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High-quality reference genome of the Siamese fighting fish Betta splendens, a model species for the study of aggression --Manuscript Draft--

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Abstract:	Background: Siamese fighting fish Betta spl aggressiveness and accordingly have been the lack of a reference genome has so far lin basis of aggression in this species. Here we assembly of the Siamese fighting fish. Findings: We first sequenced and de novo a splendens variety Giant, with a weighted av an N50 contig size of 19.01 Kb, covering 99 obtain a chromosome-level genome assemi sequenced 69.7 Gb reads using the BGISE approximately 93% of the scaffold sequence quality of our assembly using the high conta also performed comparative chromosome a splendens, revealing a chromosome conser predicted a total of 23,202 genes assisted b liver, muscle and heart tissues of Giant, and the genome. Additionally, we resequenced of detected ~3.4M single-nucleotide variations Conclusions: We provide the first chromoso fish. The genome will lay a valuable foundat splendens. Keywords: Betta splendens; fish genome; a assembly; resequencing	endens are notorious for their widely used to study aggression. However, mited the understanding of the genetic present the first reference genome assembled a 465.24 Mb genome for the B. erage (N50) scaffold size of 949.03 Kb and 0.93% of the estimated genome size. To bly, we constructed one Hi-C library and Q-500 platform. We anchored es into 21 chromosomes and evaluated the act frequency heatmap and BUSCO. We nalyses between Oryzias latipes and B. vation evolution in B. splendens. We by RNA-seq data generated from brain, d annotated 15% repetitive sequences in other five B. splendens varieties and (SNVs) and 27,305 indels. me-level genome for the Siamese fighting tion for future research on aggression in B.		
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6 7	2	splendens, a model species for the study of aggression				
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38 39	19					
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42	21	Abstract				
43 44	22	Background: Siamese fighting fish Betta splendens are notorious for their				
45 46	23	aggressiveness and accordingly have been widely used to study aggression. However, the				
47 48	24	lack of a reference genome has so far limited the understanding of the genetic basis of				
49 50	25	aggression in this species. Here we present the first reference genome assembly of the				
51 52	26	Siamese fighting fish.				
53 54	27	Findings: We first sequenced and <i>de novo</i> assembled a 465.24 Mb genome for the <i>B</i> .				
55	28	splendens variety Giant, with a weighted average (N50) scaffold size of 949.03 Kb and				
56 57	29	an N50 contig size of 19.01 Kb, covering 99.93% of the estimated genome size. To				
58 59 60 61 62 63	30	obtain a chromosome-level genome assembly, we constructed one Hi-C library and				
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sequenced 75.24 Gb reads using the BGISEQ-500 platform. We anchored approximately 93% of the scaffold sequences into 21 chromosomes and evaluated the quality of our assembly using the high contact frequency heatmap and BUSCO. We also performed comparative chromosome analyses between Oryzias latipes and B. splendens, revealing a chromosome conservation evolution in B. splendens. We predicted a total of 23,202 genes assisted by RNA-seq data generated from brain, liver, muscle and heart tissues of Giant, and annotated 15% repetitive sequences in the genome. Additionally, we resequenced other five B. splendens varieties and detected ~3.4M single-nucleotide variations (SNVs) and 27,305 indels.

40 Conclusions: We provide the first chromosome-level genome for the Siamese fighting
41 fish. The genome will lay a valuable foundation for future research on aggression in *B*.
42 *splendens*.

Keywords: Betta splendens; fish genome; aggression; Hi-C; chromosomal genome
assembly; resequencing

Data Description

Males of the Siamese fighting fish *Betta splendens* are notorious for their aggressiveness. In nature, males establish and vigorously defend territories where they construct a bubble nest to hold fertilized eggs. In laboratory settings, males will readily attack an opponent, their mirror image, physical models of conspecifics or video images of other males, and accordingly the species has been widely used to study the neurobiological mechanisms of aggression. However, the lack of a reference genome limited so far studies on the genetic basis of aggression in *B. splendens*. The species is also one of the most relevant for the ornamental fish trade as it is easy to keep and reproduce in captivity and throughout its long domestication period many varieties have been selected for their exuberant fins and colors, size or aggressive behavior. Here, we sequenced the genome of B. splendens to provide the genomic foundation for future research on aggression and development of genomic tools.

