Supplemental Methods and Materials

Patients

A total of 799 patients with various myeloid neoplasms (MNs) (n = 575), BMFs (n = 105) and other hematologic disorders (n = 119) were screened. All samples were obtained after written informed consent, according to protocols approved by the Institutional Review Board of participating institutions. The patients with myeloid neoplasm consisted of MDS (n = 422), acute myeloid leukemia (AML) (n = 98), and MDS/myeloproliferative neoplasms (MPN) (n = 55). Most (78/98, 80%) AML patients had secondary (post-MDS) disease. The BMF cohorts were aplastic anemia (AA) (n = 47), paroxysmal nocturnal hemoglobinuria (PNH) (n = 40), and pure red cell aplasia (PRCA) (n = 18). The other hematologic disorders were large granular lymphocytic leukemia (n = 18)= 55) and idiopathic cytopenia of undetermined significance (n = 20). We sequenced all patients for the presence of SAMD9/SAMD9L variants. Paired disease and normal GL DNA were obtained from 220 MN patients for whole exome sequencing (WES). GL DNA was obtained from either buccal mucosa or CD3 positive T cells which were purified from peripheral blood with or without prior culture in the presence of PHA and IL-2¹. Disease DNA was extracted from bone marrow or peripheral blood. 579 patients were evaluated by targeted capture sequencing. We used Broad Institute's Genome Aggregation Database (gnomAD) as healthy controls (n = 138,632) which included 123.136 WES and 15.496 whole-genome sequences from unrelated individuals². After comparing ethnicities between gnomAD and our cohort^{2,3}, we matched healthy controls to patients by ethnicities (Supplemental Figure 3).

Whole exome sequencing

WES was performed as previously described^{1,4}. Paired disease and normal GL DNAs were used. Whole-exome capture was accomplished through hybridization of sonicated genomic DNA to a bait cDNA library synthesized on magnetic beads (SureSelect Human All Exon 50Mb or V4 kit, Agilent Technologies). Captured targets were subjected to massively parallel sequencing using a HiSeq 2000 (Illumina) according to the standard protocol for 100-bp paired-end reads. Subsequent validation and confirmatory sequencing are described below. Briefly, sequencing reads were aligned to the human genome (hg19) by a Burrows-Wheeler aligner (http://bio-bwa.sourceforge.net/). We used a GATK pipeline to extract candidate variants/polymorphisms and to remove sequencing errors. Validations were performed by Sanger or PCR amplicon sequencing⁴.

Targeted Capture Sequencing

Targeted capture sequencing was performed using a TruSeq Custom Amplicon or Nextera Rapid capture custom enrichment kit (Illumina) as previously described^{4,5}. The targeted 46 genes were captured (Supplemental Table 2 and 3). Sequencing libraries were generated according to an Illumina paired-end library protocol. The enriched targets were subjected to massive sequencing using HiSeq 2000 or MiSeq sequencer (Illumina), with sufficient read coverage (Supplemental Figure 2). Variants were annotated using ANNOVAR⁶ and filtered by removing: i) synonymous single nucleotide variants; ii) variants only present in unidirectional reads; iii) variants in repetitive genomic regions. Only variants with minimum depth of 20 and with 5 positive, high quality reads, were considered. Variant allelic frequencies were adjusted according to zygosity and copy number based on single nucleotide polymorphism array results⁵. Finally, mapping errors were removed by visual inspection with the Integral Genomics Viewer. Validation by Sanger sequencing or PCR amplicon sequencing was performed as previously described⁵.

Criteria of rare variants and pathogenic missense variants

We defined "rare variants" of *SAMD9/SAMD9L* genes as those present in <0.1% of ethnicallymatched healthy controls (Supplemental Figure 3). To predict functional effects of specific *SAMD9* missense variants, we focused on *SAMD9* germline variants reported in previous papers^{7,8} or public databases of healthy donors. 39 variants were identified, 23 out of 39 were deemed "truly pathogenic" variants by confirming that there was a significant phenotype using a functional assay and by examining familial disease phenotype co-segregations (Supplemental Table 5). ROC curves of 8 different algorithms⁹⁻¹³ were created (Supplemental figure 4), and Areas Under the Curves (AUC) were compared between them (Supplemental Table 6). We set a strict cut-off to discover missense variants with high probabilities of pathogenesis, even if it reduces the sensitivity. The top 5 algorithms in terms of greatest AUC were chosen and variants with \geq 3 positive scores out of the top 5 algorithms were defined as "Pathogenic". Applying this criteria to 39 variants already confirmed to be pathogenic or not yielded 9 as positive and 7 as truly pathogenic (Supplemental figure 5). Given that 23 were truly pathogenic and 16 were not, sensitivity and specificity were 30% (7/(7+16)) and 88% (14/(14+2)), respectively.

Metaphase cytogenetics

Chromosome preparations were G-banded using trypsin and Giemsa, and karyotypes were described in 799 patients according to the International System for Human Cytogenetic Nomenclature¹⁴.

Single-nucleotide polymorphism array analysis

Single-nucleotide polymorphism (SNP) array karyotyping for confirming metaphase cytogenetics and detecting copy-number neutral loss of heterozygosity was performed as previously described^{15,16}. Briefly, Affymetrix 250K and 6.0 SNP arrays were used to evaluate copy number and loss of heterozygosity. Using our internal and a publicly available database (http://dgv.tcag.ca/dgv/app/home), the screening algorithm validated each lesion as somatic. Non-somatic lesions were excluded from further analysis. Affected genomic positions in each lesion were visualized and extracted using CNAG (v3.0) or Genotyping Console (Affymetrix)^{17,18}.

