

## **Supplementary information**

### **Diverse novel resident *Wolbachia* strains in Culicine mosquitoes from Madagascar**

Claire L Jeffries, Luciano M Tantely, Fara N Raharimalala, Eliot Hurn, Sébastien Boyer & Thomas Walker

**Supplementary table S1.** Additional sample details and CO1 GenBank accession numbers

**Supplementary table S2.** Additional sample details and *Wolbachia* 16S, wsp and MLST gene GenBank accession numbers.

**Supplementary table S3.** Summary of qualitative characteristics, median humidity, median temperature and altitude of collection sites.

**Supplementary table S4.** Arbovirus screening assays including PCR primer/probes sequences and cycling conditions used to screen mosquitoes.

**Supplementary table S1. Additional sample details and CO1 GenBank accession numbers.** The sample code, location and species are shown, in addition to the *Wolbachia* / RVFV status and GenBank CO1 accession number.

| Sample ID       | Alternative ID                | Location       | Species                        | <i>Wolbachia</i> / RVFV status | CO1 accession number |
|-----------------|-------------------------------|----------------|--------------------------------|--------------------------------|----------------------|
| TSA-AMAD-1      | Aedo. madagascarica (W+)      | Tsaramandroso  | <i>Aedeomyia madagascarica</i> | W+                             | MK033247             |
| TSA-CANT-1      | Cx. antennatus (W+) (RVFV+)   | Tsaramandroso  | <i>Culex antennatus</i>        | W+ / RVFV+                     | MK033248             |
| TSA-CDEC-1      | Cx. decens (W+) 1             | Tsaramandroso  | <i>Culex decens</i>            | W+                             | MK033249             |
| TSA-CDEC-2      | Cx. decens (W+) 2             | Tsaramandroso  | <i>Culex decens</i>            | W+                             | MK033250             |
| TSA-CDEC-3      | Cx. decens (W+) 3             | Tsaramandroso  | <i>Culex decens</i>            | W+                             | MK033251             |
| TSA-CDUT-1      | Cx. duttoni (W+)              | Tsaramandroso  | <i>Culex duttoni</i>           | W+                             | MK033252             |
| TSA-CTRI-1      | Cx. tritaeniorhynchus (RVFV+) | Tsaramandroso  | <i>Culex tritaeniorhynchus</i> | W- / RVFV+                     | MK033253             |
| TSA-MUNI-1      | Man. uniformis (W+)           | Tsaramandroso  | <i>Mansonia uniformis</i>      | W+                             | MK033254             |
| TSA-USP1-1      | Ura. sp. 1 (W+)               | Tsaramandroso  | <i>Uranotaenia</i> sp. 1       | W+                             | MK033255             |
| TSA-USP2-1      | Ura. sp. 2 (W+) 1             | Tsaramandroso  | <i>Uranotaenia</i> sp. 2       | W+                             | MK033256             |
| TSA-USP2-2      | Ura. sp. 2 (W+) 2             | Tsaramandroso  | <i>Uranotaenia</i> sp. 2       | W+                             | MK033257             |
| CU1-ANK-CSPI-1  | Culex sp. 1 (W-) 1            | Ankazobe       | <i>Culex</i> sp. 1             | W-                             | MK033258             |
| CU2-ANK-CUNI-1  | Cx. univittatus (W-) 1        | Ankazobe       | <i>Culex univittatus</i>       | W-                             | MK033259             |
| CU3-ANK-CPIP-1  | Cx. pipiens (W-) 1            | Ankazobe       | <i>Culex pipiens</i>           | W-                             | MK033260             |
| CU4-ANK-CUNI-2  | Cx. univittatus (W-) 2        | Ankazobe       | <i>Culex univittatus</i>       | W-                             | MK033261             |
| CU5-ANK-CUNI-3  | Cx. univittatus (W-) 3        | Ankazobe       | <i>Culex univittatus</i>       | W-                             | MK033262             |
| CU6-ANK-CSP1-2  | Culex sp. 1 (W-) 2            | Ankazobe       | <i>Culex</i> sp. 1             | W-                             | MK033263             |
| CU7-TSA-CSPI-1  | Culex sp. 1 (W-) 3            | Tsaramandroso  | <i>Culex</i> sp. 1             | W-                             | MK033264             |
| CU8-TSA-CPIP-1  | Cx. pipiens (W-) 2            | Tsaramandroso  | <i>Culex pipiens</i>           | W-                             | MK033265             |
| CU9-ANK-CPIP-2  | Cx. pipiens (W-) 3            | Ankazobe       | <i>Culex pipiens</i>           | W-                             | MK033266             |
| CU10-ANK-CPIP-3 | Cx. pipiens (W-) 4            | Ankazobe       | <i>Culex pipiens</i>           | W-                             | MK033267             |
| CU11-TSA-CPIP-2 | Cx. pipiens (W-) 5            | Tsaramandroso  | <i>Culex pipiens</i>           | W-                             | MK033268             |
| CU12-IVA-CPIP-1 | Cx. pipiens (W-) 6            | Ivato Aeroport | <i>Culex pipiens</i>           | W-                             | MK033269             |

