Corresponding Author Name: Neil A R Gow

Manuscript Number: NCOMMS-16-15066

Reporting Checklist For Life Sciences Articles

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

Figure legends

☑ Check here to confirm that the following information is available in all relevant figure legends (or Methods section if too long):

- the **exact sample size (*n*)** for each experimental group/condition, given as a number, not a range;
- a **description of the sample collection** allowing the reader to understand whether the samples represent **technical or biological replicates** (including how many animals, litters, culture, etc.);
- a **statement of how many times the experiment shown was replicated in the laboratory**;
- **definitions of statistical methods and measures**: (For small sample sizes (n<5) descriptive statistics are not appropriate, instead plot individual data points)
  - o very common tests, such as *t*-test, simple χ^2 tests, Wilcoxon and Mann-Whitney tests, can be unambiguously identified by name only, but more complex techniques should be described in the methods section;
  - o are tests one-sided or two-sided?
  - o are there adjustments for multiple comparisons?
  - o **statistical test results**, e.g., ***P* values**;
  - o definition of '**center values**' as **median** or **mean**;
  - o definition of **error bars** as **s.d.** or **s.e.m.** or **c.i.**

This checklist will not be published. Please ensure that the answers to the following questions are reported in the manuscript itself. We encourage you to include a specific subsection in the Methods section for statistics, reagents and animal models. Below, provide the page number or section and paragraph number (e.g. "Page 5" or "Methods, 'reagents' subsection, paragraph 2").

Statistics and general methods	Reported in section/paragraph or page #:
1. How was the sample size chosen to ensure adequate power to detect a pre-specified effect size? (Give section/paragraph or page #)	Methods - Proof-of-concept in 3-day systemic candidiasis infection model
For animal studies, include a statement about sample size estimate even if no statistical methods were used.	To determine the minimum group size required to detect a statistical difference in outcomes (P<0.05), power analyses
2. Describe inclusion/exclusion criteria if samples or animals were excluded from the analysis. Were the criteria pre-established? (Give section/paragraph or page #)	No animals or samples were excluded from the original study. In the study conducted by Evotec, one mouse from the control group was excluded from analysis because it was
3. If a method of randomization was used to determine how samples/animals were allocated to experimental groups and processed, describe it. (Give section/paragraph or page #)	Methods - Proof-of-concept in 3-day systemic candidiasis infection model
For animal studies, include a statement about randomization even if no randomization was used.	Animals were randomly assigned into groups.
4. If the investigator was blinded to the group allocation during the experiment and/or when assessing the outcome, state the extent of blinding. (Give section/paragraph or page #)	Methods - Proof-of-concept in 3-day systemic candidiasis infection model
For animal studies, include a statement about blinding even if no blinding was done.	Investigators were not blinded during allocation or outcome assessment in this study
5. For every figure, are statistical tests justified as appropriate?	Methods - Live cell video microscopy phagocytosis assays and Analysis of disease progression
Do the data meet the assumptions of the tests (e.g., normal distribution)?	Yes
Is there an estimate of variation within each group of data?	Yes, where relevant.
Is the variance similar between the groups that are being statistically compared? (Give section/paragraph or page #)	Methods - Analysis of disease progression

(Continues on following page)

Reagents

Reported in section/paragraph or page #:

6. To show that antibodies were profiled for use in the system under study (assay and species), provide a citation, catalog number and/or clone number, supplementary information or reference to an antibody validation profile (e.g., Antibodypedia, 1DegreeBio).

Methods

7. Cell line identity:

a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by ICLAC (also available in NCBI Biosample)?

No

b. If yes, include in the Methods section a scientific justification of their use – indicate here on which page (or section and paragraph) the justification can be found.

Not applicable

c. For each cell line, include in the Methods section a statement that specifies:

- the source of the cell lines

J774.1 cell line from the European Collection of Authenticated Cell Cultures (ECACC) and Expi293F cell line from Gibco, USA.

- have the cell lines been authenticated? If so, by which method?

The cell lines have not been authenticated by the authors.

- have the cell lines been tested for mycoplasma contamination? In this checklist, indicate on which page (or section and paragraph) the information can be found.

Cells were not tested for mycoplasma contamination.

Animal Models

Reported in section/paragraph or page #:

8. Report species, strain, sex and age of animals

Methods -Proof-of-concept in 3-day systemic candidiasis infection model and Proof-of-concept in prophylactic treatment model of systemic candidiasis

9. For experiments involving live vertebrates, include a statement of compliance with ethical regulations and identify the committee(s) approving the experiments.

Methods - Ethics statement

10. We recommend consulting the ARRIVE guidelines (PLoS Biol. 8(6), e1000412,2010) to ensure that other relevant aspects of animal studies are adequately reported.

Human subjects

Reported in section/paragraph or page #:

11. Identify the committee(s) approving the study protocol.

Methods - Ethics statement

12. Include a statement confirming that informed consent was obtained from all subjects.

Methods - Ethics statement

13. For publication of patient photos, include a statement confirming that consent to publish was obtained.

Not applicable

14. Report the clinical trial registration number (at ClinicalTrials.gov or equivalent).

Not applicable

15. For phase II and III randomized controlled trials, please refer to the CONSORT statement and submit the CONSORT checklist with your submission.

Not applicable

16. For tumor marker prognostic studies, we recommend that you follow the REMARK reporting guidelines.

Not applicable

(Continues on following page)

Data deposition

Reported in section/paragraph or page #

17. Provide accession codes for deposited data.

Data deposition in a public repository is mandatory for:

a. Protein, DNA and RNA sequences
b. Macromolecular structures
c. Crystallographic data for small molecules
d. Microarray data

Not applicable

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available here. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad. We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse

18. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

Not applicable