Mutagenesis and constructs

A plasmid encoding N-terminal FLAG-tagged human SAMD9 with the tetracycline-inducible system was used as described previously¹⁹. Plasmids encoding five SAMD9 variants (Thr205Pro, Ile247Thr, Ile268Thr, Asp550Val, and Leu574Pro) were generated with a standard site-directed mutagenesis technique with use of wild-type (WT) SAMD9-expressing vector as a template. We also created a vector encoding N-terminal TagRFP-tagged human SAMD9L by inserting the TagRFP sequence (derived from pTagRFP-C [Evrogen, Moscow, Russia]) and human SAMD9L cDNA sequence, and deleting the SAMD9 sequence. Plasmids encoding 9 SAMD9L variants (Leu50Ser, Glu220Gly, Cys228Tyr, Gly235Ser, Gly247Ala, Trp333Cys, Trp517Arg, His880Glu, and Leu1323fs) were generated with a site-directed mutagenesis technique.

Inducible stably transfected HEK293 cells

HEK293 cells were purchased from the American Type Culture Collection (ATCC CRL-1573). Mycoplasma infection was excluded through the standard indirect DNA staining method. HEK293 cell lines stably expressing each FLAG-SAMD9 and TagRFP-SAMD9L proteins (wild type, mutant, or empty vector) in the presence of doxycycline (1 μ g/mL) were established according to the manufacturer's protocol^{7,19}. Cells were maintained in DMEM supplemented with 10% FBS in a humidified incubator at 37°C and 5% CO2.

Intracellular localization analysis

Inducible stable HEK293 cells were grown on glass-bottom dishes (Greiner bio-one). Forty-eight hours after induction, cells were fixed and permeabilized by incubation in cold methanol at -20°C for 10 min. Nuclei were stained with Hoechst 33342 (Dojindo). Images were obtained by confocal laser scanning microscopy FV1000D (Olympus).

Cell proliferation assays

For cell growth assays, inducible stable HEK293 cells, were seeded into 96-well plate with about 5% confluence in 200 μ L of culture medium with or without 1 μ g/mL doxycycline. The degree of confluence was quantified every 3 hours using an IncuCyte ZOOM time-lapse microscope (Essen BioScience). Growth curve data are representative of three independent assays that were performed in triplicate.

Statistical analyses

Proportions were compared using two-sided Fisher exact tests using R.

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Supplemental Figure 1. Characteristics of familial thrombocytopenia with SAMD9L variant. (A) A family tree identifying those with a SAMD9L variant (Trp517Arg) and their thrombocytopenic history (filled: thrombocytopenia; open: confirmed no thrombocytopenia). Ages are on the upper right of pedigree symbols. Paternity was confirmed by whole exome sequencing of patients' and parental germline DNAs. WT denotes wild type. (B) Graphical representation of SAMD9L protein. Functional sterile alpha motif (SAM) is depicted in green. Distribution of germline SAMD9L variants reported in pediatric MDS patients are shown in the lower panel. Amino acid alignment of SAMD9L proteins from different species shows evolutionarily conserved amino acids in red. Position of SAMD9L variants are indicated by a red arrow. (C) Clinical course of the 9-months-old patient with thrombocytopenia, including peripheral-blood counts, platelet transfusion times, and cytogenetics of marrow aspirates. After severe pancytopenia requiring transfusions for four months, this patient's cytopenias resolved spontaneously and he became transfusion-independent.



Supplemental Figure 2. Read coverage for SAMD9 and SAMD9L in targeted capture sequencing. Shown are the percentages of patients with read coverages of more than $50 \times$, $100 \times$, $200 \times$ and $500 \times$ for SAMD9 and SAMD9L.



Supplemental Figure 3. Healthy controls were selected to have ethnicities that matched those of cases in our cohort.

Asian

African

25%

We used Broad Institute's Genome Aggregation Database (gnomAD) to obtain ethnicity matched healthy controls (Center table) from a total of 138,176 subjects (Left table); to compare frequencies of germline *SAMD9/SAMD9L* variants in our sporadic adult MDS cohort (Right table) to these controls, ethnicities had to be matched. Thus, Latinos and Finns that were not included in our cohort were omitted, but Caucasians, Asians and African-Americans were included. Fractions of these races are depicted using pie charts.



Supplemental Figure 4. Receiver Operating Characteristic curve for prediction algorithms for missense variants.

ROC curves of 8 different algorithms were created by using 39 variants which include 23 that were deemed "truly pathogenic" variants by confirming that there was a significant phenotype using a functional assay and by examining familial disease phenotype co-segregations. Blue line graphs depicted the top 5 algorithms in terms of greatest Area Under the Curve.



Supplemental Figure 5. Contingency table for prediction of SAMD9 missense variants

We set a strict cut-off (variants with \geq 3 positive scores out of the top 5 algorithms in terms of greatest AUC) to discover missense variants with high possibilities of pathogenesis, even if it reduces the sensitivity. Applying this criteria to 39 variants already confirmed to be pathogenic or not yielded 9 as positive and 7 as truly pathogenic. Given that 23 were truly pathogenic and 16 were not, sensitivity and specificity were 30% (7/(7+16)) and 88% (14/(14+2)), respectively. Positive predict values and negative predict values were also 78% [7/(7+2)] and 47% [14/(16+14)].



