

Table S1. The structures and functions of current documented transcription factors in diverse insects.

TFs ^a	Structures ^b	Species	Target genes	Functions	References ^c
Abd-A	A	<i>Bombyx mori</i>	<i>WCP4</i>	Cuticle protein synthesis	[1,2]
		<i>Drosophila melanogaster</i>	—	NB apoptosis and quintessence	[3,4]
		<i>Manduca sexta</i>	—	The expression patterns in the embryos	[5]
		<i>Plutella xylostella</i>	—	Abdominal segment development	[6]
		<i>Spodoptera litura</i>	—	Embryonic development	[7]
Abd-B	A	<i>Bombyx mori</i>	—	Mating structure	[8]
		<i>Drosophila melanogaster</i>	—	Inhibition of specific NB formation in the embryonic tail region	[9]
AhR, ARNT	B	<i>Drosophila melanogaster</i>	<i>CYP6B1</i>	Xenobiotic response	[10]
		<i>Aphis gossypii</i>	<i>CYP6DA2</i>	Gossypol tolerance	[11]
		<i>Papilio polyxenes</i>	<i>CYP6B1v3</i>	Tolerance to toxic furanocoumarins	[12,13]
Amos	B	<i>Drosophila melanogaster</i>	—	Promoting olfactory sensory formation and inhibiting bristle formation	[14]
Antp	A	<i>Bombyx mori</i>	<i>sericin-3, fhxh4, fhxh5</i>	Silk synthesis	[15]
			Silk gland specific genes	The terminal differentiation of the silk gland	[16]
		<i>Bicyclus anynana</i>	—	Embryonic body plan and wing patterns	[17,18]
		<i>Drosophila melanogaster</i>	—	NB quintessence	[3,4]
			—	The role of YPWM motif in homeotic transformations	[19]
<i>Manduca sexta</i>	—	The expression patterns in the embryos	[5]		
AP-4	B	<i>Drosophila melanogaster</i>	—	Tracheal terminal branching and cell growth	[20]
		<i>Helicoverpa armigera</i>	<i>DH-PBAN</i>	Diapause	[21]
Apt-like	A	<i>Bombyx mori</i>	The single genetic <i>p</i>	Larval pigment patterns	[22]

			locus		
Ase	B	<i>Drosophila melanogaster</i>	—	CNS development	[23]
ATF-2	B	<i>Drosophila melanogaster</i>	—	Fat metabolism	[24]
ATF-2, THAP	B	<i>Aedes aegypti</i>	SCP-2	Energy metabolism	[25]
ATF-3	B	<i>Drosophila melanogaster</i>	Or47b	Olfactory system	[26]
			—	Metamorphosis	[27]
			—	Immune and metabolic homeostasis	[28]
Ato	B	<i>Drosophila melanogaster</i>	—	Chordotonal organs formation	[29]
		<i>Bombyx mori</i>	—	Molecular cloning and characterization of <i>ato</i>	[30,31]
Awh	A	<i>Bombyx mori</i>	<i>fibH</i>	Silk protein synthesis	[16,32,33]
Bcd	A	<i>Drosophila melanogaster</i>	—	Embryonic axis establishment	[34,35]
BCFI, BCFII	C	<i>Bombyx mori</i>	chorionic gene	Regulation of chorionic gene expression	[36]
bHLH TFs	B	<i>Aedes aegypti</i> <i>Anopheles gambiae</i> <i>Culex quinquefasciatus</i>	—	Genome-wide identification	[37]
		<i>Acyrtosiphon pisum</i>	—	Genome-wide identification	[38]
		<i>Apis mellifera</i>	—	Genome-wide identification	[39]
		<i>Bombyx mori</i>	—	Genome-wide identification	[40]
		<i>Diaphorina citri</i>	—	Genome-wide identification	[41]
		<i>Drosophila melanogaster</i>	—	Study of the origin and diversification of the bHLH TFs	[42]
		<i>Harpegnathos saltator</i>	—	Genome-wide identification	[43]
		<i>Leptinotarsa decemlineata</i>	—	Genome-wide identification	[44]
		<i>Nasonia vitripennis</i>	—	Genome-wide identification	[45]
		<i>Nilaparvata lugens</i>	—	Genome-wide identification	[46]
		<i>Pediculus humanus corporis</i>	—	Genome-wide identification	[47]

		<i>Plutella xylostella</i>	—	Genome-wide identification	[48]	
		<i>Tribolium castaneum</i>	—	Female reproduction	[49]	
BMFA	D	<i>Bombyx mori</i>	<i>P25</i>	Silk protein synthesis	[50]	
Bmx	B	<i>Drosophila melanogaster</i>	—	Fat metabolism	[51,52]	
BrC	C	<i>Aedes aegypti</i>	<i>vitellogenin</i>	Vitellogenesis	[53–56]	
		<i>Bombyx mori</i>	—	Nuclear import of BrC	[57]	
		<i>Bombyx mori</i>	—	Post-transcription and -translation regulation	[58]	
		<i>Nilaparvata lugens</i>	—	Ovary development	[59]	
		<i>Tribolium castaneum</i> <i>Chrysopa perla</i>	—	Metamorphosis	[60]	
BrC-Z1	C	<i>Drosophila melanogaster</i>	<i>Lcp65A-b, Edg78E</i>	Pupal cuticle protein synthesis	[61]	
			<i>phm, dib, sad</i>	Ecdysteroidogenesis	[62]	
BrC-Z2	C	<i>Bombyx mori</i>	<i>vitellogenin</i>	Reproduction	[63,64]	
			<i>WCP10, CPG13</i>	Cuticle protein synthesis	[65–67]	
BrC-Z2, BrC-Z3	C	<i>Drosophila melanogaster</i>	Activate <i>E75A</i> and repress <i>E75B</i> under nutritional shortage	Egg chamber apoptosis	[68]	
BrC-Z4	C	<i>Drosophila melanogaster</i>	<i>npc1</i>	Ecdysteroidogenesis	[69]	
			<i>Bombyx mori</i>	<i>WCP5</i>	Cuticle protein synthesis	[70]
				<i>lebocin</i>	Immune response	[71]
Btd	C	<i>Drosophila melanogaster</i>	<i>Pros</i>	Nervous system development	[72]	
Cad	A	<i>Acyrtosiphon pisum</i>	—	Early embryo development	[73]	
			<i>Drosophila melanogaster</i>	<i>ftz, spalt</i>	Embryonic development	[74–76]
				AMPs	Immune response and commensal-gut mutualism	[77–79]
				—	Embryo development	[9,35]