Sampling and sequencing

We purchased five different varieties of adult male Siamese fighting fish including Giant, Half-moon (HM), Half-moon plakat (HMPK), Fighter, and Elephant Ear (EE) from HK supplier TC Northern Betta (Supplementary Fig. 1). We constructed and sequenced five DNA libraries for the *B. splendens* variety Giant, including three short insert size libraries and three mate-pair libraries (Supplementary Table 1), and five RNA-seq libraries (Supplementary Table 2) using the HiSeq 2000 sequencing platform. One Hi-C library for Giant was also constructed and sequenced using the BGISEQ-500 sequencing platform, yielding 75.24 Gb of reads. Additionally, we sequenced five short insert size DNA libraries for the other five *B. splendens* varieties.

Genome assembly

We obtained 52.34 Gb of clean reads using SOAPnuke¹ with strict parameters, including removal of low-quality reads, adapter contamination and PCR duplicates. Then, we performed the *de novo* assembly of the Giant high-quality reads using the SOAP*denovo*² assembler. For the genome assembly, the short insert size libraries were used to construct the contig sequences and the mate-paired libraries were used to link the scaffolds. We

filled the gaps within the scaffolds using Gapcloser. We obtained a genome with a size of 465.24 Mb, with an N50 scaffold size of 949.03 Kb and an N50 contig size of 19.01 Kb (Table 1), covering 99.93% of the estimated genome size of 465.55 Mb using kmer analysis (Supplementary Table 3 and Supplementary Fig. 2). To construct the reference genome at the chromosome-level, we used a MBOI endonuclease to cut the DNA, and constructed a Hi-C library based on a previous protocol³. We sequenced 75.24 Gb of data using the BGISEQ-500 sequencing platform, and obtained 34.5Gb valid reads (~45.8%) that could be used to anchor the scaffolds into chromosomes after quality control using the HiC-Pro pipeline⁴ (Supplementary Fig. 3-7). Lastly, we constructed 21 chromosomes that occupied 95.3% of the genome (Fig. 1, Table 1 and Supplementary **Table 4**) using Juicer⁵ and 3D-dna pipeline⁶ based on the draft genome assembly. To evaluate the quality of the assembly, we found 95.4% of BUSCO genes that could be completely covered by our genome (**Table 2**) and approximately 98% of the transcripts assembled from four RNA-seq data could be aligned against the genome with more than 90% coverage, revealing the high quality of the assembly (Supplementary Table 5).

Genome annotation

We annotated the repetitive sequences by combining *de novo-* and homolog-based approaches. We firstly used LTR-FINDER⁷ and RepeatModeler to construct a repetitive sequence library, and then used RepeatMasker⁸ to classify these repeat sequences. We also detected repetitive sequences using RepeatMasker and ProteinMasker based on the Repbase library⁹. We identified a total of 15.12% transposable elements in the genome (**Supplementary Table 6**).

For the protein-coding prediction we combined several approaches: 1) gene model prediction using Augustus¹⁰ and GENSCAN¹¹; 2) gene prediction using GeneWise¹² based on the alignment results of protein sequences of other published species against our assembly; and 3) four RNA-seq data samples were used to assist in predicting the gene structure with Cufflinks¹³. Lastly, we integrated all of this evidence into a nonredundancy gene set using GLEAN¹⁴. The final gene set contained 23,202 genes (Supplementary Table 7), close to the number for *Oryzias latipes*¹⁵ (24,674) and slightly less than that for Danio rerio¹⁶ (26,046). We identified 90% of the 2,586 BUSCO gene

models to be complete in the gene set (Table 2), indicating the high-quality assembly andannotation.

