

Table S1. Biofilm screening conditions tested on SK and ON microbial isolates.

Bacterial species (No of isolates)^a	Biofilm media tested^b	Temperature Time	Reference^c
<i>Staphylococcus Epidermidis</i> (77)	TSB	37°C	(1, 2)
	TSB + 0.5%, 1% or 2% glucose	24h or 48h	
	0.5 TSB + 1% glucose		(3)
	TSB + 1% glucose + 4% NaCl		
	MHII + 2% NaCl		(4)
	BHI + 1% glucose + 4% NaCl,		
	M9 + 0.25% glucose + 0.5% CAA		
	M9 + 0.5% glucose + 1% CAA		
M9 + 1.0% glucose + 2% CAA			
M63 + 0.3% glucose + 0.5% CAA			
<i>Staphylococcus aureus</i> (10)	TSB	37°C	(1)
	TSB + 0.5%, 1%, 1.5%, or 2% glucose	24h or 48h	
	TSB + 1% glucose + 4% NaCl		
	MHII + 2% NaCl		(4)
	BHI + 0.5%, 1% or 2% glucose + 1% NaCl		
	BHI + 1%, 2% or 3% glucose + 4% NaCl		(5)
	M9 + 0.25%, 0.5%, or 1% glucose + 0.5%, 1% or 2% CAA		
	M63 + 0.3% glucose + 0.5% CAA		
<i>Enterococcus faecalis</i> (5)	TSB	37°C	
	TSB + 0.25%, 0.5%, 1%, or 2% glucose	24h or 48h	(6, 7)
	TSB + 1% glucose + 4% NaCl		
	MHII + 2% NaCl		
	BHI + 0.5%, 1% or 2% glucose + 1% NaCl		
	BHI + 1%, 2% or 3% glucose + 4% NaCl		
	M9 + 0.25%, 0.5% or 1% glucose + 0.5%, 1% or 2% CAA		
	M9 + 0.5% yeast extract + 1% CAA		(8)
M63 + 0.3% glucose + 0.5% CAA			
<i>Escherichia coli</i> (5)	TSB	37°C	(9)
	TSB + 0.5%, 1%, or 2% glucose	24h or 48h	
	TSB + 1% glucose + 4% NaCl		
	MHII + 2% NaCl		
	BHI + 1% glucose + 4% NaCl		
	M9 + 0.1%, 0.25%, 0.5% or 1% glucose + 0.2%, 0.5%, 1% or 2% CAA		
	M63 + 0.3% glucose + 0.5% CAA		(10)
	LB		

Bacterial species (No of isolates) ^A	Biofilm media tested ^B	Temperature Time	Reference ^C
<i>Stenotrophomonas maltophilia</i> (3)	TSB	37°C	(11)
	TSB + 0.5% glucose	24h or 48h	
<i>Pseudomonas aeruginosa</i> (3)	M9 + 0.25% glucose + 0.5% CAA		(12)
	M63 + 0.3% glucose + 0.5% CAA		
	LB		
<i>Enterobacter agglomerans</i> (4)	BHI + 0.5%, 1% or 2% glucose + 1% NaCl	37°C	
	BHI + 1%, 2% or 3% glucose + 4% NaCl	24h or 48h	
	M9 + 0.1% or 0.2% glucose		
	M9 + 0.1% glucose + 0.5% or 1% CAA		
	M9 + 0.2% glucose + 0.2% CAA		
	M63 + 0.3% glucose + 0.5% CAA		
	LB		
<i>Serratia marcescens</i> (3)	TSB	37°C	
	M9 + 0.25% glucose + 0.5% CAA	24h or 48h	
	M9 + 0.5% glucose		(13)
	LB		(14)
<i>Proteus mirabilis</i> (3)	TSB	37°C	
	TSB + 0.5% glucose	24h or 48h	
	M63 + 0.3% glucose + 0.5% CAA		
	M9 + 0.25% glucose + 0.5% CAA		
	LB		
<i>Klebsiella pneumoniae</i> (3)	TSB	37°C	
	1/2 TSB	24h or 48h	
	M9 + 0.25% glucose + 0.5% CAA		(15)
	LB		(16)
<i>Candida albicans</i> (22)	M9 + 0.25% glucose + 0.5% CAA	28°C or 37°C	
<i>Candida glabrata</i> (7)	RPMI 1640 + MOPS pH7	24h or 48h	(17)
	TSB		
	YPD + 2% glucose		(18)
	YNB + 0.9% glucose		(18)
<i>Salmonella serovar Typhimurium</i> (1)	1/2 LB (no salt) + 40uM 2,2'-dipyridyl	28°C or 37°C	(19)
		24h or 48h	

^a Each species is listed with the number of isolates screened in parentheses

^b Abbreviations: TSB, Tryptic soy broth; MHII, Mueller Hinton II Broth; M9, M9 minimal media; CAA, Casamino acids; M63, M63 minimal media; BHI, Brain Heart Infusion; LB, Lysogeny broth; MOPS, 3-(*N*-morpholino)propanesulfonic acid; YPD, yeast extract-peptone-dextrose; YNB, yeast nitrogen base

^c References were used as the starting point for choosing growth media. For consistency of testing, each strain was grown in TSB.

