

1 **Supplementary Material for:**  
2 **Metabolic physiology explains macroevolutionary trends in the melanic**  
3 **colour system across amniotes**

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11

## 12 Supplementary Methods

13 *Assessing taxon sampling*—In total, we obtained morphological data for 255 species in the  
14 final tree (see figure S1). To assess the evenness of this taxonomic sampling, we used mean  
15 nearest taxonomic distance (MNTD) [1]. We calculated the MNTD for species with  
16 morphological data and compared this value to a null distribution generated by randomly  
17 shuffling tips [1]. The observed MNTD was ~58 My, which was not significantly smaller than  
18 expected by chance ( $P_{\text{rand}} = 0.13$ ) suggesting that our sampling is not significantly biased or  
19 uneven [1].

20  
21 *Incorporating fossils*—We added extinct lepidosaurs and turtles with available melanosome  
22 data either as polytomies at the base of the Iguanidae clade or as the sister taxon to the lone  
23 extant representative of the Testudines clade, respectively, following [2]. For theropod  
24 dinosaurs, we used the R code, fossil ages, and published cladogram of [3] to generate a fossil  
25 subtree with the timePaleoPhy function in paleotree [4]. We grafted this time-calibrated  
26 fossil tree along the branch leading to birds in our extant supertree using custom R code  
27 (available on Dryad). Additional extinct archosaur taxa not represented in the published  
28 theropod tree [3] were added as sister taxa to clades in the supertree based on published  
29 divergence times [3,5] and with fossils as minimum ages [2]. Finally, we added three  
30 additional fossil taxa as polytomies (1 undescribed pterosaur, 1 undescribed psittacosaur,  
31 and 1 undescribed enantiornithine; see table S1 for specimen details).

32  
33 *Sensitivity analysis*—Comparative methods that estimate shifts in the rate of phenotypic  
34 evolution generally do not require a fully resolved phylogeny, but rather that taxa are  
35 sampled uniformly with respect to the trait of interest [6]. To assess the sensitivity of our rate  
36 analyses to taxon sampling, we employed a jackknife approach following [7]. To do this, we  
37 created 25 subtrees by randomly removing 10 taxa. We then re-ran rate shift analyses on all  
38 random subtrees. The results show that the inferred rate shifts in Maniraptora (dinosaurs and  
39 birds) and Passeres (songbirds) are robust to removal of taxa ( $n=10$ ), while the decrease in  
40 evolutionary rate at the base of the Sauropsida clade (non-avian reptiles, birds, crocodiles) is  
41 not (figure S3). This could be because of denser sampling of taxa with fossil melanosomes in  
42 feathered dinosaurs and birds (figure S2).

43  
44 *Clade-specific trends in evolutionary rates*— We compared among clade differences in  
45 evolutionary rates and covariation among traits using a Bayesian approach implemented in  
46 the ratematrix package [8]. This method reconstructs shifts in evolutionary trait correlations  
47 and rates while accounting for uncertainty in when different integumentary structures  
48 evolved. We used stochastic character mapping [9] to map integument type (hair, skin/scales,  
49 and feathers) onto the pruned phylogeny, with the ancestral state of integument type to  
50 skin/scales based on extant and fossil evidence [10]. We fit separate models for brown and  
51 black colours for melanosome length and melanosome diameter. We ran two MCMC chains  
52 for 2 million generations each and assessed convergence (Gelman-Rubin  $R < 1.01$ ) with the  
53 checkConvergence function [8]. We compared evolutionary rates and covariation among

54 clades by calculating posterior distribution overlaps in the testRatematrix function [8].  
55 Overlaps >5% suggest that clades do not show distinct macroevolutionary trends.  
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57 **SUPPLEMENTARY TABLES**

58 **Table S1. List of fossil taxa used.** Data for samples 1-18 taken from Li et al. 2014 [2] and images  
59 used to measure melanosomes in sample 19 taken from McNamara et al. 2009 [11].  
60