Supplemental Figure 6. Three-dimensional structure of SAM domain in SAMD9 and SAMD9L

Three-dimensional structure of SAM domain were evaluated by the University of California–San Francisco Chimera Program and PDB 2B6G of a protein data bank administered by the RCSB (Research Collaboratory for Structural Bioinformatics). Wild type (Left) and His32Leu and Leu50Ser variants (Right) were shown respectively.



Time (Hours)

Supplemental Figure 7. Cell proliferation curves for wild type (WT) or mutants of SAMD9/SAMD9L before and after doxycycline induction

Chr	7
Start	92763736
End	92763736
Ref	А
Alt	G
Function	exonic
Genes	SAMD9L
ExonicFunc	nonsynonymous SNV
Alterations	SAMD9L:NM_001303500:c.T1549C:p.W517R
gnomAD frequency ALL	0.0003
gnomAD frequency AFR	0
gnomAD frequency AMR	2.98E-05
gnomAD frequency ASJ	0
gnomAD frequency EAS	0
gnomAD frequency FIN	4.49E-05
gnomAD frequency NFE	0.0006
gnomAD frequency OTH	0
gnomAD frequency SAS	0
avsnp147	rs199714577
SIFT score	0
SIFT converted rankscore	0.912
SIFT pred	D
Polyphen2 HDIV score	1
Polyphen2 HDIV rankscore	0.899
Polyphen2 HDIV pred	D
Polyphen2 HVAR score	0.992
Polyphen2 HVAR rankscore	0.79
Polyphen2 HVAR pred	D
LRT score	0
LRT converted rankscore	0.629
LRT pred	D
MutationTaster score	1
MutationTaster converted rankscore	0.487
MutationTaster pred	D
MutationAssessor score	2.8
MutationAssessor score rankscore	0.819
MutationAssessor pred	М
FATHMM score	2.15
FATHMM converted rankscore	0.193
FATHMM pred	T
PROVEAN score	-7.18
PROVEAN converted rankscore	0.942
PROVEAN pred	D

Supplementary Table 1. A novel SAMD9L variant identified in a family with thrombocytopenia

Abbreviations; AFR, African; AMR, Latno; EUR, European (Non-Finnish); ASJ, AsjkenaziJewish; EAS, East Asian; FIN, Finnish; NFE, Non-Finnish European; OTH, Other; SAS, South Asia; FATHMM, Functional Analysis through Hidden Markov Models; PROVEAN, Protein Variation Effect Analyzer

APC	ETV6	NPM1	SRSF2
ASXL1	EZH2	NRAS	STAG2
BCOR	FLT3	PHF6	STAT3
BCORL1	GATA2	PRPF8	SUZ12
CALR	IDH1	PTPN11	TET2
CBL	IDH2	RAD21	TP53
CEBPA	JAK2	RUNX1	U2AF1
CSF1R	JAK3	SAMD9	U2AF2
CUX1	KDM6A	SAMD9L	WT1
DDX41	KIT	SETBP1	ZRSR2
DNMT3A	KRAS	SF3B1	
EED	NF1	SMC3	

Supplementary Table 2. Panel of 46 genes for targeted capture sequencing

Chromosomo Probes		0	# of unables	
Chromosome	Start	End	Gene name	# of probes
chr5	112043415	112043579	APC	1
chr5	112090588	112090722	APC	1
chr5	112102023	112102107	APC	1
chr5	112102886	112103087	APC	1
chr5	112111326	112111434	APC	1
chr5	112116487	112116600	APC	1
chr5	112128143	112128226	APC	1
chr5	112126140	112120220	APC	1
chr5	112150070	112151290	APC	1
chr5	112154663	112155041	APC	2
chr5	112157593	112157688	APC	1
chr5	112162805	1121620//		1
chr5	112163626	112102344		1
chr5	11216/553	11216/660		1
chr5	112104000	112104003		1
chr5	112170040	112170002	AFC	20
chi S	20046570	20046625	AFC ACVL1	29
chi20	2005/19	20054260	ASALI	1
chii20	30934107	30934209	ASALI	1
CIII 20	30930013	30930920	ASXLI	1
CIII 20	30939907	30939909	ASXLI	1
chr20	31015931	31010051	ASXLI	1
cnr20	31016128	31016225	ASXL1	1
chr20	31017141	31017234	ASXL1	1
chr20	31017704	31017856	ASXL1	1
chr20	31019124	31019287	ASXL1	1
chr20	31019386	31019482	ASXL1	1
chr20	31020683	31020788	ASXL1	1
chr20	31021087	31021720	ASXL1	3
chr20	31022235	31025138	ASXL1	13
chrX	39911365	39911653	BCOR	2
chrX	39913139	39913295	BCOR	1
chrX	39913509	39913586	BCOR	1
chrX	39914621	39914766	BCOR	1
chrX	39916408	39916574	BCOR	1
chrX	39921392	39921646	BCOR	2
chrX	39921999	39922324	BCOR	2
chrX	39922861	39923205	BCOR	2
chrX	39922861	39923103	BCOR	2
chrX	39923589	39923852	BCOR	2
chrX	39930226	39930412	BCOR	1
chrX	39930890	39930943	BCOR	1
chrX	39931602	39934433	BCOR	13
chrX	39935707	39935785	BCOR	1
chrX	39937097	39937182	BCOR	1
chrX	129139208	129139293	BCORL1	1
chrX	129146554	129146644	BCORL1	1
chrX	129146926	129150189	BCORL1	15
chrX	129154960	129155125	BCORL1	1