112 Comparative genomic analysis

We compared the fighting fish genome with other species, both at the whole genome- and gene-level. All of the 21 chromosomes assembled for the fighting fish could be matched to chromosomes of Oryzias latipes with a mean coverage ratio of 75.3%. From these, 18 chromosomes had a single hit to one chromosome of O. latipes, and 3 chromosomes (1, 19 and 21) had a hit in two chromosomes of O. latipes (Fig. 2 and Supplementary Table 8), indicating conservative evolution for most of chromosomes, as well as several chromosome reshuffling events between these two species. Furthermore, from the gene set level, KO (KEGG Orthology) terms of animals from 109 different species were counted and compared with the fighting fish gene set using the KEGG database¹⁷ (version 79). There were five KO terms notably expanded in fighting fish compared with all other animals, including 147 NACHT, LRR and PYD domains-containing protein 3 (NLRP3, K12800), 86 tripartite motif-containing protein 47 (TRIM47, K12023), 43 chloride channel 7 (CLCN7, K05016), 29 arginine vasopressin receptor 2 (AVPR2, K04228) and 17 maltase-glucoamylase (MGAM, K12047) (Fig. 3). NLRP3 has two prominent expansions, corresponding to clade 1, containing 56 genes, and clade2, containing 79 genes, whereas other fish species in these two clades have less than three gene copies (Fig. 4). NLRP3 encodes a pyrin-like protein containing a pyrin domain, a nucleotide-binding site (NBS) domain, and a leucine-rich repeat (LRR) motif, and plays a role in the regulation of inflammation, the immune response, and apoptosis 18 .

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133 Resequencing

To evaluate the genetic diversity among the four varieties of *Betta splendens*, we called the SNVs (single-nucleotide variations) and Indels (insertions and deletions) based on the read alignment result using Giant assembly as a reference. We obtained 70.25 Gb of clean reads filtered from 79.18 Gb of raw reads (**Supplementary Table 9**). We used BWA¹⁹ (v0.6.2) to align all the re-sequencing data to the reference genome and the UnifiedGenotyper in Genome Analysis Toolkit (GATK²⁰, v2.8.1) to call variations. In

total, we detected approximately 3.4 M SNVs and 27,305 indels, which will provide б abundant genetic polymorphism for use in future research and applications. Moreover, we found that the cluster result was in accordance with the behavior shown by each B. splendens variety, by constructing a phylogenetic tree and performing a PCA analysis (Supplementary Fig. 8). Availability of data We have deposited the data into the GenBank under the BioProject number PRJNA416843. Abbreviations bp: base pair; Gb: gigabases; Kb: kilobases; KO: KEGG Orthology; M: million; Mb: megabases; PCA: principal component analysis; SNV: single-nucleotide variation Acknowledgements We thank the Macau Science and Technology Development Fund for financial support (project 011/2014/A1), and Shenzhen Municipal Government China(JCYJ20151015162041454 to W.C. and JCYJ20150529150505656 to X.L.). **Authors contributions** G.F., S.L. and J.C. conceived the project. G.F., X.L. and J.C. supervised the research. H.Z., C.S. and X.Y. conceived and designed the experiments. K.M., X.L. and B.Y. performed genome assembly and gene annotation. H.L., Z.R., Q.L. and Q.X. prepared the fighting fish sample. Jiahao.W., W.C., X.X. and L.S. performed sequencing. A.R., M.G., Jing.C., H.Y. and J.W. performed comparative genomic analysis. G.F., S.C., Y. W. and D.G. revised the paper.K.M. and X.Y. performed data accession. Tables Table 1. Statistics of the assembly using SOAPdenovo and Hi-C data.

of

T	Scaffold	Contig	Scaffold	Contig (Hi-C)	
Гуре	Original	Original	(Hi-C)		
Total Number	92,886	138,929	3,057	48,943	
Toltal length (bp)	465,240,853	421,527,246	452,537,980	409,013,355	
Average length (bp)	5008.73	3034.12	19,754,490	19,674	
N50 Length (bp)	949,032	19,014	35,205,731	189,132	
N90 Length (bp)	59,769	3,504	14,840,089	4,434	

Table 2. Evaluation results of the genome and gene set using BUSCO.

	Genome		Genes	
	Number	Percentage (%)	Number	Percentage (%)
Complete	2,466	95.4	2,327	90.0
Single-copy complete	2,422	93.7	2,254	87.2
Duplicated complete	44	1.7	73	2.8
Fragmented	69	2.7	204	7.9
Missing	51	1.9	55	2.1
Total	2,586	-	2,586	-

Figure legends

174 Fig. 1. Hi-C interaction heatmap for *B. Splendens* reference genome, showing
175 interactions between the 21 chromosomes.

