

Life Sciences Reporting Summary

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Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study. [For final submission](#): please carefully check your responses for accuracy; you will not be able to make changes later.

► Experimental design

1. Sample size

Describe how sample size was determined.

Each experiment was usually performed thrice. The sample size was chosen in order to achieve statistical significance. The exact number of samples (donors/mice) for the individual experiments are mentioned in the figure legends.

2. Data exclusions

Describe any data exclusions.

Data were generally not excluded

3. Replication

Describe the measures taken to verify the reproducibility of the experimental findings.

Experiments were repeated using different human donors and mice to check for reproducibility. Attempts at replication were successful.

4. Randomization

Describe how samples/organisms/participants were allocated into experimental groups.

Mice were randomly assigned to the experimental groups.

5. Blinding

Describe whether the investigators were blinded to group allocation during data collection and/or analysis.

No blinding was done but the major findings were also independently verified to be consistent by our collaborators in this study.

Note: all in vivo studies must report how sample size was determined and whether blinding and randomization were used.

6. Statistical parameters

For all figures and tables that use statistical methods, confirm that the following items are present in relevant figure legends (or in the Methods section if additional space is needed).

- | n/a | Confirmed |
|--------------------------|--|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> The <u>exact sample size</u> (<i>n</i>) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.) |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A statement indicating how many times each experiment was replicated |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> The statistical test(s) used and whether they are one- or two-sided
<i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i> |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A description of any assumptions or corrections, such as an adjustment for multiple comparisons |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Test values indicating whether an effect is present
<i>Provide confidence intervals or give results of significance tests (e.g. P values) as exact values whenever appropriate and with effect sizes noted.</i> |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A clear description of statistics including <u>central tendency</u> (e.g. median, mean) and <u>variation</u> (e.g. standard deviation, interquartile range) |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Clearly defined error bars in <u>all</u> relevant figure captions (with explicit mention of central tendency and variation) |

See the web collection on [statistics for biologists](#) for further resources and guidance.

► Software

Policy information about [availability of computer code](#)

7. Software

Describe the software used to analyze the data in this study.

Data were statistically analysed using GraphPad Prism version 5.04

For manuscripts utilizing custom algorithms or software that are central to the paper but not yet described in the published literature, software must be made available to editors and reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). *Nature Methods* [guidance for providing algorithms and software for publication](#) provides further information on this topic.

► Materials and reagents

Policy information about [availability of materials](#)

8. Materials availability

Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a third party.

No restrictions

9. Antibodies

Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).

All antibodies used in this study were purchased from commercial sources. The source of the antibodies are mentioned in the methods section.

10. Eukaryotic cell lines

a. State the source of each eukaryotic cell line used.

Primary human dendritic cells and neutrophils were freshly isolated from buffy coat of healthy donors provided by Karolinska University Hospital and Uppsala University Hospital. Human THP-1 cell line was purchased from ATCC [TIB-202].

b. Describe the method of cell line authentication used.

Cell-lines were authenticated by using different frozen batches of cell lines and experiments were repeated using different batches of cells.

c. Report whether the cell lines were tested for mycoplasma contamination.

The THP-1 cell line was tested for mycoplasma and found to be negative. The primary cells were freshly isolated from buffy coat and used directly for the experiments.

d. If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by [ICLAC](#), provide a scientific rationale for their use.

N/A

► Animals and human research participants

Policy information about [studies involving animals](#); when reporting animal research, follow the [ARRIVE guidelines](#)

11. Description of research animals

Provide all relevant details on animals and/or animal-derived materials used in the study.

All mice experiments were performed in accordance with the local ethical committee (Stockholms Norra djurförsöksetiska nämnd). Six- to seven- weeks old male wild-type C57BL/6J mice were purchased from Charles River Laboratories. Experiments with MRC-1^{-/-} mice were done at the University of Liverpool with the approval of the UK Home Office and the University of Liverpool ethics committee. MRC-1^{-/-} mice were generated on a mixed 129SvJ and C57BL/6 background, and then backcrossed to C57BL/6 strain for at least 7 generations. Homozygous knockout mice were bred and maintained at the University of Nottingham and were a generous gift of Dr. Luisa Martinez-Pomares (University of Nottingham).

Policy information about [studies involving human research participants](#)

12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants.

N/A