Supplementary Table 3. Targeted regions for a panel of 46 genes

chrX	129156872	129156952	BCORL1	1
chrX	129158965	129159354	BCORL1	2
chrX	129162610	129162836	BCORL1	1
chrX	129171342	129171508	BCORL1	1
chrX	129173112	129173257	BCORL1	1
chrX	129184692	129184769	BCORL1	1
chrX	129185835	129185991	BCORL1	1
chrX	129189829	129190108	BCORL1	2
chr19	13049494	13049584	CALR	1
chr19	13049948	13050049	CALR	1
chr19	13050242	13050445	CALR	1
chr19	13050867	13050961	CALR	1
chr19	13051057	13051266	CALR	1
chr19	13051355	13051468	CALR	1
chr19	13051558	13051701	CALR	1
chr19	13054351	13054443	CALR	1
chr19	13054527	13054724	CALR	1
chr11	119077128	119077322	CBL	1
chr11	119103158	119103405	CBL	2
chr11	119142445	119142591	CBL	1
chr11	119144578	119144734	CBL	1
chr11	119145542	119145663	CBL	1
chr11	119146707	119146844	CBL	1
chr11	119148467	119148554	CBL	1
chr11	119148876	119149007	CBL	1
chr11	119149220	119149423	CBL	1
chr11	119155679	119155810	CBL	1
chr11	119155899	119156276	CBL	2
chr11	119158562	119158656	CBL	1
chr11	119167628	119167744	CBL	1
chr11	119168094	119168191	CBL	1
chr11	119169068	119169250	CBL	1
chr11	119170205	119170488	CBL	2
chr19	33792247	33793320	CEBPA	5
chr5	149433635	149433787	CSF1R	1
chr5	149433885	149433993	CSF1R	1
chr5	149434800	149434899	CSF1R	1
chr5	149435589	149435700	CSF1R	1
chr5	149435782	149435904	CSF1R	1
chr5	149436850	149436947	CSF1R	1
chr5	149437067	149437155	CSF1R	1
chr5	149439263	149439425	CSF1R	1
chr5	149440425	149440535	CSF1R	1
chr5	149441054	149441158	CSF1R	1
chr5	149441286	149441412	CSF1R	1
chr5	149447778	149447893	CSF1R	1
chr5	149449436	149449626	CSF1R	1
chr5	149449745	149449865	CSF1R	1
chr5	149450019	149450134	CSF1R	1
ohrE				
CHID	149452864	149453056	CSF1R	1
chr5 chr5	149452864 149456839	149453056 149456998	CSF1R CSF1R	1

chr5	149459615	149459899	CSF1R	2
chr5	149460330	149460587	CSF1R	2
chr5	149465942	149465990	CSF1R	1
chr7	101459311	101459373	CUX1	1
chr7	101460920	101460949	CUX1	1
chr7	101559395	101559505	CUX1	1
chr7	101671378	101671425	CUX1	1
chr7	101713619	101713697	CUX1	1
chr7	101740644	101740781	CUX1	1
chr7	101747616	101747739	CUX1	1
chr7	101754978	101755054	CUX1	1
chr7	101758487	101758553	CUX1	1
chr7	101801840	101801888	CUX1	1
chr7	101813726	101813830	CUX1	1
chr7	101821749	101821937	CUX1	1
chr7	101821755	101821937	CUX1	1
chr7	101833093	101833151	CUX1	1
chr7	101837122	101837170	CUX1	1
chr7	101838787	101838883	CUX1	1
chr7	101839914	101840585	CUX1	3
chr7	101842082	101842147	CUX1	1
chr7	101843351	101843452	CUX1	1
chr7	101844640	101845484	CUX1	4
chr7	101847671	101847836	CUX1	1
chr7	101848394	101848450	CUX1	1
chr7	101870647	101870949	CUX1	2
chr7	101877332	101877520	CUX1	1
chr7	101882600	101882864	CUX1	2
chr7	101891692	101892319	CUX1	3
chr7	101916637	101916764	CUX1	1
chr7	101917515	101917581	CUX1	1
chr7	101918518	101918630	CUX1	1
chr7	101921220	101921336	CUX1	1
chr7	101923329	101923412	CUX1	1
chr7	101924096	101924152	CUX1	1
chr7	101925132	101925212	CUX1	1
chr7	101926004	101926068	CUX1	1
chr7	101926313	101926379	CUX1	1
chr5	176938795	176938928	DDX41	1
chr5	176939097	176939207	DDX41	1
chr5	176939323	176939394	DDX41	1
chr5	176939497	176939646	DDX41	1
chr5	176939781	176939877	DDX41	1
chr5	176940012	176940083	DDX41	1
chr5	176940354	176940485	DDX41	1
chr5	176940686	176940848	DDX41	1
chr5	176941702	176941838	DDX41	1
chr5	176941917	176942070	DDX41	1
chr5	176942187	176942259	DDX41	1
chr5	176942686	176942822	DDX41	1
chr5	176942930	176942990	DDX41	1
chr5	176943120	176943194	DDX41	1