Sample	Taxon	Catalogue number
1	<i>Anchiornis huxleyi</i>	BMNHC PH828
2	<i>Archaeopteryx lithographica</i>	MB.Av.100
3	<i>Caudipteryx zoui</i>	PMOL AD00020
4	<i>Confuciusornis sanctus</i>	CUGB G20070001
5	<i>Inkayacu paracasensis</i>	MUSM 1444
6	Undescribed ornithurine	CUGB G20100053
7	<i>Microraptor</i>	BMNHC PH881
8	Undescribed enantiornithine	CUGB P1201
9	Undescribed enantiornithine	CUGB G20120001
10	<i>Beipiaosaurus</i>	BMNHC PH000911
11	<i>Sinosauropteryx</i>	IVPP V14202
12	Undescribed pterosaur	BMNHC PH000988
13	Undescribed pterosaur	PMOL AP00022
14	<i>Psittacosaurus lujiatunensis</i>	PKUP V1050
15	<i>Psittacosaurus lujiatunensis</i>	PKUP V1051
16	Undescribed turtle	PKUP V1070
17	<i>Xianglong zhaoi</i>	PMOL 000666
18	<i>Yabeinosaurus sp.</i>	PKUP V1059
19	Undescribed frog	MNCN 63805

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63 **Table S2. Relationship between rates of melanosome shape evolution and predicted**  
64 **metabolic rates in amniotes.** Results of 'rate-by-state' tests. P values < 0.05 indicate **rates of**  
65 **melanosome shape evolution are** significantly correlated with predicted ancestral metabolic  
66 **rates for black (eumelanin-consistent) colours, brown (phaeomelanin-consistent) colours, and**  
67 **species mean values ("All").** Significance was assessed by simulating trait evolution 500 times.  
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Colour	r	P value
Black	0.270	0.004
Brown	0.303	0.010
All	0.189	0.006

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71 **Table S3. Bayesian phylogenetic mixed models (BPMM) results.** Columns show response  
 72 variables and estimates with 95% Bayesian credible intervals calculated with the HPDinterval  
 73 function in coda [12]. For multivariate response models, Wald tests showed overall significance  
 74 between melanosome morphology and metabolic rate for black (**eumelanin-consistent**) colours  
 75 ( $p < 0.001$ ) but not brown (**phaeomelanin-consistent**) colours ( $p = 0.52$ ).  
 76

Response	Term	Estimate (95% CI)	$P_{\text{MCMC}}$
Univariate (aspect ratio)	Intercept	2.36 (2.07, 2.65)	< 0.001
	Colour (brown)	-0.46 (-0.86, -0.10)	< 0.001
	BMR	0.65 (0.30, 1.04)	0.019
	BMR:Colour (brown)	-1.00 (-1.48, -0.51)	< 0.001
Multivariate (length, diameter)	Length	6.63 (6.54, 6.73)	< 0.001
	Diameter	5.85 (5.75, 5.95)	< 0.001
	Length:BMR	0.36 (0.24, 0.48)	< 0.001
	Diameter:BMR	0.15 (0.02, 0.29)	0.027
	Length:Colour (brown)	-0.27 (-0.41, -0.13)	< 0.001
	Diameter:Colour (brown)	-0.07 (-0.20, 0.07)	0.33
	Length:BMR:Colour (brown)	-0.44 (-0.62, -0.24)	< 0.001
	Diameter:BMR:Colour (brown)	-0.13 (-0.31, 0.05)	0.16

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79 **Table S4. Quadratic discriminant function analysis performance with and without including**  
80 **body mass.** Variables included: melanosome length, melanosome diameter, melanosome  
81 aspect ratio, and log mass (for 'Mass' model only). See [13] for details.  
82