**Supplementary table S2. Additional sample details and *Wolbachia* 16S, wsp and MLST gene GenBank accession numbers.** Sample codes, *Wolbachia* strain names and *Wolbachia* MLST gene sequence GenBank accession numbers.

| Sample ID  | Location       | Host species                   | Strain    | 16S      | wsp      | gatB     | coxA     | hcpA     | ftsZ     | fbpA     |
|------------|----------------|--------------------------------|-----------|----------|----------|----------|----------|----------|----------|----------|
| TSA-AMAD-1 | Tsaramandroso  | <i>Aedeomyia madagascarica</i> | wMad      | MK026554 | MK033270 | MK033279 | MK033288 | MK033297 | MK033305 | MK033312 |
| TSA-AMAD-2 | Tsaramandroso  | <i>Aedeomyia madagascarica</i> | wMad      | -        | MK033271 | -        | -        | -        | -        | -        |
| TSA-AMAD-3 | Tsaramandroso  | <i>Aedeomyia madagascarica</i> | wMad      | -        | MK033272 | -        | -        | -        | -        | -        |
| TSA-CANT-1 | Tsaramandroso  | <i>Culex antennatus</i>        | wAnt      | MK026555 | -        | -        | -        | -        | -        | -        |
| TSA-CDEC-1 | Tsaramandroso  | <i>Culex decens</i>            | wDec      | MK026556 | MK033273 | MK033280 | MK033289 | MK033298 | -        | MK033313 |
| TSA-CDEC-2 | Tsaramandroso  | <i>Culex decens</i>            | wDec      | MK026557 | MK033274 | MK033281 | MK033290 | MK033299 | MK033306 | MK033314 |
| TSA-CDUT-1 | Tsaramandroso  | <i>Culex duttoni</i>           | wDutt     | MK026558 | -        | -        | -        | -        | -        | -        |
| TSA-FCIR-1 | Tsaramandroso  | <i>Ficalbia circumtestacea</i> | wCir      | -        | -        | MK033282 | MK033291 | MK033300 | MK033307 | MK033315 |
| TSA-MUNI-1 | Tsaramandroso  | <i>Mansonia uniformis</i>      | wUnif-Mad | MK026559 | -        | MK033283 | MK033292 | -        | MK033308 | MK033316 |
| ANI-USP1-1 | Anivorano Nord | <i>Uranotaenia</i> sp. 1       | wUral     | MK026560 | MK033275 | MK033284 | MK033293 | MK033301 | MK033309 | MK033317 |
| TSA-USP1-1 | Tsaramandroso  | <i>Uranotaenia</i> sp. 1       | wUra1     | MK026561 | MK033276 | MK033285 | MK033294 | MK033302 | MK033310 | MK033318 |
| TSA-USP2-1 | Tsaramandroso  | <i>Uranotaenia</i> sp. 2       | wUra2     | MK026562 | MK033277 | MK033286 | MK033295 | MK033303 | MK033311 | MK033319 |
| TSA-USP2-2 | Tsaramandroso  | <i>Uranotaenia</i> sp. 2       | wUra2     | MK026563 | MK033278 | MK033287 | MK033296 | MK033304 | MK033307 | MK033320 |

**Supplementary table S3.** Summary of qualitative characteristics, median humidity, median temperature and altitude of collection sites.

| Commune        | Village             | Nearest water bodies | Natural flora              | Natural fauna | Agriculture                     | Animal husbandry      | Construction / housing                                   | Human population density | Median humidity (25th percentile, 75th percentile) | Median temperature (25th percentile, 75th percentile) | Altitude (meters above sea level) |
|----------------|---------------------|----------------------|----------------------------|---------------|---------------------------------|-----------------------|--|--------------------------|--|---|-----------------------------------|
| Anivorano Nord | Anivorano III       | river, swamp         | forested areas             | cats, dogs    | n/a                             | zebu, poultry         | thatched roofs, sheet metal walls                        | sparsely populated       | 74 (65, 83)  | 20.5 (19.5, 22.5)                                     | 357                               |
| Tsaramandroso  | Ambomiharina        | lake                 | scattered trees, bushes    | dogs          | Rice paddies, swede, cane       | zebu, goats           | thatched roofs, wood walls                               | populated                | 57 (53,70)   | 24.0 (21.5, 28.0)                                     | 84                                |
| Bemokotra      | Antafia             | lake                 | trees, scattered shrubbery | bats, cats    | cane                            | zebu, poultry, swine  | concrete houses/wood walls thatched roofs                | populated                | 50 (40, 60)  | 26.0 (23.5, 29.5)                                     | 64                                |
| Ankazobe       | Ambohimarina        | n/a (large city)     | patches of dense greenery  | cats          | n/a                             | zebus, poultry, swine | clay walls/thatched roofs, brick walls/sheet metal roofs | population               | 79 (63, 90)  | 14.5 (13.0, 18.5)                                     | 1212                              |
| Ivato Aeroport | Ivato Imerimandroso | canals, rice paddies | grassland                  | birds         | Rice paddies, banana plantation | zebu, poultry         | concrete houses  | sparsely populated       | 88 (61, 100)                                       | 11.0 (9.0, 15.5)                                      | 1261                              |