ohrE	1760/2200	1760/2//0		1
CIII D	1/0943209	170943440		1
chr5	1/0943/20	170943830		1
CUL2	1/0943920	1/0943940		1
CNFZ	20407101	25457289	DNMT 3A	1
cnr2	25458576	25458694	DNMT 3A	1
chr2	25459805	25459874	DNMT3A	1
chr2	25461999	25462084	DNMT3A	1
chr2	25463171	25463319	DNMT3A	1
chr2	25463509	25463599	DNMT3A	1
chr2	25464431	25464576	DNMT3A	1
chr2	25466767	25466851	DNMT3A	1
chr2	25467024	25467207	DNMT3A	1
chr2	25467409	25467521	DNMT3A	1
chr2	25468122	25468201	DNMT3A	1
chr2	25468889	25468933	DNMT3A	1
chr2	25469029	25469178	DNMT3A	1
chr2	25469489	25469645	DNMT3A	1
chr2	25469920	25470027	DNMT3A	1
chr2	25470460	25470618	DNMT3A	1
chr2	25470906	25471121	DNMT3A	1
chr2	25472526	25472593	DNMT3A	1
chr2	25475063	25475066	DNMT3A	1
chr2	25497810	25497956	DNMT3A	1
chr2	25498369	25498412	DNMT3A	1
chr2	25505260	25505580	DNMT3A	2
chr2	25505310	25505580	DNMT3A	2
chr2	25523008	25523112	DNMT3A	1
chr2	25536782	25536853	DNMT3A	1
chr11	85956272	85956385	FFD	1
chr11	85961338	85961490	FED	1
chr11	85963190	85963282	FED	1
chr11	85966264	85966329	EED	1
ohr11	85067420	85067551		1
ohr11	85068557	85068638		1
obr11	95075214	85075205		1
ohr11	05975214	05975505		1
CIII I I obr11	009//120	009//200		1
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chrX	123184036	123184159	STAG2	1
chrX	123184971	123185069	STAG2	1
chrX	123185165	123185244	STAG2	1
chrX	123189978	123190085	STAG2	1
chrX	123191716	123191827	STAG2	1
chrX	123195074	123195191	STAG2	1
chrX	123195621	123195724	STAG2	1

chrX chrX	123196752	123196844	STAG2	1
chrX				
	123196966	123197055	STAG2	1
chrX	123197698	123197901	STAG2	1
chrX	123199726	123199796	STAG2	1
chrX	123200025	123200112	STAG2	1
chrX	123200206	123200286	STAG2	1
chrX	123202414	123202506	STAG2	1
chrX	123204999	123205173	STAG2	1
chrX	123210182	123210321	STAG2	1
chrX	123211807	123211908	STAG2	1
chrX	123215230	123215378	STAG2	1
chrX	123217271	123217399	STAG2	1
chrX	123220397	123220620	STAG2	1
chrX	123224425	123224614	STAG2	1
chrX	123224704	123224814	STAG2	1
chrX	123227868	123227994	STAG2	1
chrX	123229222	123229299	STAG2	1
chrX	123234424	123234444	STAG2	1
chr17	40467766	40467818	STAT3	1
chr17	40468807	40468919	STAT3	1
chr17	40468848	40468869	STAT3	1
chr17	40469200	40469242	STAT3	1
chr17	40474300	40474512	STAT3	1
chr17	40474303	40474512	STAT3	1
chr17	40475022	40475161	STAT3	1
chr17	40475278	40475372	STAT3	1
chr17	40475591	40475643	STAT3	1
chr17	40476729	40476864	STAT3	1
chr17	40476981	40477079	STAT3	1
chr17	40478134	40478217	STAT3	1
chr17	40481428	40481475	STAT3	1
chr17	40481572	40481665	STAT3	1
chr17	40481765	40481794	STAT3	1
chr17	40483490	40483549	STAT3	1
chr17	40485691	40485783	STAT3	1
chr17	40485909	40486067	STAT3	1
chr17	40489453	40489604	STAT3	1
chr17	40489781	40489875	STAT3	1
chr17	40490749	40490830	STAT3	1
chr17	40491332	40491427	STAT3	1
chr17	40497577	40497675	STAT3	1
chr17	40498587	40498731	STAT3	1
chr17	40500407	40500534	STAT3	1
chr17	30264266	30264539	SUZ12	2
chr17	30267305	30267351	SUZ12	1
chr17	30267441	30267505	SUZ12	1
chr17	30274636	30274704	SUZ12	1
chr17	30293166	30293215	SUZ12	1
chr17	30300165	30300250	SUZ12	1
chr17	30302501	30302732	SUZ12	2
chr17	30303540	30303633	SUZ12	1
chr17	30310018	30310123	SU712	1

chr17	30315339	30315516	SUZ12	1
chr17	30320261	30320352	SUZ12	1
chr17	30320884	30321027	SUZ12	1
chr17	30321583	30321740	SUZ12	1
chr17	30322583	30322781	SUZ12	1
chr17	30323817	30323896	SUZ12	1
chr17	30325677	30326019	SUZ12	2
chr4	106155100	106158594	TET2	16
chr4	106155100	106158508	TET2	15
chr4	106162496	106162586	TET2	1
chr4	106163991	106164084	TET2	1
chr4	106164727	106164935	TET2	1
chr4	106180776	106180926	TET2	1
chr4	106182916	106183005	TET2	1
chr4	106190767	106190904	TET2	1
chr4	106193721	106194075	TET2	2
chr4	106196205	106197673	TET2	7
chr17	7572930	7573008	TP53	1
chr17	7573927	7574033	TP53	1
chr17	7576540	7576584	TP53	1
chr17	7576628	7576657	TP53	1
chr17	7576853	7576926	TP53	1
chr17	7577019	7577155	TP53	1
chr17	7577499	7577608	TP53	1
chr17	7578177	7578289	TP53	1
chr17	7578371	7578554	TP53	1
chr17	7578371	7578533	TP53	1
chr17	7579312	7579569	TP53	2
chr17	7579312	7579590	TP53	2
chr17	7579700	7579721	TP53	1
chr17	7579839	7579912	TP53	1
chr21	44513215	44513359	U2AF1	1
chr21	44514581	44514673	U2AF1	1
chr21	44514765	44514898	U2AF1	1
chr21	44515548	44515646	U2AF1	1
chr21	44515804	44515833	U2AF1	1
chr21	44515804	44515853	U2AF1	1
chr21	44520563	44520629	U2AF1	1
chr21	44521476	44521542	U2AF1	1
chr21	44524425	44524512	U2AF1	1
chr21	44527561	44527604	U2AF1	1
chr19	56166471	56166519	U2AF2	1
chr19	56170576	56170711	112AF2	1
chr19	56171543	56171587	112AF2	1
chr19	56171882	56171985	112AF2	1
chr19	56172404	56172555	112AF2	1
chr10	56173868	5617308/	112AF2	1
chr10	56174072	56175110	1120F2	1
chr10	56170272	56170052	112052	1
chr10	56120026	56120152	112052	1
chr10	56120///0	56180535	112052	1
chr10	56120/1/0	561805/7	112052	1
	00100440	00100047		1