		<i>Bombyx mori</i>	<i>eve</i>	Embryo body segmentation	[80]
Cato	B	<i>Drosophila melanogaster</i>	—	Sensory neuron morphology	[81]
CbZ	B	<i>Bombyx mori</i>	Chorion genes	Vitellogenesis	[82]
C/EBP	B	<i>Drosophila melanogaster</i>	—	Embryogenesis; cell migration, apical constriction and epithelial invagination during oogenesis;	[83–85]
		<i>Bombyx mori</i>	Chorion genes, itself	Vitellogenesis; choriogenesis	[82,86,87]
		<i>Anopheles gambiae</i> <i>Aedes aegypti</i>	<i>def</i>	Immune response	[88]
		<i>Spodoptera litura</i>	<i>SlSCPx</i>	Energy metabolism	[89]
CncB	B	<i>Drosophila melanogaster</i>	—	Segmental diversity in the head	[90]
CncC	B	<i>Drosophila melanogaster</i>	<i>nvd, spok, dib, sad</i>	Ecdysteroidogenesis	[91]
			<i>gstD1, ref(2P), Atg8a</i>	Oxidative stress response	[92,93]
			<i>Cyp6a2, Cyp6a8, Jheh1, GstD2 and CG6188</i>	Response to phenobarbital (PB; malathion and DDT resistance)	[94,95]
			<i>Cyp6a2</i>	DDT resistance	[96]
			<i>Jheh1</i> and <i>Jheh2</i> (Transposable element Bari-Jheh was added in their promoters)	Paraquat and malathion resistance	[97]
			—	Antioxidant response,; insecticide resistance; endocrine signaling and aging	[98,99]
		<i>Tribolium castaneum</i>	CYP6BQ cluster genes	Deltamethrin resistance	[100]
	<i>CYP4G7, CYP4G14, GST-1, ABCA-UB, ABCAA1, ABCA-A1L</i>	Pyrethroid resistance	[101]		

			and <i>ABCA-9B</i>		
		<i>Leptinotarsa decemlineata</i>	<i>CYP6BJa/b</i> , <i>CYP6BJ1v1</i> , <i>CYP9Z25</i> , and <i>CYP9Z29</i>	Imidacloprid resistance and adaptation to potato plant allelochemicals	[102]
		<i>Aphis gossypii</i>	<i>CYP6DA2</i>	Gossypol tolerance	[103]
		<i>Bombyx mori</i>	—	Response after phoxim and phoxim treatment	[104,105]
Col	B	<i>Drosophila melanogaster</i>	—	Muscle identity and shape; multidendritic neuron differentiation; head development; blood cell progenitor maintenance	[106–110]
Crc	B	<i>Drosophila melanogaster</i>	—	Molting and metamorphosis	[111–113]
CREB	B	<i>Drosophila melanogaster</i>	<i>period</i>	Circadian rhythm	[114]
		<i>Drosophila melanogaster</i>	cAMP-response elements	Long-term memory	[115,116]
		<i>Bombyx mori</i>	—	Diapause process induced by environmental factors; Aluminum (Al-induced neurotoxicity or neurodegeneration)	[117,118]
		<i>Cotesia glomerata</i> <i>Cotesia rubecula</i>	—	Long-term memory formation	[119]
		<i>Apis mellifera</i>	—	Neuronal and behavioral plasticity	[120,121]
		<i>Antheraea pernyi</i>	—	Identification and function of CREB	[122]
dCREB2	B	<i>Drosophila melanogaster</i>	—	Circadian rhythm; formation of long-term memory	[114–116]
CrebA	B	<i>Drosophila melanogaster</i>	SPCGs	Embryonic SG development	[123]
		<i>Drosophila melanogaster</i>	—	Secretory capacity and activity in salivary glands	[123,124]
		<i>Drosophila melanogaster</i>	—	Nervous system development	[125]

