Supplementary figures.



Supplementary 1. Figure KDM3A promotes cardiomyocyte hypertrophy. (a) Immunofluorescence staining of Flag-tagged KDM3A in NRVMs transduced with adenovirus expressing Flag-Kdm3a using anti-Flag antibody. Scale bar, 20 µM. (b) WB of H3K9me2 of NRVMS transduced with adenoviruses expressing LacZ (Ad-LacZ) or Kdm3a (Ad-Kdm3a). Gapdh was used as loading control. Overexpression of KDM3A in NRVMs resulted in downregulation of H3K9me2. (c) Relative mRNA of genes indicated from NRMVs transduced with either Ad-LacZ or Ad-Kdm3a. Overexpression of Kdm3a upregulated canonical genes associated with cardiomyocyte hypertrophy and Timp1. n=3±SEM, \*, p<0.05 (t test), relative to LacZ-transduced cells. (d-f) NRVMs were transfected with control or Kdm3a specific siRNA and stimulated with or without PE. Kdm3a knockdown (KD) resulted in approximately 50% downregulation of Kdm3a mRNA relative to control siRNA transfected cells (d). Kdm3a KD also resulted in upregulation of H3K9me2 (e). (f) Relative cell size of NRVMs in Kdm3a siRNA or control siRNA transfected cells and stimulated with or without PE. Kdm3a KD attenuated PEstimulated myocyte hypertrophy \*, p < 0.05 (ANOVA). Scale bar, 100  $\mu$ m.



**Supplementary Figure 2. KDM3A promotes TAC-induced hypertrophic remodeling.** (a) Schematics of *Kdm3a* transgenic construct. (b) Western blot of expression levels of Flag-Kdm3a and H3K9me2 in the heart. GADPH was used as loading control. (c) H&E (top two panels) and trichrome (lower two panels) staining of histological sections of WT and *Kdm3a*-Tg (line 29) mouse hearts at week 6 post-sham and TAC surgery. Scale bars, 1mm (first panel), 20  $\mu$ m (second panel), 100  $\mu$ m (3<sup>rd</sup> and 4<sup>th</sup> panels). (d) HW/BW, LW/BW, FS, and Heart rate of *Kdm3a*-Tg (line 24) mouse hearts at week 6 post-sham and TAC surgery. n=5-9±SEM. \*, *p<0.05* (ANOVA).



Supplementary Figure 3. *Kdm3a*-deficiency protects mice against hypertrophic remodeling in response to TAC injury. (a) H&E (top two panels) and trichrome (lower two panels) staining of histological sections of WT and *Kdm3a* KO mouse (in FVB/C57/BI6 mixed background) hearts at week 6 post-sham and TAC surgery. Scale bars, 1mm (first panel), 20  $\mu$ m (second panel), 100  $\mu$ m (3<sup>rd</sup> and 4<sup>th</sup> panels). (b) HW/BW, LW/BW, ejection fraction (EF) and FS of *Kdm3a* KO mouse (in C57/BI6 background) hearts at week 6 post-sham and TAC surgery. n=3-4±SEM. \*, *p*<0.05. #, *p*<0.01 (ANOVA).



**Supplementary Figure 4.** (a) GO analysis of upregulated genes in *Kdm3a* KO TAC mouse hearts vs WT TAC littermates. (b) Relative fold change of mRNA of collagens in WT and Kdm3a-Tg mice at week 6 post-sham and TAC surgery. n=4±SEM. \*, relative to sham, p<0.05. #, relative to WT TAC, p<0.05 \_(ANOVA). (c) Adam9 and Adam19 in WT and TG mouse heart at week 6 post-sham and TAC surgery. n=4±SEM, \*,p<0.05 (ANOVA). (d) Western blot of indicated proteins in WT and Kdm3a-Tg mouse hearts at week 6 post-sham and TAC surgery.



**Supplementary Figure 5.** (a) Relative fold change of transcripts of genes involved in fibrosis in cardiomyocytes (CM) and fibroblasts (cFb, non-cardiomyocytes) fractions of WT and Kdm3a-Tg mouse hearts at week 6 post-sham and TAC surgery n=3 $\pm$ SEM, \*,*p*<0.05 (ANOVA). (b) Western blot of TIMP1, showing upregulation of TIMP1 in *Kdm3a*-Tg mouse hearts and in response to TAC surgery. (c) Western blot of TIMP1 in CM and cFb isolated from WT and *Kdm3a*-Tg mouse at week 6 post-Sham and TAC surgery. (d) *Kdm3a*-Tg mouse hearts at weeks 6 post-Sham and TAC surgery were used for ChIP assay with antibodies against KDM3A (left panel) or H3K9me2 (right panel). The relative occupancies of KDM3A and H3K9me2 at Timp1 promoter were normalized against Input and expressed relative to Sham control. n=3 $\pm$ SEM, \*,*p*<0.05 (t test). (e) Relative fold change of Timp1 mRNA in Sham and TAC *Kdm3a*-Tg mouse hearts treated with control or Timp1 shRNA n=3 $\pm$ SEM, \*,*p*<0.05 (ANOVA). (f) Relative mRNA of hypertrophic and fibrotic genes in Sham and TAC *Kdm3a*-Tg mouse hearts treated with control shRNA TAC vs Sham, \* Timp1 shRNA TAC vs control shRNA TAC. \*, \*,*p*<0.05 (ANOVA).