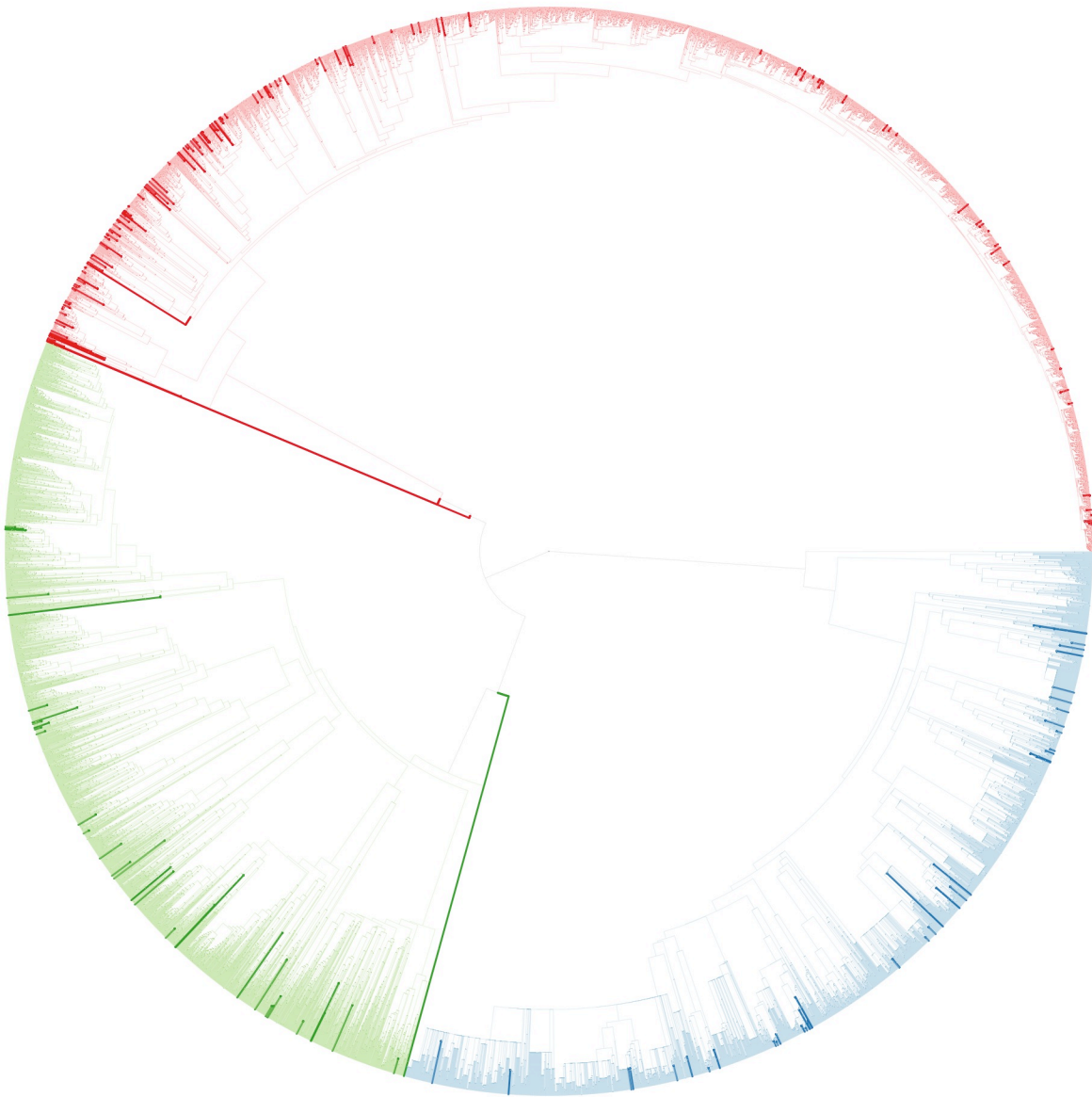
Model	Cross-validation	Self-test
No mass	0.753	0.630
Mass	0.798	0.686

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85 **SUPPLEMENTARY FIGURES**