CYC	B	<i>Nilaparvata lugens</i>	P450 genes	The susceptibility to imidacloprid	[126]
CYC, CLK	B	<i>Drosophila melanogaster</i>	<i>per, tim, vri, Pdp1ε, cwo</i>	Circadian rhythm	[127–130]
		<i>Pyrrhocoris apterus</i>	<i>Pdp1_{iso1}, cry2</i>	Adult diapause regulation	[131]
Cwo	B	<i>Drosophila melanogaster</i>	itself and other clock genes	Circadian rhythm	[132]
Da	B	<i>Drosophila melanogaster</i>	—	Photoreceptor cell determination; eye morphogenesis; cell proliferation	[133,134]
Dac	A	<i>Drosophila melanogaster</i>	—	Neuronal specification	[135]
Deaf1, lola	C	<i>Anopheles gambiae</i>	—	Immune response	[136]
Dei	B	<i>Drosophila melanogaster</i>	Muscle creatine kinase	Epidermal cells differentiate into muscle attachment sites	[137]
			<i>βPS integrin</i>	Wing development	[138]
Dichaete	D	<i>Drosophila melanogaster</i>	<i>commisureless, ase</i>	Nervous system development	[139]
Dif	D	<i>Drosophila melanogaster</i>	AMPs	Immune response	[140,141]
Dimm	B	<i>Drosophila melanogaster</i>	—	Molecular and cellular properties of all major neuroendocrine cells	[112,142–146]
			—	Protection against Gram-negative infection	[147]
		<i>Bombyx mori</i>	<i>fibH</i>	Silk protein synthesis	[148,149]
Dfd, Lab	A	<i>Drosophila melanogaster</i>	—	Embryonic development	[150]
Dip3	D	<i>Drosophila melanogaster</i>	AMPs	Immune response	[151]
Dl	D	<i>Drosophila melanogaster</i>	—	Embryonic axis establishment	[35,152]
		<i>Manduca sexta</i>	AMPs	Immune response	[153]
Dm		<i>Drosophila melanogaster</i>	<i>Sxl</i>	Sex determination	[154]
Dnato3	B	<i>Drosophila melanogaster</i>	—	Nervous system development	[155]
Dpn	B	<i>Drosophila melanogaster</i>	<i>Sxl</i>	Sex determination	[156,157]
			<i>erm</i>	Nervous system development	[158–160]

			—	Mitotic activity during optic lobe development	[161]
			—	R7 specification during eye development	[162]
Dsx	C	<i>Aedes aegypti</i>	—	Multiple sex-specific morphological, physiological, and behavioral traits of adult females; the olfactory phenotypes	[163]
		<i>Antheraea assama</i> <i>Antheraea mylitta</i>	—	Female sexual differentiation	[164]
		<i>Apis mellifera</i>	—	Reproductive division of labor	[165]
		<i>Bombyx mori</i>	—	Sex determination; sexually dimorphic traits	[166–168]
		<i>Cyclommatus metallifer</i>	—	sex-specific mandible growth	[169]
		<i>Drosophila melanogaster</i>	<i>yolk protein genes;</i> <i>bab1/2; desaturase-F</i>	Reproduction; sexually dimorphic traits; sex determination	[170–174]
		<i>Drosophila immigrans</i> species group	—	The origin and diversification of male sexual ornaments	[175]
		<i>Nilaparvata lugens</i>	—	Male BPH somatic development and mating behavior	[176]
		<i>Onthophagus taurus</i>	—	Sexual dimorphism	[177]
		<i>Ostrinia scapulalis</i>	—	Sex determination	[178]
		<i>Sciara</i>	—	Sex determination	[179]
		<i>Spodoptera litura</i>	—	Testis development and the formation and function of external genitalia	[180]
		<i>Tribolium castaneum</i> <i>Onthophagus taurus</i>	Multiple potential Dsx targets	Sexual dimorphism	[174]
		<i>Tribolium castaneum</i>	Vitellogenins, Vitellogenin receptors	Reproduction	[181]

		<i>Trypoxylus dichotomus</i>	—	Sexual dimorphism	[182,183]
Dys	B	<i>Drosophila melanogaster</i>	—	Tracheal migration; dhesion and fusion; leg development	[184–187]
E2F4	A	<i>Antheraea pernyi</i>	—	Immune response	[188]
E74	C	<i>Bombyx mori</i>	Atg genes	Autophagy in the fat body	[189]
E74A	C	<i>Bombyx mori</i>	<i>EO</i>	Ecdysone degradation	[190]
		<i>Bradysia hygida</i>	<i>BhC4-1</i>	Ecdysone signaling	[191]
E74B	C	<i>Aedes aegypti</i>	<i>vitellogenin</i>	Vitellogenesis	[192]
E75	C	<i>Bombyx mori</i>	E75A, B and C repress <i>HR3</i> ; E75A and E75C directly induce Halloween genes; E75B down-regulates Halloween genes	Ecdysteroidogenesis	[193]
		<i>Aedes aegypti</i>	<i>vitellogenin</i>	Vitellogenesis	[53–56]
		<i>Thermobia domestica</i>	—	Circadian clock; ecdysis	[194]
		<i>Drosophila melanogaster</i>	<i>period</i>	Circadian clock	[195]
E75A	C	<i>Drosophila melanogaster</i>	<i>Eip63F-1</i>	Salivary gland development	[196]
		<i>Manduca sexta</i>	<i>MHR3</i>	Ecdysone signaling	[197]
E75B	C	<i>Manduca sexta</i>	<i>ddc</i>	Molting	[198]
		<i>Plodia interpunctella</i>	—	Differentiation of imaginal wing cells	[199]
E93	C	<i>Bombyx mori</i>	—	Larval-pupal metamorphosis	[200]
E93, Br-C, Kr-h1	C	<i>Xenos vesparum</i>	—	Female neoteny	[201]
EcR	C	<i>Bombyx mori</i>	—	Programmed cell death in the anterior silk glands; silk protein synthesis	[202,203]

