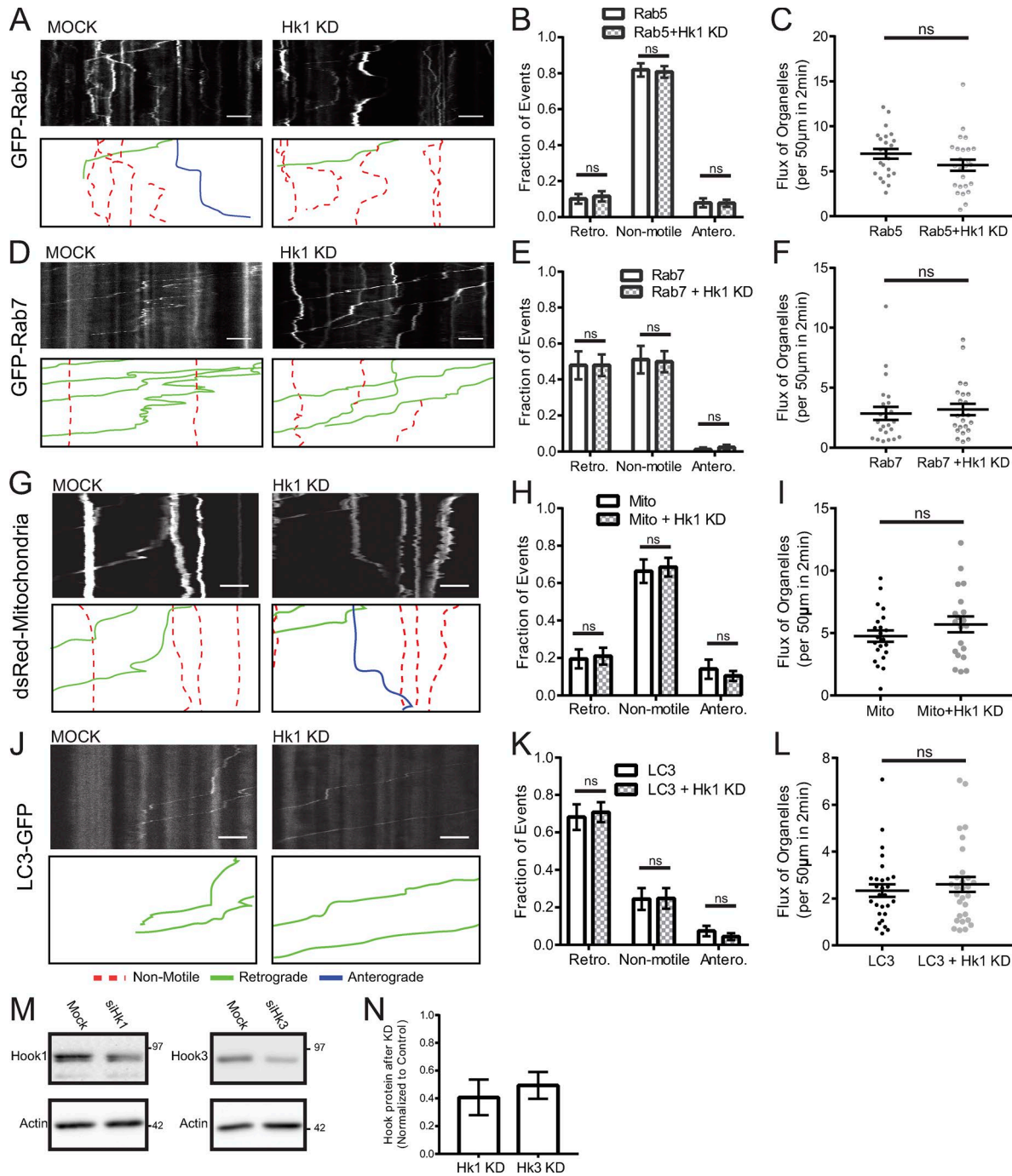


## Supplemental material

Olenick et al., <https://doi.org/10.1083/jcb.201805016>



**Figure S1. Hook1 KD does not significantly change Rab5, Rab7, mitochondria, or autophagosome motility.** (A) Kymograph of Rab5-GFP in control or Hk1 KD neurons. Traced events below are color coded for ease of interpretation. (B) Motility fractionated into retrograde, anterograde, and nonmotile events per neuron. Bar graph shows mean  $\pm$  SEM; two-way ANOVA ( $P > 0.9999$ ). Rab5:  $n = 23$  neurons; Rab5 + siHk1:  $n = 25$  neurons. (C) Flux of Rab5 organelles in control or Hk1 KD neurons. Scatter plot shows mean  $\pm$  SEM; unpaired  $t$  test ( $P = 0.1303$ ). Rab5:  $n = 23$  neurons; Rab5 + siHk1:  $n = 25$  neurons. (D) Kymograph of Rab7-GFP in control or Hk1 KD neurons. Traced events below are color-coded for ease of interpretation. (A and D) Bars, 5  $\mu$ m; 2 min total. (E) Motility fractionated into retrograde, anterograde, and nonmotile events per neuron. Bar graph shows mean  $\pm$  SEM; two-way ANOVA ( $P > 0.9999$ ). Rab7:  $n = 23$  neurons; Rab7 + siHk1:  $n = 23$  neurons. (F) Flux of Rab7 organelles in control or Hk1 KD neurons. Scatter plot shows mean  $\pm$  SEM; unpaired  $t$  test ( $P = 0.6504$ ). Rab7:  $n = 23$  neurons; Rab7 + siHk1:  $n = 23$  neurons. (G) Kymographs of mitochondria in control and Hook1 KD