Fig. 2. Collinear relationship between *B. splendens* and *Oryzias latipes*. Green represents
the chromosomes of *B. splendens* and the other multicolor represent the chromosomes of *O. latipes*.

⁵¹ 52 180

Fig. 3. Five gene families with prominent expansion in *B. splendens* when compared withother species.

Fig. 4. The gene phylogenetic tree of NLRP3 gene family (KO: K12800) using the genes

185 of *B. splendens* and other species. Clade 1 and clade 2 show two prominent expansion

186 sub-families of *B. splendens*.

References

 Chen, Y. *et al.* SOAPnuke: a MapReduce acceleration-supported software for integrated quality control and preprocessing of high-throughput sequencing data.
 GigaScience 7, 1-6 (2018).

192 2. Luo, R. *et al.* SOAPdenovo2: an empirically improved memory-efficient short-193 read de novo assembler. *Gigascience* **1**, 18 (2012).

- 1943.Durand, N.C. *et al.* Juicer Provides a One-Click System for Analyzing Loop-195195Resolution Hi-C Experiments. *Cell Syst* **3**, 95-8 (2016).
- ⁴ 196 4. Servant, N. *et al.* HiC-Pro: an optimized and flexible pipeline for Hi-C data ⁵ 197 processing. *Genome Biol* **16**, 259 (2015).
- 1985.Belton, J.M. *et al.* Hi-C: a comprehensive technique to capture the conformation8199of genomes. *Methods* 58, 268-76 (2012).
- 292006.Dudchenko, O. *et al.* De novo assembly of the Aedes aegypti genome using Hi-C30201yields chromosome-length scaffolds. Science **356**, 92-95 (2017).
- 12027.Xu, Z. & Wang, H. LTR_FINDER: an efficient tool for the prediction of full-13203203length LTR retrotransposons. Nucleic Acids Res 35, W265-8 (2007).
- 342048.Tarailo-Graovac, M. & Chen, N. Using RepeatMasker to identify repetitive
elements in genomic sequences. Curr Protoc Bioinformatics Chapter 4, Unit 4
10 (2009).
- 2079.Jurka, J. et al. Repbase Update, a database of eukaryotic repetitive elements.208208Cytogenet Genome Res 110, 462-7 (2005).
- 20910.Stanke, M. et al. AUGUSTUS: ab initio prediction of alternative transcripts.12210Nucleic Acids Res 34, W435-9 (2006).
- 1221111.Burge, C. & Karlin, S. Prediction of complete gene structures in human genomic14212DNA. J Mol Biol 268, 78-94 (1997).
- 15 213 12. Birney, E. GeneWise and Genomewise. *Genome Research* 14, 988-995 (2004).
- ⁶ 214 13. Trapnell, C. *et al.* Differential gene and transcript expression analysis of RNA-seq
 ⁷ 215 experiments with TopHat and Cufflinks. *Nature Protocols* 7, 562-578 (2012).
- ⁴⁰ 216 14. Elsik, C.G. *et al.* Creating a honey bee consensus gene set. *Genome Biol* **8**, R13 (2007).
- 218 15. Kasahara, M. *et al.* The medaka draft genome and insights into vertebrate genome evolution. *Nature* 447, 714-719 (2007).
- ⁵³₅₄ 220 16. Howe, K. *et al.* The zebrafish reference genome sequence and its relationship to the human genome. *Nature* **496**, 498-503 (2013).
- 56 222 17. Kanehisa, M. The KEGG Database. **247**, 91-103 (2002).
- ⁵⁷ 223 18. Hirota, S.A. *et al.* NLRP3 inflammasome plays a key role in the regulation of intestinal homeostasis. *Inflammatory Bowel Diseases* 17, 1359-1372 (2011).

1 2			
3 4	225	19.	Li, H. & Durbin, R. Fast and accurate short read alignment with Burrows-Wheeler
5 6	226	20	transform. <i>Bioinformatics</i> 25 , 1754-1760 (2009).
7 8	227 228	20.	analyzing next-generation DNA sequencing data. Genome Research 20, 1297-
9 10	229		1303 (2010).
11 12	230		
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14 15			
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Supplementary Material

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