		<i>Bicyclus anynana</i>	—	Variation in wing eyespot patterns	[204]
		<i>Drosophila melanogaster</i>	<i>Kr-h1</i>	Photoreceptor maturation	[205]
EcR, USP	C	<i>Drosophila melanogaster</i>	—	20E signaling pathway	[206]
		<i>Schistocerca gregaria</i>	—	Metamorphosis and development	[207]
EcR-B1	C	<i>Plodia interpunctella</i>	—	Differentiation of imaginal wing cells	[199]
Emc	B	<i>Drosophila melanogaster</i>	<i>string</i>	Cell cycle and dorsal-ventral patterning of the imaginal discs	[134,208,209]
		<i>Bombyx mori</i>	—	Blastokinesis	[210]
Ems	A	<i>Drosophila melanogaster</i>	—	Embryonic and olfactory sense organ development	[211]
En, Nej	A	<i>Drosophila melanogaster</i>	<i>dpp</i>	Ovarian stem cell niche	[212]
Erm	C	<i>Drosophila melanogaster</i>	—	Nervous system development	[160,213,214]
ERR, EcR, Usp	C	<i>Bombyx mori</i>	<i>vitellogenin</i>	Reproduction	[215]
ERR, EcR	C	<i>Teleogryllus emma</i>	—	Maintenance of the function of adult cricket testis	[216]
Ets	A	<i>Bombyx mori</i>	<i>lebocin</i>	Immune response	[71]
Eve	A	<i>Bombyx mori</i>	—	Embryo segmentation	[217]
Eya	D	<i>Drosophila melanogaster</i>	—	Neuronal specification	[135]
Fer1	B	<i>Drosophila melanogaster</i>	odorant receptor	Odor identification	[218]
Fer2	B	<i>Drosophila melanogaster</i>	—	Nervous system development; locomotor rhythms regulation	[219]
Fkh	A	<i>Drosophila melanogaster</i>	<i>Dipt, Mtk</i>	Immune response	[220]
			autoregulation, <i>CrebA, Sage</i>	Salivary gland development	[123,221–223]
			<i>reaper, hid, sens</i>	Maintenance of SG cells survival	[196,224,225]

			<i>PH4αSG2, PH4αSG1</i>	Maintenance of a uniform and patent SG lumen	[222]
			<i>Sgs4</i>	Larval SG development	[226,227]
			<i>Eip63F-1</i>	Ca ²⁺ -signaling and secretion in SG and Malpighian tubes	[228]
			<i>CG6770, cabut</i>	Growth inhibition	[229]
			SPCGs	Secretory activity in salivary glands and epidermis	[123]
		<i>Bombyx mori</i>	<i>fibH</i>	Silk protein synthesis	[230]
			<i>fibL, P25, sericin-1</i>	Silk protein synthesis	[231–233]
		<i>Manduca sexta</i>	<i>moricin, lysozyme, defensin-1, defensin-3, attacin-2</i>	Immune response	[234]
FoxA	A	<i>Helicoverpa armigera</i>	<i>DH-PBAN</i>	Diapause	[235]
		<i>Helicoverpa armigera</i> <i>Spodoptera litura</i>	<i>ABCC2, ABCC3</i>	Cry1Ac toxin resistance	[236]
FoxL1	A	<i>Drosophila melanogaster</i>	<i>Sema2a</i>	SG positioning in embryos	[237]
FoxL2	A	<i>Nilaparvata lugens</i>	<i>Fcp3C</i>	Choriogenesis	[238]
FOXO	A	<i>Drosophila melanogaster</i>	<i>lipase 4, brummer</i>	Lipid metabolism	[239,240]
			<i>Drs</i>	Immune response	[241]
			—	Ecdysone biosynthesis	[242]
		<i>Bombyx mori</i>	<i>brummer, acid lipase-1</i>	Lipolysis	[243]
			<i>JHE, JHEH, JHDK</i>	JH degradation	[244]
		<i>Glossina morsitans</i>	—	Lipolysis suppression	[245]
βFtz-F1	C	<i>Blattella germanica</i>	—	JH biosynthesis	[246]
		<i>Drosophila melanogaster</i>	—	JH signaling	[247]

			<i>phm, dib, sad</i>	Ecdysteroidogenesis	[248,249]
			<i>EDG84A, EDG78E</i>	Pupal cuticle protein synthesis	[250–252]
			<i>MMP2</i>	Fat body remodeling	[253]
		<i>Bombyx mori</i>	<i>BmWCP5, BmWCP4, CPR55</i>	Cuticle protein synthesis	[66,67,70,254]
			<i>BmACP-6.7</i>	Formation of adult epidermis	[255]
			Atg genes	Autophagy in the fat body	[189]
			<i>fibH</i>	Silk protein synthesis	[256]
		<i>Aedes aegypti</i>	<i>vitellogenin</i>	Vitellogenesis	[257]
FTZ	A	<i>Drosophila melanogaster</i>	<i>slp1, wg, en</i>	Embryonic axis establishment	[258,259]
GATA β	C	<i>Bombyx mori</i>	late chorion genes	Choriogenesis	[36,260,261]
GATA-1	C	<i>Manduca sexta</i>	<i>moricin</i>	Immune response	[262]
Gce	B	<i>Drosophila melanogaster</i>	—	A cofactor of Met	[263]
Gt	B	<i>Drosophila melanogaster</i>	<i>kr, hb, eve</i>	Embryonic development	[264]
Hand	B	<i>Drosophila melanogaster</i>	—	Cardiogenesis; hematopoiesis; wing heart and muscle formation	[265–268]
H, Gro	B	<i>Aedes aegypti</i>	15% of Met-repressed genes	Reproduction	[269]
Hb	A	<i>Bombyx mori</i>	—	Embryo development	[270]
			—	NB cell fate decision	[271]
		<i>Drosophila melanogaster</i>	<i>rhomboid, Star, CBP, Fasciclin2, sprout</i>	Retinal glia cell development and blood-brain barrier integrity	[272]
HNF4	C	<i>Aedes aegypti</i>	—	Lipid metabolism	[273]
bHLH54F	B	<i>Drosophila melanogaster</i>	—	Expressed in the precursor of longitudinal visceral muscle	[274]
Hinge 1/2/3	D	<i>Drosophila melanogaster</i>	—	Wing hinge formation	[275]