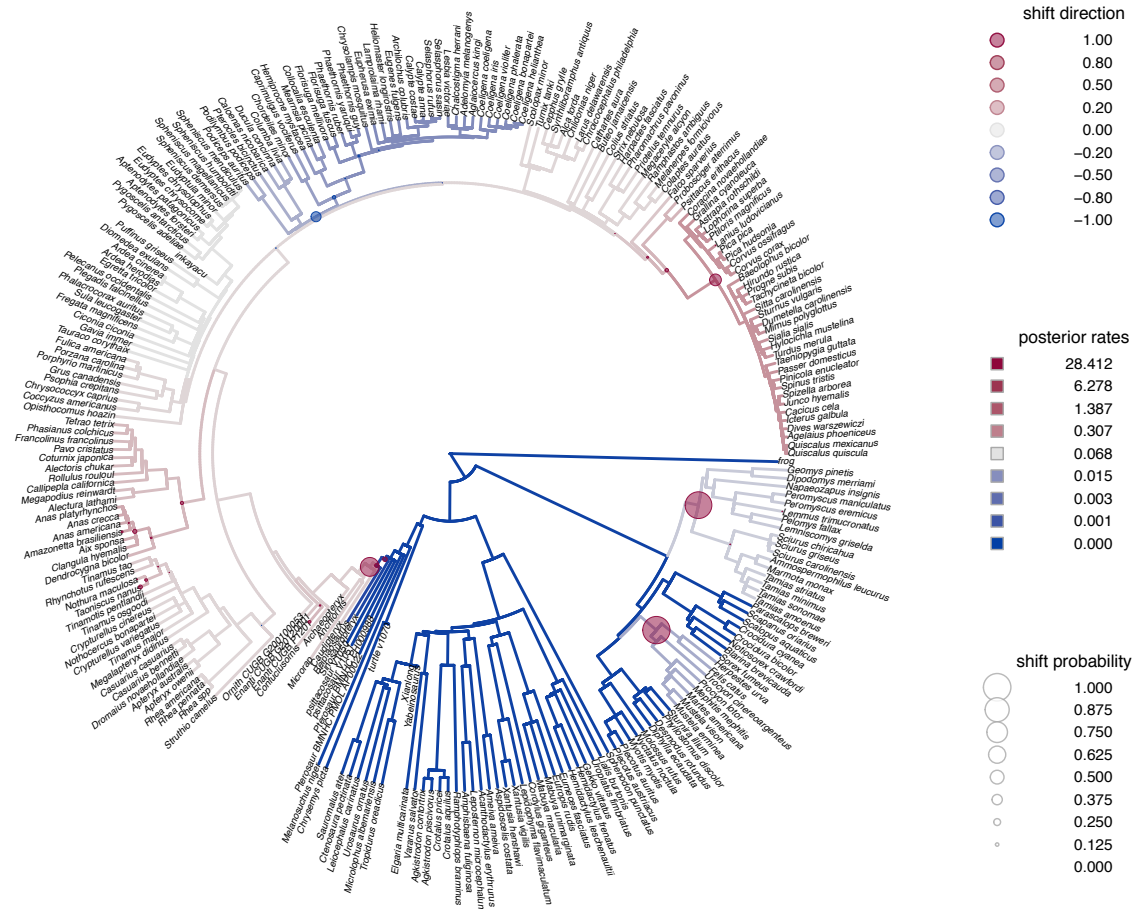
86 **Figure S1. Full synthetic supertree and taxon sampling in amniotes.** Tips coloured according to  
87 clade (red: birds, blue: mammals, green: non-avian reptiles), with darker shades indicating  
88 species with morphological data (n = 255) and lighter shades without morphological data.  
89 Species without morphological data were dropped from the tree prior to analysis. A tree file  
90 with readable tips is available on Dryad.