chr19	56180810	56181058	U2AF2	2
chr19	56185300	56185431	U2AF2	1
chr11	32410607	32410725	WT1	1
chr11	32413518	32413610	WT1	1
chr11	32413527	32413610	WT1	1
chr11	32414212	32414301	WT1	1
chr11	32417803	32417953	WT1	1
chr11	32421494	32421590	WT1	1
chr11	32438036	32438086	WT1	1
chr11	32439123	32439200	WT1	1
chr11	32449502	32449604	WT1	1
chr11	32450043	32450165	WT1	1
chr11	32452076	32452085	WT1	1
chr11	32456246	32456891	WT1	3
chrX	15808619	15808659	ZRSR2	1
chrX	15809057	15809136	ZRSR2	1
chrX	15817995	15818076	ZRSR2	1
chrX	15821811	15821919	ZRSR2	1
chrX	15822234	15822320	ZRSR2	1
chrX	15826356	15826394	ZRSR2	1
chrX	15827323	15827441	ZRSR2	1
chrX	15833800	15834013	ZRSR2	1
chrX	15836710	15836765	ZRSR2	1
chrX	15838330	15838439	ZRSR2	1
chrX	15840854	15841362	ZRSR2	3

Supplementary Table 4. Rare SAMD9/SAMD9L germline variants in our cohort (n = 26)

UPN	Age [#]	Sex	Diagnosis [†]	Gene	Alterations	VAFs of germline [¶]	VAFs of tumor	Chr7 copy number alteration [‡]	Frequency in healthy controls*	Family history	Bone marrow cellularity	Infectious history	lgG (mg/dL) [§]
UPN004	78	F	MDS	SAMD9L	1327V	49%	53%	-7	.02%	-	60%	-	NA
UPN005	77	М	sAML	SAMD9L	L1323fs	50%	90%	-7	<.01%	-	50%	-	NA
UPN006	73	М	MDS	SAMD9	Q154X, I268T	46%, 47%	40%, 41%	No	.01%, <.01%	NA	50%	NA	NA
UPN007	88	F	MDS/MPN	SAMD9	H32L	38%	45%	No	.04%	NA	90%	NA	NA
UPN012	83	F	MDS	SAMD9	T205P	39%	49%	No	.08%	-	80%	NA	NA
UPN013	78	F	MDS	SAMD9	T205P	47%	46%	No	.08%	-	70%	NA	NA
UPN014	49	F	MDS	SAMD9	R221Q	54%	48%	No	.02%	-	70%	Pneumonia due to Pseudomonas aeruginosa	308
UPN015	66	М	MDS/MPN	SAMD9	I268T	44%	47%	No	<.01%	NA	60%	NA	NA
UPN017	75	М	MDS/MPN	SAMD9	D550V	44%	51%	No	0%	-	20%	-	NA
UPN020	79	М	MDS	SAMD9	N1244D	42%	48%	No	<.01%	NA	30%	NA	NA
UPN022	73	М	MDS	SAMD9L	L50S	48%	49%	No	0%	NA	40%	NA	NA
UPN023	86	F	MDS	SAMD9L	P123L	46%	52%	No	.02%	Breast cancer (M)	30%	-	NA
UPN024	61	F	MDS	SAMD9L	E220G	47%	46%	No	.01%	-	70%	NA	NA
UPN025	42	М	MDS	SAMD9L	G235S	45%	47%	No	.01%	-	40%	NA	NA
UPN026	70	F	AA/PNH	SAMD9L	W333C	-	45%	No	0%	-	<5%	-	NA
UPN028	34	F	AA/PNH	SAMD9L	A637T	-	53%	No	.02%	-	30%	-	985
UPN029	59	М	MDS	SAMD9L	S728P, Y705C	48%, 51%	55%, 47%	No	.02%, .01%	NA	50%	NA	NA
UPN_ADD1	61	F	MDS	SAMD9	A949S	-	36%	No	.08%	-	60%	-	1490
UPN_ADD2	75	F	MDS	SAMD9	R106H	-	53%	No	.02%	-	90%	-	NA
UPN_ADD3	41	F	MDS	SAMD9	R902Q	-	49%	No	<.01%	-	10%	Positive blood culture including <i>VRE, Klebsiella</i> , and <i>C. albicans</i>	352
UPN_ADD4	78	F	MDS	SAMD9L	R1298G	-	54%	No	<.01%	-	60%	-	544
UPN_ADD5	52	М	AA/PNH	SAMD9L	A637S	-	45%	No	<.01%	-	20%	-	NA
UPN_ADD6	57	М	MDS	SAMD9L	G247A	-	48%	No	0%	-	50%	Pneumonia due to Pneumocystis jirovecii	1,120
UPN_ADD7	70	М	sAML	SAMD9	T1370fs	-	51%	No	.02%	-	40%	-	590
#**		vo 4 00.			0		· · · · · · · · · · · · · · · · · · ·	1.11					