DHR3	C	<i>Drosophila melanogaster</i>	<i>EDG84A</i>	Pupal cuticle protein synthesis	[250,252]
			<i>EcR, E74B, βFTZ-1</i>	Metamorphosis	[276]
			<i>phm, dib, sad</i>	Ecdysteroidogenesis	[249]
HR3	C	<i>Aedes aegypti</i>	<i>SCP-2</i>	Energy metabolism	[257]
			<i>vitellogenin</i>	Vitellogenesis	[53–56]
		<i>Bombyx mori</i>	Atg genes	Autophagy in the fat body	[189]
		<i>Manduca sexta</i>	<i>MHR3</i>	Ecdysone signaling	[197]
		<i>Plodia interpunctella</i>	—	Differentiation of imaginal wing cells	[199]
<i>Thermobia domestica</i>	—	Circadian clock; ecdysis	[194]		
DHR4	C	<i>Drosophila melanogaster</i>	<i>Cyp6t3</i>	Repress ecdysteroidogenesis	[277]
HR4	C	<i>Manduca sexta</i>	<i>ddc</i>	Molting	[198]
HR38	C	<i>Bombyx mori</i>	—	The effect of PTH on HR38 expression	[278]
DHR39	C	<i>Drosophila melanogaster</i>	<i>EDG84A</i>	Pupal cuticle protein synthesis	[250,252]
DHR96	C	<i>Drosophila melanogaster</i>	Many PB-regulated genes	PB tolerance and DDT resistance	[279]
HSF	A	<i>Drosophila melanogaster</i>	Heat shock genes	Heat stress	[280,281]
		<i>Bombyx mori</i>	—	Activation of <i>Hsp70a</i> and <i>Samui</i> under 5 °C -incubation	[282]
Hth	A	<i>Drosophila melanogaster</i>	—	Development of the wing imaginal disc	[283,284]
ILF	C	<i>Bombyx mori</i>	<i>POUM2</i>	Metamorphosis	[285]
Keap1	D	<i>Bombyx mori</i>	—	Response after phoxim and phoxim treatment	[104,105]
Kni	C	<i>Drosophila melanogaster</i>	<i>Phm, Dib</i>	Ecdysteroidogenesis	[286]
Kr	C	<i>Drosophila melanogaster</i>	<i>Rh5, Rh6</i>	Photoreceptor neuron differentiation	[287]
Kr-h1	C	<i>Bombyx mori</i>	<i>Br-C, E93</i>	Metamorphosis	[288,289]
Kr-h1	C	<i>Drosophila melanogaster</i>	Steroidogenic enzymes	Ecdysone biosynthesis; metabolic homeostasis; metamorphosis	[240,290,291]

		<i>Locusta migratoria</i>	<i>vitellogenin</i>	Vitellogenesis and oocyte maturation	[292]
		<i>Nilaparvata lugens</i>	—	Ovary development	[59]
		<i>Tribolium castaneum</i>	E93	Metamorphosis	[290,293]
		<i>Blattella germanica</i>			
		<i>Pyrrhocoris apterus</i>	<i>vitellogenin</i>	Vitellogenesis	[294]
LL3	D	<i>Anopheles gambiae</i>	SPRN6	Anti-Plasmodium immunity	[295]
<i>l'sc</i>	B	<i>Drosophila melanogaster</i>	—	Muscle progenitors	[296]
Lolal	D	<i>Drosophila melanogaster</i>	<i>crumbs, Moesin</i>	SG and trache elongation	[297]
Islet, Lim3	A	<i>Drosophila melanogaster</i>	—	Motoneuron identity	[298]
Max	B	<i>Drosophila melanogaster</i>	—	Regulation of Myc activity	[299]
MBF2	D	<i>Bombyx mori</i>	<i>fibH</i>	Silk protein synthesis	[149]
Met	B	<i>Bombyx mori</i>	<i>Kr-h1</i>	Metamorphosis	[300,301]
		<i>Cimex lectularius</i>	<i>vitellogenin</i>	Vitellogenesis and ovigenesis	[302]
		<i>Drosophila melanogaster</i>	<i>Dronc, Drice</i>	Programmed cell death	[303]
		<i>Glossina morsitans</i>	—	Lipolysis suppression	[245]
		<i>Leptinotarsa decemlineata</i>	—	Growth and development	[304,305]
		<i>Locusta migratoria</i>	<i>vitellogenin</i>	Vitellogenesis and oocyte maturation	[292]
			<i>Mcm4, Mcm7</i>	Polyploidy and vitellogenesis	[306]
			<i>Cdc6, Grp78-2</i>	Fat body cell homeostasis, polyploidy, vitellogenesis and oogenesis	[307,308]
		<i>Pyrrhocoris apterus</i>	<i>vitellogenin</i>	Vitellogenesis	[294]
			Accessory gland proteins and hexamerins	Male reproduction	[309]
			<i>Pdp1^{iso1}, cry2</i>	Adult diapause regulation	[131]
		<i>Tribolium castaneum</i>	<i>Kr-h1, Br-C</i>	Metamorphosis	[60,290,310–314]