**Supplementary table S4.** Arbovirus screening assays including PCR primer/probes sequences and cycling conditions used to screen mosquitoes.

| Arbovirus | Primer and probe sequences  | Cycling conditions  | Reference |
|-----------|---|---|-----------|
| DENV      | 5'-TTGAGTAAACYRTGCTGCCTGTAGCTC-3'<br>5'-GAGACAGCAGGATCTGGTCTYTC-3'  | 95°C for 10 min<br>50 cycles: 95°C for 10 sec, 60°C for 10 sec, 72°C for 10 sec | [1]       |
| ZIKV      | 5'-CCGCTGCCAACACAAG-3'<br>5'-CCACTAACGTTCTTGCAAGACAT-3'<br>5'-FAM-AGCCTACCTTGACAAGCAGTCAGACACTCAA-TAMRA3' | 95°C for 10 min<br>45 cycles: 95°C for 10 sec, 52°C for 30 sec                  | [2]       |
| CHIKV     | 5'-CTCATCGCATCCGATCAG-3'<br>5'-ACATTGGCCCCACAATGAATTG-3'  | 95°C for 10 min<br>40 cycles: 95°C for 10 sec, 56°C for 10 sec, 72°C for 15 sec | [3]       |
| WNV       | 5'-CCTGTGTGAGCTGACAAACTTAGT-3'<br>5'-GCGTTTAGCATATTGACAGCC-3'   | 95°C for 10 min<br>45 cycles: 95°C for 10 sec, 60°C for 10 sec, 72°C for 20 sec | [4]       |
| YFV       | 5'-AATCGAGTTGCTAGGCAATAAACAC-3'<br>5'-TCCCTGAGCTTACGACCAGA-3'   | 95°C for 10 min<br>40 cycles: 95°C for 10 sec, 58°C for 10 sec, 72°C for 10 sec | [5]       |
| RVFV      | 5'-CTAGCCGTTCACAAACTGGG-3'<br>5'-GACTGARGAYCTGAATTGCACC-3'  | 95°C for 10 min<br>45 cycles: 95°C for 10 sec, 60°C for 10 sec, 72°C for 20 sec | [6]       |

1. Lai YL, Chung YK, Tan HC, Yap HF, Yap G, Ooi EE, et al. Cost-effective real-time reverse transcriptase PCR (RT-PCR) to screen for Dengue virus followed by rapid single-tube multiplex RT-PCR for serotyping of the virus. *J Clin Microbiol.* 2007/01/12. 2007;45: 935–941. doi:10.1128/JCM.01258-06
2. Lanciotti RS, Kosoy OL, Laven JJ, Velez JO, Lambert AJ, Johnson AJ, et al. Genetic and serologic properties of Zika virus associated with an epidemic, Yap State, Micronesia, 2007. *Emerg Infect Dis.* 2008;14: 1232–1239. doi:10.3201/eid1408.080287
3. Ali UH, Vasan SS, Thayan R, Angamuthu C, Lim LH, Sekaran SD. Development and evaluation of a one-step SYBR-Green I-based real-time RT-PCR assay for the detection and quantification of Chikungunya virus in human, monkey and mosquito samples. *Trop Biomed.* 2010;27: 611–623.
4. Linke S, Ellerbrok H, Niedrig M, Nitsche A, Pauli G. Detection of West Nile virus lineages 1 and 2 by real-time PCR. *J Virol Methods.* 2007;146: 355–358. doi:10.1016/j.jviromet.2007.05.021
5. Dash PK, Boutonnier A, Prina E, Sharma S, Reiter P. Development of a SYBR green I based RT-PCR assay for yellow fever virus: application in assessment of YFV infection in *Aedes aegypti*. *Virol J.* 2012/01/24. 2012;9: 27. doi:10.1186/1743-422X-9-27
6. Maquart M, Temmam S, Héraud J-M, Leparc-Goffart I, Cêtre-Sossah C, Dellagi K, et al. Development of real-time RT-PCR for the detection of low concentrations of Rift Valley fever virus. *J Virol Methods.* 2014;195: 92–99. doi:10.1016/j.jviromet.2013.10.001.