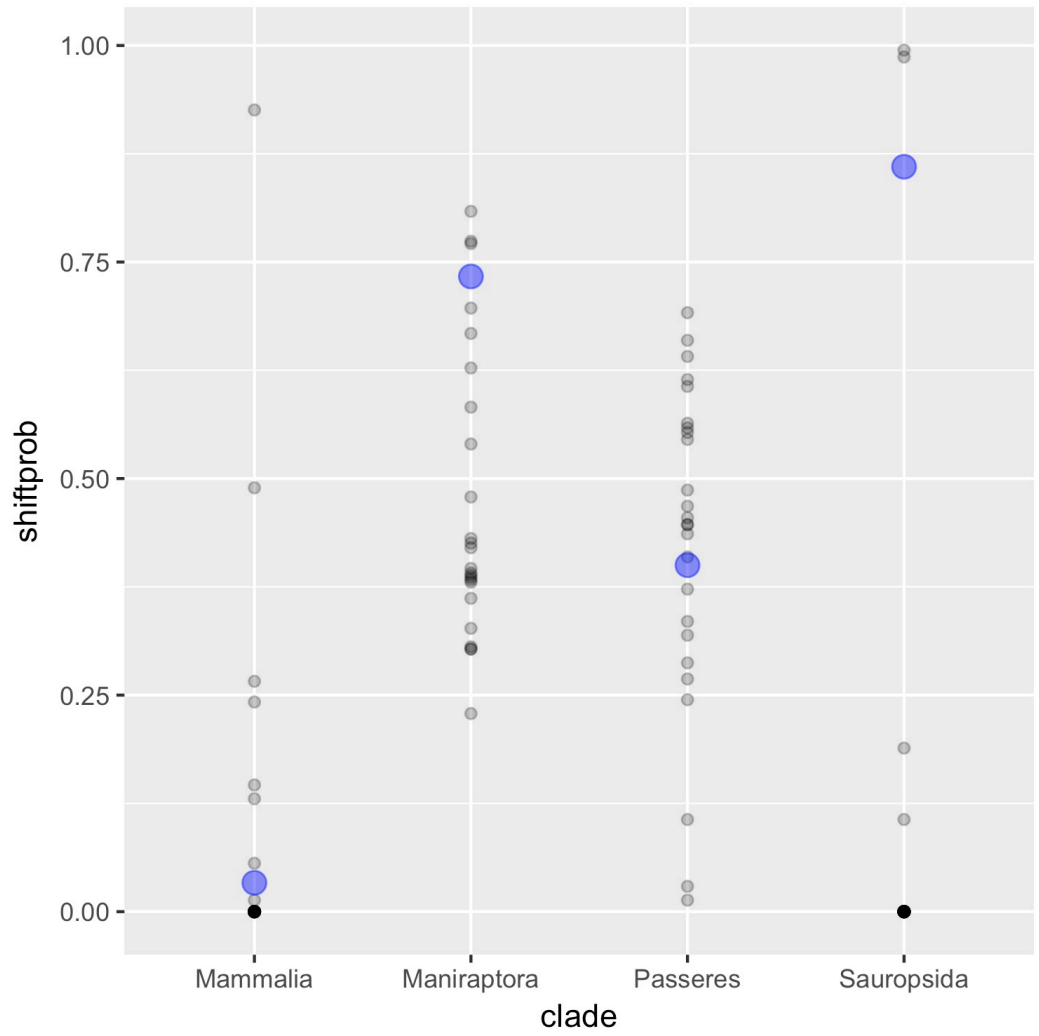
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93 **Figure S2. Full auteur rate shift results.** Branch colours correspond to estimated rates of  
 94 melanosome evolution (blue: slow, red: fast). Circles at nodes indicate estimated locations of  
 95 rate shifts, with size of circle indicating probability of a rate shift occurring at a given node.  
 96