Met, SRC	B	<i>Aedes aegypti</i>	<i>Kr-h1;RRS1</i> ; six ribosomal protein; <i>ET</i>	Reproduction	[315–318]
Met, CYC, SRC	B	<i>Aedes aegypti</i>	<i>Kr-h1, Hairy</i>	Light-dependent circadian regulation	[319]
MIC	D	<i>Bombyx mori</i>	<i>sericin-1</i>	Silk protein synthesis	[320–322]
Mid	D	<i>Drosophila melanogaster</i>	<i>wingless, hedgehog</i>	Wing disc development	[323]
			<i>gooseberry</i>	Neuroblast development	[324]
Mitf	B	<i>Drosophila melanogaster</i>	—	Eye development	[325]
			V-ATPase	Lysosomal-autophagy pathway	[326]
Mld	C	<i>Drosophila melanogaster</i>	<i>spok</i>	Ecdysteroidogenesis	[286,327–329]
Mlx, Mondo	B	<i>Drosophila melanogaster</i>	<i>Cabut, Aldh-III</i>	Dietary sugar tolerance	[330]
Mnt	B	<i>Drosophila melanogaster</i>	—	Body size regulation	[331]
dMyd	B	<i>Drosophila melanogaster</i>	—	A transiently expressed muscle marker	[332]
Nej, Tgo	B	<i>Drosophila melanogaster</i>	—	Peripheral glial differentiation	[333]
Nkx-2.5-like, transcription factor 3-like	A	<i>Apis mellifera</i>	Unknown	Fipronil resistance	[334]
Odd	C	<i>Drosophila melanogaster</i>	—	Embryonic axis establishment	[335]
		<i>Bombyx mori</i>	—	Embryo segmentation	[217]
Oli	B	<i>Drosophila melanogaster</i>	—	Nervous system development; larval and adult locomotion; neuronal remodeling	[336,337]
Otd	A	<i>Drosophila melanogaster</i>	<i>kr-h1</i>	Photoreceptor maturation	[205]
		<i>Bombyx mori</i>	<i>eve</i>	Embryo body segmentation	[80]
		<i>Onthophagus binodis</i> <i>Digitonthophagus gazella</i> <i>Liatongus militaris</i>	—	Development of functional ectopic compound eyes	[338]

Pb	A	<i>Drosophila melanogaster</i>	—	Embryonic axis establishment	[339]
Pdm1, Pdm2	A	<i>Drosophila melanogaster</i>	<i>CecA1</i>	Immune response	[340]
Pdp1	B	<i>Drosophila melanogaster</i>	—	Regulation of somatic muscle gene expression	[341]
Pdp1ε	B	<i>Drosophila melanogaster</i>	—	Circadian rhythm	[130]
Pdp1γ	B	<i>Drosophila melanogaster</i>	—	Pphysiological and drug-influenced lipid regulation	[342]
Pitx	A	<i>Bombyx mori</i>	<i>DH-PBAN, PTTH</i>	Neuroendocrine system activation	[343,344]
PntP1	A	<i>Drosophila melanogaster</i>	<i>Ase, erm, Pros</i>	Nervous system development	[345,346]
POU-M1	A	<i>Bombyx mori</i>	<i>sericin-1</i>	Silk protein synthesis	[321,347]
Prd	A	<i>Drosophila melanogaster</i>	<i>EcR</i>	The function of male accessory glands and fertility	[348]
Prey2	D	<i>Helicoverpa armigera</i>	<i>CYP6B6</i>	Response to plant secondary toxicant 2-tridecanone	[349]
Vri	B	<i>Drosophila melanogaster</i>	—	A dominant enhancer of Dpp signaling in embryonic D-V patterning and wings; cardiac aging; hair and cell growth; embryonic tracheal development; circadian clock	[350–353]
		<i>Spodoptera frugiperda</i>	—	Circadian rhythm regulation	[354]
Vvl	A	<i>Drosophila melanogaster</i>	<i>CecA1, DiptA, AttA, AttB, Drs</i>	Immune response	[79,340]
			<i>Phm, Dib</i>	Ecdysteroidogenesis	[286]
			—	Motoneuron identity	[298]
		<i>Bombyx mori</i>	<i>vitellogenin</i>	Reproduction	[64]
			<i>fibH</i>	Silk synthesis	[355]
<i>Phm</i>	Ecdysteroidogenesis	[356]			

