promote the bacterial growth.</p>	Tarabichi et al., 2015
nasopharynx	16S rRNA sequence	—	—	Influenza A pdm09	<p>Actinobacteria Firmicutes Proteobacteria -A trend towards increasing diversity with patient age</p>	Chaban et al., 2013
*nasopharynx	Bacterial culture	—	—	LAIV	<p>↑<i>S. pneumoniae</i> ↑<i>S. aureus</i> Type I IFN was increased in the nasopharynx and bonchoalveolar lavage specimens.</p>	Mina et al., 2014
nasopharynx	16S rRNA sequence	Firmicutes Actinobacteria Proteobacteria	<i>Alloiococcus</i> <i>Corynebacterium</i> <i>Staphylococcus</i> <i>Haemophilus</i> <i>Propionibacterium</i> <i>Streptococcus</i>	RV	<p>↓<i>Haemophilus</i> ↓<i>Neisseria</i> ↑<i>Propionibacterium</i></p>	Allen et al., 2014
nasopharynx	16S rRNA sequence	Proteobacteria Firmicutes Actinobacteria	<i>Streptococcus</i> <i>Rothia</i> <i>Prevotella</i>	Influenza A and B RSV RV metapneumovirus	<p>Decreased diversity indices with symptomatic respiratory infection. ↓Anaerobes ↓<i>Prevotella</i> ↑<i>S. aureus</i> ↑<i>H. Influenzae</i> ↑<i>S. Pneumoniae</i> ↑<i>M. Catarrharis</i> ↑<i>Dol osigranulum pigrum</i> ↑<i>Corynebacterium propinquum/pseudodiphtheriticum</i></p>	Edouard et al., 2018

Abbreviations: RV, rhinovirus; RSV, respiratory syncytial virus; LAIV, live attenuated influenza vaccine; IFN, interferon; * Mouse study

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throat	16S rRNA sequence	Firmicutes Bacteroidetes Proteobacteria Actinobacteria	<i>Streptococcus</i> <i>Prevotella</i> <i>Veillonella</i> <i>Rothia</i> <i>Haemophilus</i>	RV	↑ <i>H. parainfluenzae</i> ↑ <i>Neisseria subflava</i> ↑ <i>S. aureus</i>	Hofstra et al., 2015
oropharynx	Bacterial culture Automated mass spectrometry	Bacteroidetes Proteobacteria Firmicutes	<i>Prevotella</i> <i>Acinetobacter</i>	Influenza A pdm09	↑ <i>Pseudomonas</i> ↑ <i>Acinetobacter</i>	Leung et al., 2013
oropharynx	16S rRNA sequence	Bacteroidetes Proteobacteria Firmicutes	<i>Haemophilus</i> <i>Bacteroides</i>	Influenza A (H7N9)	Significantly increased diversity compared to healthy controls, particularly H7N9 patients with secondary bacterial pneumonia.	Lu et al., 2017
Mixed URT (nasopharynx, oropharynx, and sputum)	16S rRNA sequence	—	<i>S. pneumoniae</i>	Influenza Parainfluenza RV RSV Coronavirus adenovirus metapneumovirus	Decreased diversity indices during viral infection. Following 6 types of oropharyngeal microbiome were detected; <i>Streptococcus + Prevotella + Veillonella</i> <i>Streptococcus + Haemophilus + Neisseria</i> <i>Streptococcus</i> <i>Moraxella</i> <i>Haemophilus</i> <i>Klebsiella</i>	Yi et al., 2014
LRT (induced sputum)	16S rRNA sequence	Firmicutes (65%) Bacteroidetes (17%) Proteobacteria (5%)	<i>Streptococcus</i> (42.5%) <i>Veillonella</i> (15.2%) <i>Prevotella</i> (15.0%)	RV	-are not associated with virus type but are linked to the age of subjects. Six-fold increase in bacterial burden. ↑Proteobacteria (<i>H. influenzae</i>) ↑Neutrophil concentration ↑Neutrophil elastase level	Molyneaux et al., 2013
*LRT	16S rRNA sequence	Firmicutes (90%)	<i>Lactobacillus</i>	Influenza A	Minor qualitative change in microbiota and no quantitative impact on bacterial colonization. Robust antimicrobial host response and increased sensitivity to bacterial super-infection.	Yildiz et al., 2018

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		Predominant phyla	Predominant genera			
*Gut	16S rRNA sequence	Bacteroidetes (71%)	<i>S24-7 (Bacteroidia)</i>	Influenza A	Depletion of bacterial content (↓Bacteroidetes, ↑Firmicutes) Disruption of mucus layer integrity and higher levels of antimicrobial peptides in Paneth cells.	Yildiz et al., 2018
*Gut	16S rRNA sequence	Bacteroidetes Firmicutes Tenericutes Actinobacteria Proteobacteria Deferribacteres	—	RSV Influenza	↑Bacteroidetes ↓Firmicutes	Groves et al., 2018
*Gut	16S rRNA sequence	Fimicutes Bacteroidetes	—	Influenza A (PR8)	↑Proteobacteria	Bartley et al., 2017
*Gut	16S rRNA sequence	Bacteroidetes Firmicutes	<i>S24-7 (Bacteroidia)</i>	Influenza A (PR8)	↑Proteobacteria -Influenza-induced antiviral type I IFN pathway is partly responsible for the inadequate innate immune cell responses during host defense against secondary bacterial infections.	Deriu et al., 2016
*Gut	16S rRNA sequence	—	<i>Lactobacillus</i> <i>Bacteroides</i>	Influenza A (PR8)	↓ <i>Lactobacillus</i> ↑ <i>Enterobacteriaceae</i> -Influenza infection altered intestinal microbiota composition, which was mediated by type II IFN produced by lung-derived T cells recruited to the intestine.	Wang et al., 2014

Abbreviations: RSV, respiratory syncytial virus ; IFN, interferon; * Mouse study