[#]Median age and range; 72 (34-88) in patients with SAMD9/SAMD9L variants (n = 24) vs. 68 (17-94) in patients without them (n = 775), P = 0.3 (T test)

[†]Previous treatment history; 0% (0/24) in patients with SAMD9/SAMD9L variants vs. 0.6% (5/775) in patients without them, P = 1 (Fisher's exact test) [¶] "-" depicts that germline DNAs were not available.

[‡]Aberrant karyotype; 46% (11/24) in patients with SAMD9/SAMD9L variants vs. 28% (220/775) in patients without them, P = 0.07 (Fisher's exact test);

Cytogenetic subsets in detal were shown (Supplemental table 7)

*Ethnically-matched healthy controls from gnomAD (n = 99,860) (Supplemental Figure 1)

[§]Immunoglobulin G; normal range; 717-1,411 (mg/dl)

Abbreviations; UPN, universal patient number; sAML, secondary acute myeloid leukemia; MDS, myelodysplastic syndrome; -7, monosomy 7;

VAFs, variant allele frequencies; NA, not applicable; M, mother; VRE, vancomycin resistant Enterococci; C. albicans, Candida albicans.

Supplementary Table 5. Truly pathogenic SAMD9 germ line variants (n=23)

Amino Acid change	Prediction (3>positive)
SAMD9:A1195V	Pathogenic
SAMD9:1983S	Pathogenic
SAMD9:R982H	Pathogenic
SAMD9:R982C	Pathogenic
SAMD9:1897M	Pathogenic
SAMD9:Y684C	Pathogenic
SAMD9:L641P	Pathogenic
SAMD9:R1293Q	VUS
SAMD9:R1293W	VUS
SAMD9:Q1286K	VUS
SAMD9:P1280L	VUS
SAMD9:E1136Q	VUS
SAMD9:F1017V	VUS
SAMD9:E974K	VUS
SAMD9:R824Q	VUS
SAMD9:K821M	VUS
SAMD9:D769N	VUS
SAMD9:D769G	VUS
SAMD9:A722E	VUS
SAMD9:R685Q	VUS
SAMD9:K1569N	VUS
SAMD9:L1539I	VUS
SAMD9:R459Q	VUS

Abbreviations; P, pathogenic; VUS, variants of unknown significance

Supplementary Table 6. Area Under the Curve (AUC) for 8 prediction algorithms

Algorithms	AUC
PolyPhen-2	0.823
Mutation Assessor	0.810
MutationTaster	0.780
SIFT	0.777
PROVEAN	0.764
M-CAP	0.726
DANN	0.712
CADD	0.688

Predicted UPN Gene Alterations pathogenesis UPN006 SAMD9 1268T Ρ Ρ UPN012 SAMD9 T205P Ρ UPN013 SAMD9 T205P Ρ SAMD9 UPN015 1268T Ρ UPN017 SAMD9 D550V Ρ UPN022 SAMD9L L50S Ρ UPN024 SAMD9L E220G Ρ UPN025 SAMD9L G235S Ρ UPN026 SAMD9L W333C Ρ UPN_ADD6 SAMD9L G247A VUS, P S728P, Y705C UPN029 SAMD9L VUS UPN007 SAMD9 H32L VUS SAMD9 R221Q UPN014 VUS UPN020 SAMD9 N1244D VUS UPN_ADD1 SAMD9 A949S VUS UPN_ADD2 SAMD9 R106H VUS UPN_ADD3 SAMD9 R902Q VUS UPN004 SAMD9L 1327V UPN023 SAMD9L P123L VUS VUS **UPN028** SAMD9L A637T VUS UPN_ADD4 SAMD9L R1298G VUS UPN_ADD5 SAMD9L A637S

Supplementary Table 7. SAMD9 and SAMD9L missense rare variants with predicted pathogenesis based on top 5 algorithms

Abbreviations; P, pathogenic; VUS, variants of unknown significance

Supplementary Table 8. Rare SAMD9/SAMD9L GL variants in myeloid neoplasms database (n = 7)

Sample	Age	Sex	Genes	Alterations	Location	Diagnosis	Subtypes	Frequency in healthy controls	Predicted pathogenesis
UPN39	88	F	SAMD9	H32L	N-terminus	MDS/MPN	CMML	0.04%	VUS
AML139	58	М	SAMD9	I247T	N-terminus	AML	Normal Karyotype, NPM1 mutations	<.01%	Р
UPN41	66	М	SAMD9	I268T	N-terminus	MDS/MPN	CMML	<.01%	Р
PD9412	NA	NA	SAMD9	L574P	Central	MPN	PMF	<.01%	Р
AML082	37	М	SAMD9	Q1427P	C-terminus	AML	46, XY, del (11) (q14 q25), CEBPA (double mutant)	0%	VUS
PD4780	NA	NA	SAMD9L	C228Y	N-terminus	MPN	PMF	<.01%	Р
PD4996	NA	NA	SAMD9L	R285Q	N-terminus	MPN	PMF	<.01%	VUS