			<i>DH-PBAN</i>	Diapause	[357]
			<i>BmWCP4</i>	Cuticle protein synthesis	[1,2]
		<i>Tribolium castaneum</i>	<i>jhamt3, HR3, Phm, Spo</i>	JH biosynthesis and ecdysteroidogenesis	[358]
		<i>Helicoverpa armigera</i>	<i>DH-PBAN, PTEN</i>	Diapause	[359,360]
Rel	D	<i>Drosophila melanogaster</i>	—	Immune response	[141,361,362]
		<i>Helicoverpa armigera</i>	<i>cathepsin L</i>	Fat body dissociation	[363]
Rel2	D	<i>Manduca sexta</i>	AMPs	Immune response	[153]
REL2	D	<i>Anopheles gambiae</i>	AMPs	Immune response	[364]
REL1-A, REL1-B	D	<i>Aedes aegypti</i>	<i>Dipt, Drs,</i>	Antifungal immune responses	[365]
REL2	D	<i>Aedes aegypti</i>	<i>Defensins A, C, D Cec A, Cec N</i>	Defense against Gram-positive, Gram-negative bacteria and <i>Plasmodium gallinaceum</i>	[366–368]
Rel2	D	<i>Culex quinquefasciatus</i>	<i>Vago</i>	Anti-viral responses	[369]
RelA, RelB	D	<i>Bombyx mori</i>	<i>Lebocin4, Attacin</i>	Immune response	[370]
Rel1, Rel2	D	<i>Bombyx mori</i>	<i>Cecropin B1, Attacin, Lebocin 4</i>	Immune response	[371]
RelA	D	<i>Allomyrina dichotoma</i>	<i>Coleoptericin A</i>	Immune response	[372]
Rib	A	<i>Drosophila melanogaster</i>	<i>crumbs, Moesin,</i>	SG and trache elongation	[297]
			Autoregulation	Embryonic SG morphogenesis	[373]
Run	D	<i>Drosophila melanogaster</i>	<i>Slx</i>	Sex determination	[374]
			<i>engrailed, slp1</i>	Embryonic development	[258,259]
		<i>Bombyx mori</i>	—	Embryo segmentation	[217]
Sage	B	<i>Bombyx mori</i>	<i>fibH</i>	Silk protein synthesis	[148,230]
		<i>Drosophila melanogaster</i>	Sage target genes	Salivary gland development	[223]
			<i>PH4αSG2, PH4αSG1</i>	Maintenance of a uniform and patent SG	[222]

				lumen	
Sal	C	<i>Pieris rapae</i>	—	Wing pattern development	[375]
Sal, E75	C	<i>Papilio Xuthus</i>	—	Larval eyespot markings	[376]
Sc (Sis-B)	B	<i>Drosophila melanogaster</i>	<i>Sxl</i>	Sex determination	[377]
Scr	C	<i>Drosophila melanogaster</i>	<i>Fkh, CrebA, Sage, Hkb</i>	Salivary gland specification	[378–381]
		<i>Bombyx mori</i>	—	Development of the embryonic silk gland	[382]
Séan, Ouib, Mld	C	<i>Drosophila melanogaster</i>	<i>nvd, spok</i>	Ecdysteroidogenesis	[329]
Sens	C	<i>Drosophila melanogaster</i>	<i>reaper, hid</i>	Maintenance of salivary gland cells survival	[225]
SGF 1/2/3/4	D	<i>Bombyx mori</i>	—	Silk protein synthesis	[231,383]
Sim, Tgo	B	<i>Drosophila melanogaster</i>	—	CNS midline cell development	[384,385]
Sima, Tgo	B	<i>Drosophila melanogaster</i>	—	Normal development and hypoxia response	[386,387]
Sis-A	B	<i>Drosophila melanogaster</i>	<i>Sxl</i>	Sex determination	[388]
			—	Midgut formation; Brahma complex recruitment	[389] [390]
slp1	A	<i>Drosophila melanogaster</i>	—	Embryonic axis establishment	[259]
Sna, Esg, Wor	C	<i>Drosophila melanogaster</i>	—	Asymmetric cell division of embryonic NBs	[391–393]
Nine Sox genes	D	<i>Bombyx mori</i>	—	Identification and characterization of Sox genes	[394]
SRAM	D	<i>Sarcophaga peregrina</i>	<i>lectin</i>	Immune response	[395]
dSREBP	B	<i>Drosophila melanogaster</i>	—	Lipid synthesis	[396]

			—	Nervous system development	[397]
SRC	B	<i>Cimex lectularius</i>	—	Reproduction	[398]
		<i>Aedes aegypti</i>	—	Reproduction	[315,316,318]
		<i>Locusta migratoria</i>	—	Reproduction	[292,306–308]
		<i>Pyrrhocoris apterus</i>	—	Reproduction	[294,309]
		<i>Bombyx mori</i>	—	Metamorphosis	[300,301]
		<i>Tribolium castaneum</i>	—	Metamorphosis	[313,314]
Ss	B	<i>Drosophila melanogaster</i>	—	Nervous system development	[186]
			—	Distal antenna formation	[399]
STAT	D	<i>Drosophila melanogaster</i>	<i>Raf</i>	Immune response	[400]
STAT-A	D	<i>Anopheles gambiae</i>	<i>NOS, SOCS</i>	Immune response	[401]
STAT-B	D	<i>Anopheles gambiae</i>	<i>STAT-A</i>	Immune response	[401]
Svp	C	<i>Blattella germanica</i>	—	JH biosynthesis	[246]
		<i>Drosophila melanogaster</i>	—	CNS and photoreceptor cell development	[402–404]
Tgo	B	<i>Drosophila melanogaster</i>	—	Formation of CNS midline antennal and tarsal development peripheral glial cell migration	[405–408]
Tim	B	<i>Laodelphax striatellus</i>	—	The circadian behavioral rhythms	[409]
Tll	C	<i>Drosophila melanogaster</i>	—	Neural and embryonic development	[206,410,411]
Toy	A	<i>Drosophila melanogaster</i>	<i>ey</i>	Eye development	[412]
Trh, Tgo	C	<i>Drosophila melanogaster</i>	<i>jing</i>	Tracheal development	[407]
Twi	B	<i>Drosophila melanogaster</i>	—	Cell fate decisions	[413]
Tsh	C	<i>Drosophila melanogaster</i>	—	Wing imaginal disc development	[283,284]
Ubx	A	<i>Bicyclus anynana</i>	—	Embryonic body plan and wing patterns	[17,18]
		<i>Drosophila melanogaster</i>	<i>dad</i>	The differentiation of the gastric stem cell lineage	[414]