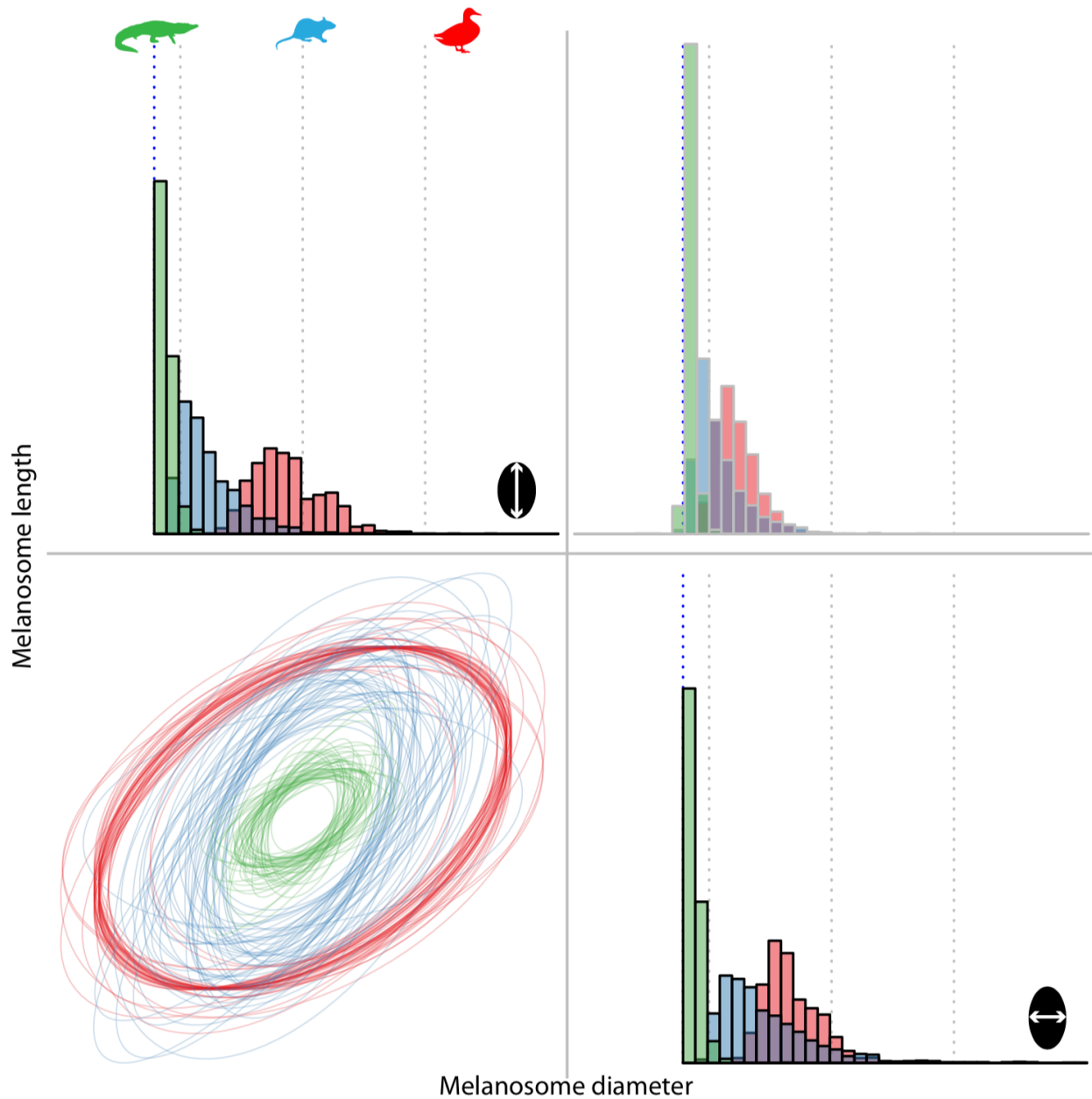


99 **Figure S3. Sensitivity analysis for rate shifts estimated with auteur.** Points show estimated  
100 shift probabilities for clades of interest (nodes highlighted in figure S2) after randomly removing  
101 10 taxa from the tree (i.e. 'taxonomic jack-knifing'). This analysis indicates the shifts in  
102 Maniraptora (feathered dinosaurs and birds) and Passeres (songbirds) are robust to taxon  
103 removal, but the decrease in evolutionary rate at the base of the Sauropsida clade (non-avian  
104 reptiles, birds, crocodiles) is not. Values prior to jack-knifing are shown in blue.  
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108 **Figure S4. Clade-specific differences in rates of melanosome shape evolution for black**  
109 **(eumelanin-consistent) integument colours.** Plots show posterior distribution of evolutionary  
