Table S1.List of primers used in this study

Primer names	**Primer sequences (5′-3′)**	**Purposes**
Cas9-F	CAGTAATACGACTCACTATTAGAAGCGACT CGTCTCAAACGG	Amplification template of the 111 nt *in vitro* transcribed RNA
Cas9-R	AGGATGACGTTCATGCTTCTTGT	
FAM-RT primer1	FAM-CTATCATCTACTTTCGCCATCTCA	RT primer of *in vitro* transcribed RNA
$A8_{AG}$-F	CAGTAATACGACTCACTATTAGAACAACGG CGCCGCGGTAACGCGGCGCCACGCGG	Amplification template for $A8_{AG}$
$G8_{GG}$-F	TAATACGACTCACTATAGGAACAACGGCGC CGCGGTAACGCGGCGCCACGCGG	Amplification template for $G8_{GG}$
$G8_{GA}$-F	TAATACGACTCACTATAGAAACAACGGCGC CGCGGTAACGCGGCGCCACGCGG	Amplification template for $G8_{GA}$
$G8_{GU}$-F	TAATACGACTCACTATAGTAACAACGGCGC CGCGGTAACGCGGCGCCACGCGG	Amplification template for $G8_{GU}$
$G8_{GC}$-F	TAATACGACTCACTATAGCAACAACGGCGC CGCGGTAACGCGGCGCCACGCGG	Amplification template for $G8_{GC}$
G0-F	TAATACGACTCACTATAGGCGCCGCGGTAA CGCGGCGCCACGCGG	Amplification template for G0
G1-F	TAATACGACTCACTATAGGGCGCCGCGGT AACGCGGCGCCACGCGG	Amplification template for G1
G2-F	TAATACGACTCACTATAGGGGCGCCGCGG TAACGCGGCGCCACGCGG	Amplification template for G2
G3-F	TAATACGACTCACTATAGGAGGCGCCGCG GTAACGCGGCGCCACGCGG	Amplification template for G3
G4-F	TAATACGACTCACTATAGGAAGGCGCCGC GGTAACGCGGCGCCACGCGG	Amplification template for G4
IVT-R	CAAGCAGAAGACGGCATACGAGATTCGCG TTTCCGCGTGGCGCCGCGTTACCGCGGC	Amplification template for $A8_{AG}$ to G4
FAM-RT primer 2	FAM-CAAGCAGAAGACGGCATACGAGAT	RT primer of *in vitro* transcribed RNA
TSO	ACACTCTTTCCCTACACGACGCTCTTCCGA TCTrGrGrG	Template switching oligonucleotide containing 3′ poly(rG) tail
RLM adaptor	ACACUCUUUCCCUACACGACGCUCUUCCG AUCU	RNA adaptor for RLM-RACE
outer primer	AATGATACGGCGACCACCGAGATCTACACT CTTTCCCTACACGACGCTCTTCCGATCT	Nest-PCR first round primer
inner primer	CTACACGACGCTCTTCCGATCT	Nest-PCR second round primer
E.ompA-GSP1	GTAGATGTCCAGGTCGTCAGTG	*E.coli* MG1655 *ompA* 5′ RACE
E.ompA-GSP2	GCCAACATACGGGTTAACCTGG	
E.sodB-GSP1	CGCTGCATCAGTAAACTGCGC	*E.coli* MG1655 *sodB* 5′ RACE
E.sodB-GSP2	CAGACCTGAGCTGCGTTGTTG	
E.shiA-GSP1	CCAATCAAGGCTGTCGCGATG	*E.coli* MG1655 *shiA* 5′ RACE
E.shiA-GSP2	CCGACGCCAAAGGTAGCAAATG	

E.ryhB-GSP1	AGCACCCGGCTGGCTAAG	*E.coli* MG1655 *ryhB* 5′ RACE
E.ryhB-GSP2	CTGGAAGCAATGTGAGCAATG	
E.micC-GSP1	GACGACTGTTCGGGCTTG	*E.coli* MG1655 *micC* 5′ RACE
E.micC-GSP2	GTTGGAAAATCAGTGGCAATGC	
B.ompA-GSP1	GACCTGATCCTGATCCGTATCG	*B. melitensis* 16M *ompA* 5′ RACE
B.ompA-GSP2	GCGGTCCATGTAGTTACCGATG	
B.rne-GSP1	GCCTTGGCTTCCGCTTCAAGC	*B. melitensis* 16M *rne* 5′ RACE
B.rne-GSP2	GCCAGGAAGCCATGGCGATTG	
B.rppH-GSP1	GGCGAACCATTTCTGCGTCTG	*B. melitensis* 16M *rppH* 5′ RACE
B.rppH-GSP2	CAGAAGCGAGACCGACGTCATG	
B.bsnc135-GSP1	AAAGAGCCGCAAGGGCGGCTC	*B. melitensis* 16M *bsnc135* 5′ RACE
B.bsnc135-GSP2	GAGTTTAACAGGGAGGCGTCAA	
B.bsnc149-GSP1	AAGCCCCTGCGCAATAGCGCAG	*B. melitensis* 16M *bsnc149* 5′ RACE
B.bsnc149-GSP2	CAGGTGGTTCTCGTTAATGAATG	