		<i>Manduca sexta</i>	—	The expression patterns in the embryos	[5]
		<i>Bombyx mori</i>	—	The abdominal appendage specification	[415]
USF	B	<i>Drosophila melanogaster</i>	<i>Stellate</i>	Male fertility	[416]
Woc	C	<i>Drosophila melanogaster</i>	—	Ecdysone biosynthesis	[417]
Zld	C	<i>Drosophila melanogaster</i>	<i>Sxl</i>	Sex determination	[418]
			<i>eve</i>	Embryonic development	[419]
Zfh1, Zfh2	C	<i>Drosophila melanogaster</i>	—	Embryonic development and immune regulation	[420,421]

^a Abbreviations of TFs in this table. **Abd-A/B**: Abdominal-A/B; **AhR**: Aryl hydrocarbon receptor; **ARNT**: Aryl hydrocarbon receptor nuclear translocator; **Antp**: Antennapedia; **Apt-like**: Apontic-like; **Ase**: Asense; **ATF-2/3**: Activating transcription factor 2/3; **Ato**: Atonal; **Bcd**: Bicoid; **Bmx**: Bigmax; **BrC**: Broad-Complex; **BrC-Z1/2/3/4**: Broad complex isoform 1/2/3/4; **Btd**: Buttonhead; **Cad**: Caudal; **Cato**: Cousin of atonal; **C/EBP**: CCAAT-enhancer-binding protein; **Cnc B/C**: Cap 'n' collar B/C; **Col**: Collier; **Crc**: Cryptocephal; **CREB**: cAMP response element binding protein; **Cyc**: Cycle; **Clk**: Clock; **Cwo**: Clockwork orange; **Da**: Daughterless; **Dac**: Dachshund; **Dei**: Delilah; **Deaf-1**: Deformed epidermal autoregulatory factor-1; **Dfd**: Deformed; **Dif**: Dorsal-related immunity factor; **Dimm**: Dimmed; **Dip3**: Dorsal interacting protein 3; **DI**: Dorsal; **Dm**: Diminutive; **Dpn**: Deadpan; **Dys**: Dysfusion; **E74/75**: Ecdysone-induced protein 74/75; **EcR**: Ecdysone receptor; **Emc**: Extramacrochaetae; **Ems**: Empty spiracles; **En**: Engrailed; **ERR**: Oestrogen-related receptor; **Esg**: Escargot; **Eve**: Even-skipped; **Eya**: Eyes absent; **Fkh**: Forkhead; **FoxA**: Forkhead box, sub-group A; **FoxL1/2**: Forkhead box transcription factor L1/2; **FOXO**: Forkhead box, sub-group O; **βFtz-F1**: β-fushi tarazu transcription factor 1; **Ftz**: Fushi tarazu; **Gce**: Germ-cell expressed; **Gro**: Groucho; **Gt**: Giant; **H**: Hairly; **Hb**: Hunchback; **HNF4**: Hepatocyte nuclear factor 4; **DHR3/4/39/96**: *Drosophila* Hormone Receptor 3/4/39/96; **HR3/4/38**: Hormone Receptor 3/4/38; **HSF**: Heat shock factor; **Hth**: Homothorax; **Kni**: Knirps; **Kr**: Krüppel; **Kr-h1**: Krüppel homolog 1; **Lab**: Labial; **LL3**: LITAF-like 3; **I^{sc}**: lethal of scute; **Lolal**: Lola Like; **MBF2**: Multiprotein bridging factor 2; **Met**: Methoprene-tolerant; **Nej**: Nejire; **Odd**: Odd-skipped; **Oli**: Olig family; **Otd**: Orthodenticle; **Pdp1**: PAR-domain protein 1; **PntP1**: Pointed P1; **Prd**: Paired; **Vri**: Vrille; **USF**: Upstream stimulatory factor; **Vvl**: Ventral veins lacking; **Rel**: Relish; **Pb**: Proboscidea; **Rib**: Ribbon; **Run**: Runt; **Sal**: Spalt; **Sc**: Scute; **Scr**: Sex combs reduced; **Séan**: Séance; **Sens**: Senseless; **Sim**: Single-minded; **Sima**: Similar; **Sis-A/B**: Sisterless A/B; **Slp1**: Sloppy-paired 1; **Sna**: Snail; **SRAM**: Sarcophaga-derived Rel/Ankyrin molecule; **SRC**: Steroid receptor coactivator; **Ss**: Spineless; **STAT**: Signal transducers and activators of transcription; **Svp**: Seven-up; **Tgo**: Tango; **Tim**: Timeless; **Tll**: Tailless; **Toy**: Twin of eyeless; **Trh**: Tracheless; **Tw**: Twist; **Tsh**: Teashirt; **Ubx**: Ultrabithorax; **Wor**: Worniu; **Woc**: Without children; **Zld**: Zelda; **Zfh1**: Zn finger homeodomain 1.

^b The structures of TF superclasses: A represents the helix-turn-helix domain factor; B represents the basic domain factor; C represents the zinc finger domain factor; D represents other DNA-binding domain factor.

^c We apologize in advance to those whose work could not be adequately discussed and cited in the main body due to space limitations.

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