**Supplementary Figure 6.** (a) NRVMs were stimulated with PBS or PE, treated with JIB-04 or vehicle DMSO. JIB-04 suppressed PE-stimulated myocyte hypertrophy. n=3±SEM, \*, *p*<0.05 (ANOVA). (b) NRVMs transduced with either Ad-LacZ or Ad-Kdm3a were treated with JIB-04 or vehicle DMSO. JIB-04 suppressed Kdm3a-promoted myocyte hypertrophy n=3±SEM, \*, *p*<0.05 (ANOVA). Scale bar, 100 µm. (c) Relative mRNA of fetal genes and genes involved in fibrosis in *Kdm3a*-Tg sham and TAC mouse hearts treated with vehicle or JIB-04 starting at post-TAC week 3 for 4 weeks. mRNA transcripts were measured by qRT-PCR, normalized against internal Gapdh, and expressed relative to Sham mice. n=3±SEM. \*, *p*<0.05, \*TAC vs Sham, <sup>#</sup>JIB-04 vs vehicle n=3±SEM, \*, *p*<0.05 (ANOVA). (d-g) Kdm3a-Tg and WT littermates were subjected to TAC surgery and treated with Lox inhibitor β-aminoproprionitrile (BAPN) (8 mg/ml in drinking water). Echocardiograph was performed at week 6 post-TAC and hearts were harvested. HW/BW (c), LW/BW (d), FS (e), and relative fibrotic area (f) were measured. n=5-9±SEM, \*, *p*<0.05 (ANOVA).

**Supplementary Figure 7** Full, un-cropped Western blots in main article figures and supplementary figures. Boxes indicate regions cropped in the main article.