Abbreviations; NA, not applicable; CMML, Chronic myelomonocytic leukemia; PMF, primary myelofibrosis; P, pathogenic; VUS, variants of ur

Gene	Locations	Amino acids
SAMD9	N-terminal	1-530
SAMD9	Center	531-1,060
SAMD9	C-terminal	1,061-1,589
SAMD9L	N-terminal	1-528
SAMD9L	Center	529-1,056
SAMD9L	C-terminal	1,057-1,584

Supplementary Table 9. Three gene locations of SAMD9 and SAMD9L

UPN	Cytogenetics	Gene	Alterations	VAFs of	VAFs of
UPN004	44,XX,-4,del(5)(q12q33),-17,add(19)(p13)[3]/45,sl,- add(19)(p13),+19,+mar1[7]/44,sl1,-6,+inv(6)(p23q25),-7 ,-10,+del(10)(q25),-14,+add(14)(q32),-15,- 22,+der(22)t(15;22)(q11.2;q13),+mar2[cp6]/46,XX[4]	SAMD9L	1327V	49%	53%
UPN005	42,XY,add(1)(q32),del(3)(q12),add(4)(q21),del(5)(q22q35),-7 ,der(12)t(12;13)(p13;q12),-13,-13,-16,-18,+mar1[cp20]	SAMD9L	L1323fs	50%	90%
UPN006	46,XY[20]	SAMD9	Q154X, I268T	46%, 47%	40%, 41%
UPN007	46,XX[20]	SAMD9	H32L	38%	45%
UPN012	46,XX,del(5)(q13q33)[14]/46,XX[6]	SAMD9	T205P	39%	49%
UPN013	46,XX,del(5)(q13q33)[6]/46,XX[14]	SAMD9	T205P	47%	46%
UPN014	47, XX, +21[20]	SAMD9	R221Q	54%	48%
UPN015	46,XY[20]	SAMD9	I268T	44%	47%
UPN017	46,XY[cp30]	SAMD9	D550V	44%	51%
UPN020	46, XY[20]	SAMD9	N1244D	42%	48%
UPN022	45, X, -Y[20]	SAMD9L	L50S	48%	49%
UPN023	46,XX[20]	SAMD9L	P123L	46%	52%
UPN024	46,XY,del(5)(q13q33)[1]/46,XX[4]	SAMD9L	E220G	47%	46%
UPN025	46,XY,del(5)(q13q33),-18,del(20)(q11.2q13.1)[4]/46,XY[16]	SAMD9L	G235S	45%	47%
UPN026	46,XX[20]	SAMD9L	W333C	-	45%
UPN028	46,XX[20]	SAMD9L	A637T	-	53%
UPN029	46,XY[20]	SAMD9L	S728P, Y705C	48%, 51%	55%, 47%
UPN_ADD1	46,XX[20]	SAMD9	A949S	-	36%
UPN_ADD2	44-46,XX, del(5)(q21q23),add(7)(q11.2), add(12)(p13), +1-2 mar(CP)[20]	SAMD9	R106H	-	53%
UPN_ADD3	46,XY[20]	SAMD9	R902Q	-	49%
UPN_ADD4	46,XX,del(20)(q11.2q13.1)[5]/46,XX[15]	SAMD9L	R1298G	-	54%
UPN_ADD5	46,XY[20]	SAMD9L	A637S	-	45%
UPN_ADD6	42-44,XY,der(5)del(5)(q13.3)t(5;12)(q13.3;q13), der(12)t(5;12),-15,-16, -17,-18, der(19)t(17;19)(q21;p13.3),add(20)(q13.3),add(21)(q22)[cp20]	SAMD9L	G247A	-	48%
UPN_ADD7	46,XY[20]	SAMD9	T1370fs	-	51%

"-" depicts germline DNAs were not available. Abbreviations; VAFs, variant allele frequencies

Gono	Mutants	Prediction	Deteced cohort	Results of cell
Gene			Deleced conon	proliferation assay
SAMD9	T205P	Pathogenic missense	Our MDS cohort	LOF
SAMD9	I247T	Pathogenic missense	Public database	LOF
SAMD9	I268T	Pathogenic missense	Our MDS cohort and public database	WT
SAMD9	D550V	Pathogenic missense	Our MDS cohort	WT
SAMD9	L574P	Pathogenic missense	Public database	LOF
SAMD9L	L50S	Pathogenic missense	Our MDS cohort	WT
SAMD9L	C228Y	Pathogenic missense	Public database	LOF
SAMD9L	E220G	Pathogenic missense	Our MDS cohort	LOF
SAMD9L	G235S	Pathogenic missense	Our MDS cohort	LOF
SAMD9L	G247A	Pathogenic missense	Our MDS cohort	WT
SAMD9L	W333C	Pathogenic missense	Our MDS cohort	LOF
SAMD9L	W517R	Pathogenic missense	Index familal cases	LOF
SAMD9L	L1323fs	Frameshift	Our MDS cohort	LOF

Supplemental Table 11. Summary of cell proliferation assay (n=13)

Supplemental Table 12. Comparison of pathogenic features of GL SAMD9/SAM	9L variants
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Parameters	Pediatric MDS	Adult MDS
Distribution of genetic variants	C-terminus	N-terminus
Genetic reversion	Frequent	Rare
Latency	Short	Long
Function of GL alterations	Gain-of-function	Loss-of-function