110 rates (plot diagonals) and covariation (off-diagonal plots) between melanosome length and  
111 diameter for birds (red), mammals (blue), and non-avian reptiles (green). Birds and mammals  
112 show higher rates of morphological evolution compared to non-avian reptiles.  
113

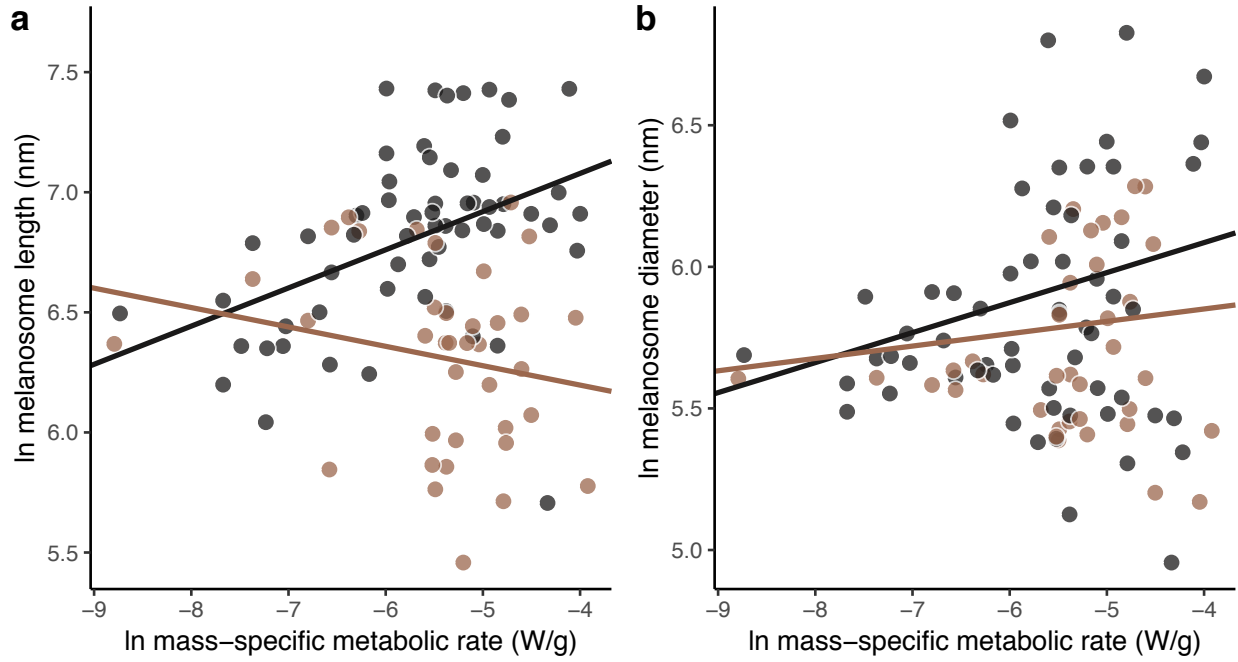


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129 **Figure S7. Relationship between melanosome morphology and mass-derived metabolic rate.**  
130 Results are shown for a subset of 77 species with both body mass and mass-specific metabolic  
131 rate data. Slopes for best-fit lines are derived from MCMCglmm analyses.  
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138 **for rain forest trees. *Am. Nat.* 156, 145–155.**
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