Fig. 8b





# Fig. 8c







JIB-04(uM)	_	0	0.4	0.8	
	1		_		
					- 50
GAPDH		-	-	-	
					25
					20
					-

## Supplementary Figure 1b



#### Supplementary Figure 1e



### Supplementary Figure 2b



## Supplementary Figure 4d



## Supplementary Figure 5b



Supplementary Figure 5c





Supplementary table 1: primer sequences

human GAPDH qPCR p1: AGCCACATCGCTCAGACAC   p2: GCCCAATACGACCAAATCC   human KDM3A qPCR p1: CCAGCCTCAAAGGAAGACCT   p2: ACTGCACCAAGAGTCGGTTT   mouse Gandh qPCR p1: GGCACAGTCAAGGCTGAGAATG	
human GAP Dirigron p1: AGCCACATCGCTCAGACAC   p2: GCCCAATACGACCAAATCC   human KDM3A qPCR   p1: CCAGCCTCAAAGGAAGACCT   p2: ACTGCACCAAGAGTCGGTTT   mouse Gandh qPCR   p1: GCCACACCAAGAGTCGAGAATG	
human KDM3A qPCR p1: CCAGCCTCAAAGGAAGACCT p2: ACTGCACCAAGAGTCGGTTT p2: ACTGCACCAAGAGTCGGTTT	
p1: CCAGCCTCAAAGGAAGACCT p2: ACTGCACCAAGAGTCGGTTT mouse Gandh gPCR p1: GGCACAGTCAAGGCTGAGAATG	
mouse Nppa qPCR p1: CAACACAGATCTGATGGATTTCA	
mouse Nppb qPCR p1: GTCAGTCGTTTGGGGCTGTAAC	
p2: AGACCCAGGCAGAGTCAGAA	
mouse Myh7 qPCR p1: CGCATCAAGGAGCTCACC	
p2: CTGCAGCCGCAGTAGGTT	
mouse FhI1 qPCR p1: GGCTTCTCAAAGACACTCAGG	
p2: TCGAACTTCTCCGACATGGT	
mouse Tgfβ1 qPCR p1: CTCCCGTGGCTTCTAGTGC	
p2: GCCTTAGTTTGGACAGGATCTG	
mouse Tgfβ2 qPCR p1: TCGACATGGATCAGTTTATGCG	
p2: CCCTGGTACTGTTGTAGATGGA	
mouse Tgfβ3 qPCR p1: CCTGGCCCTGCTGAACTTG	
p2: TTGATGTGGCCGAAGTCCAAC	
mouse Timp1 qPCR p1: AGCCTGGAGGCAGTGATTTC	
p2: GGGCCATCATGGTATCTCTGG	
mouse LoxL1qPCR p1: GAGTGCTATTGCGCTTCCC	
p2: GGTTGCCGAAGTCACAGGT	
mouse LoxL2 qPCR p1: ATTAACCCCAACTATGAAGTGCC	
p2: CTGTCTCCTCACTGAAGGCTC	
mouse Nupr1 qPCR p1: CCCTTCCCAGCAACCTCTAAA	
p2: TCTTGGTCCGACCTTTCCGA	
mouse Fibulin2 qPCR p1: CTGTGAAGACCAAGACGAGTG	
p2: CGTTGAGGATATAGCCCTCTGC	
mouse Sparc qPCR p1: GTGGAAATGGGAGAATTTGAGGA	
p2: CTCACACACCTTGCCATGTTT	
mouse Postn qPCR p1: CGGGAAGAACGAATCATTACA	
p2: ACCTTGGAGACCTCTTTTGC	
mouse Bmp10 gPCR p1: ATGGGGTCTCTGGTTCTG	
p2: CAATACCATCTTGCTCCGTGAA	
mouse Col1a1 gPCR p1: GCTCCTCTTAGGGGGCCACT	
p2: CCACGTCTCACCATTGGGG	
mouse Col1a2 gPCR p1: AGCCCTGGTTCTCGAGGT	
p2: CCGGTTGAACCACGATTG	
mouse Col3a1 gPCR p1: CTGTAACATGGAAACTGGGGAAA	
p2: CCATAGCTGAACTGAAACCACC mouse Col5a1 gPCR p1: CTTCGCCGCTACTCCTGTTC	
p2: CCATAGCTGAACTGAAAACCACC mouse Col5a1 qPCR p1: CTTCGCCGCTACTCCTGTTC p2: CCCTGAGGGCAAATTGTGAAAA	
p2: CCATAGCTGAACTGAAAACCACC   mouse Col5a1 qPCR p1: CTTCGCCGCTACTCCTGTTC   p2: CCCTGAGGGCAAATTGTGAAAA   mouse Col6a1 qPCR p1: AGGCCCTATTGGGCTTCAAG	
p2: CCATAGCTGAACTGAAAACCACC   mouse Col5a1 qPCR p1: CTTCGCCGCTACTCCTGTTC   p2: CCCTGAGGGCAAATTGTGAAAA   mouse Col6a1 qPCR p1: AGGCCCTATTGGGCTTCAAG   p2: GCCAGTGTATCCTCGCTCTC	
p2: CCATAGCTGAACTGAAAACCACC   mouse Col5a1 qPCR p1: CTTCGCCGCTACTCCTGTTC   p2: CCCTGAGGGCAAATTGTGAAAA   mouse Col6a1 qPCR p1: AGGCCCTATTGGGCTTCAAG   p2: GCCAGTGTATCCTCGCTCTC   mouse Col8a1 qPCR p1: ACTCTGTCAGACTCATTCAGGC	
p2: CCATAGCTGAACTGAAAACCACCmouse Col5a1 qPCRp1: CTTCGCCGCTACTCCTGTTCp2: CCCTGAGGGCAAATTGTGAAAAmouse Col6a1 qPCRp1: AGGCCCTATTGGGCTTCAAGp2: GCCAGTGTATCCTCGCTCTCmouse Col8a1 qPCRp1: ACTCTGTCAGACTCATTCAGGCp2: CAAAGGCATGTGAGGGGACTTG	
p2: CCATAGCTGAACTGAAAACCACCmouse Col5a1 qPCRp1: CTTCGCCGCTACTCCTGTTCp2: CCCTGAGGGCAAATTGTGAAAAmouse Col6a1 qPCRp1: AGGCCCTATTGGGCTTCAAGp2: GCCAGTGTATCCTCGCTCTCmouse Col8a1 qPCRp1: ACTCTGTCAGACTCATTCAGGCp2: CAAAGGCATGTGAGGGACTTGmouse Adam9 qPCRp1: GGAAGGCTCCCTACTCTCGA	

mouse Adam19 qPCR	p1: TCAGTGGCGGACTTCAGAAAG
	p2: GCAAAAAGGTGCTCGTTCTTC
rat Gapdh qPCR	p1: ATCACCATCTTCCAGGAGCGA
	p2: AGCCTTCTCCATGGTGGTGAA
rat Nppa qPCR	p1: CACAGATCTGATGGATTTCAAGA
	p2: CCTCATCTTCTACCGGCATC
rat Nppb qPCR	p1: GTCAGTCGCTTGGGCTGT
	p2: CAGAGCTGGGGAAAGAAGAG
rat FhI1 qPCR	p1: GGCTTCTCAAAGACACTCAGG
	p2: GTCGAACTTCTCAGACATGGTG
rat Myh6 qPCR	p1: TGCAGAAGAAACTGAAGGAAAA
	p2: GCTCCGCCTCTAGCTCCT
rat Timp1 qPCR	p1: CAGCAAAAGGCCTTCGTAAA
	p2: TGGCTGAACAGGGAAACACT
rat Kdm3a qPCR	p1: TTGCTCTGAGGTCTCTCCCAG
	p2: TGCTGTCTGTTGCTAGATGGG
Kdm3a-Tg mouse genotyping	p1: AGTGGTGGTGTAGGAAAGT
	p2: AACCACTGAGTAGATGGGTC
mouse Timp1 promoter	p1: AAAAAGCTAGCGCTGGCAGGAGGTTTTTGTG
	p2: AAAAACTCGAGAATCACTGCCTCCAGGCTTC
mouse Timp1 ChIP	p1: AGGAAGGACTGTGCATGACG
	p2: GGCCCCAGGATAAACCCAAA