REFERENCES

1. **Christensen GD, Simpson WA, Younger JJ, Baddour LM, Barrett FF, Melton DM, Beachey EH.** 1985. Adherence of coagulase-negative staphylococci to plastic tissue culture plates: a quantitative model for the adherence of staphylococci to medical devices. *Journal of Clinical Microbiology* **22**:996–1006.
2. **Ziebuhr W, Heilmann C, Götz F, Meyer P, Wilms K, Straube E, Hacker J.** 1997. Detection of the intercellular adhesion gene cluster (*ica*) and phase variation in *Staphylococcus epidermidis* blood culture strains and mucosal isolates. *Infection and Immunity* **65**:890–896.
3. **Opperman TJ, Kwasny SM, Williams JD, Khan AR, Peet NP, Moir DT, Bowlin TL.** 2009. Aryl rhodanines specifically inhibit staphylococcal and enterococcal biofilm formation. *Antimicrobial Agents and Chemotherapy* **53**:4357–4367.
4. **Saginur R, Stdenis M, Ferris W, Aaron SD, Chan F, Lee C, Ramotar K.** 2006. Multiple combination bactericidal testing of staphylococcal biofilms from implant-associated infections. *Antimicrobial Agents and Chemotherapy* **50**:55–61.
5. **O'Neill E, Pozzi C, Houston P, Humphreys H, Robinson DA, Loughman A, Foster TJ, O'Gara JP.** 2008. A novel *Staphylococcus aureus* biofilm phenotype mediated by the fibronectin-binding proteins, FnBPA and FnBPB. *Journal of Bacteriology* **190**:3835–3850.
6. **Toledo-Arana A, Valle J, Solano C, Arrizubieta MJ, Cucarella C, Lamata M, Amorena B, Leiva J, Penadés JR, Lasa I.** 2001. The enterococcal surface protein, Esp, is involved in *Enterococcus faecalis* biofilm formation. *Applied and Environmental Microbiology* **67**:4538–4545.
7. **Holmberg A, Mörgelin M, Rasmussen M.** 2012. Effectiveness of ciprofloxacin or linezolid in combination with rifampicin against *Enterococcus faecalis* in biofilms. *J Antimicrob Chemother* **67**:433–439.
8. **Dunny GM, Clewell DB.** 1975. Transmissible toxin (hemolysin) plasmid in *Streptococcus faecalis* and its mobilization of a noninfectious drug resistance plasmid. *Journal of Bacteriology* **124**:784–790.
9. **Carneiro VA, Santos HSD, Arruda FVS, Bandeira PN, Albuquerque MRJR, Pereira MO, Henriques M, Cavada BS, Teixeira EH.** 2010. Casbane diterpene as a promising natural antimicrobial agent against biofilm-associated infections. *Molecules* **16**:190–201.
10. **Chassaing B, Darfeuille-Michaud A.** 2013. Adherent-invasive *Escherichia coli* Biofilm Formation Assays. *BIO-PROTOCOL* **3**: e982.
11. **Peeters E, Nelis HJ, Coenye T.** 2008. Comparison of multiple methods for quantification of microbial biofilms grown in microtiter plates. *J Microbiol Methods* **72**:157–165.

12. **Drenkard E, Ausubel FM.** 2002. Pseudomonas biofilm formation and antibiotic resistance are linked to phenotypic variation. *Nature* **416**:740–743.
13. **Koh KS, Lam KW, Alhede M, Queck SY, Labbate M, Kjelleberg S, Rice SA.** 2007. Phenotypic diversification and adaptation of *Serratia marcescens* MG1 biofilm-derived morphotypes. *Journal of Bacteriology* **189**:119–130.
14. **Shanks RMQ, Stella NA, Kalivoda EJ, Doe MR, O'Dee DM, Lathrop KL, Guo FL, Nau GJ.** 2007. A *Serratia marcescens* OxyR Homolog Mediates Surface Attachment and Biofilm Formation. *Journal of Bacteriology* **189**:7262–7272.
15. **Chhibber S, Nag D, Bansal S.** 2013. Inhibiting biofilm formation by *Klebsiella pneumoniae* B5055 using an iron antagonizing molecule and a bacteriophage. *BMC Microbiol* **13**:174.
16. **Maldonado NC, de Ruiz CS, Cecilia M, Nader-Macias ME.** 2007. A simple technique to detect *Klebsiella* biofilm-forming-strains. Inhibitory potential of *Lactobacillus fermentum* CRL 1058 whole cells and products. *Communicating Current Research and Educational Topics and Trends in Applied Microbiology* 52–58.
17. **Melo AS, Bizerra FC, Freymüller E, Arthington-Skaggs BA, Colombo AL.** 2011. Biofilm production and evaluation of antifungal susceptibility amongst clinical *Candida* spp. isolates, including strains of the *Candida parapsilosis* complex. *Medical Mycology*, **49**:253–262.
18. **Serrano-Fujarte I, López-Romero E, Reyna-López GE, Martínez-Gámez MA, Vega-González A, Cuéllar-Cruz M.** 2015. Influence of Culture Media on Biofilm Formation by *Candida* Species and Response of Sessile Cells to Antifungals and Oxidative Stress. *BioMed Research International* **2015**:1–15.
19. **White AP, Gibson DL, Grassl GA, Kay WW, Finlay BB, Vallance BA, Surette MG.** 2008. Aggregation via the Red, Dry, and Rough Morphotype Is Not a Virulence Adaptation in *Salmonella enterica* Serovar Typhimurium. *Infection and Immunity* **76**:1048–1058.