cells. Traced events below are color-coded for ease of interpretation. (G and J) Bars, 5  $\mu$ m; 1 min total. (H) Motility fractionated into retrograde, anterograde, and nonmotile events per neuron. Bar graph shows mean  $\pm$  SEM; two-way ANOVA ( $P > 0.93$ ). Mito:  $n = 22$  neurons; Mito + siHk1:  $n = 20$  neurons. (I) Flux of mitochondria in control or Hk1 KD neurons. Scatter plot shows mean  $\pm$  SEM; unpaired  $t$  test ( $P = 0.2292$ ). Mito:  $n = 22$ ; Mito + siHk1:  $n = 20$  neurons. (J) Kymographs of LC3-GFP in control and Hook1 KD cells. Traced events below are color-coded for ease of interpretation. (G and J) Bars, 5  $\mu$ m; 1 min total. (K) Motility fractionated into retrograde, anterograde, and nonmotile events per neuron. Bar graph shows mean  $\pm$  SEM; two-way ANOVA ( $P > 0.96$ ). LC3:  $n = 27$  neurons; LC3 + siHk1:  $n = 29$  neurons. (L) Flux of LC3-GFP in control or Hk1 KD neurons. Scatter plot shows mean  $\pm$  SEM; unpaired  $t$  test ( $P = 0.5335$ ). LC3:  $n = 28$  neurons; LC3 + siHk1:  $n = 29$  neurons. (M) Western blots of Hook1 and Hook3 siRNA KD in PC12 cells. (N) Quantification of KD Western blots from three individual repeats. Bar graph shows mean  $\pm$  SEM.

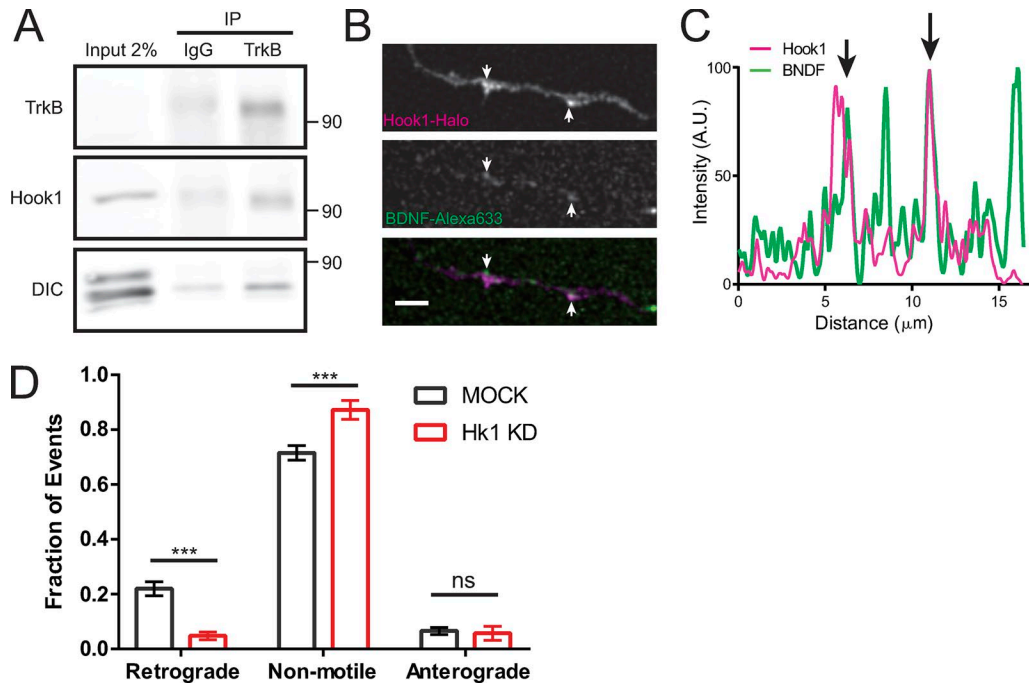


Figure S2. **Hook1 is present on TrkB-BDNF vesicles.** (A) Western blot of TrkB vesicle immunoprecipitation from mouse brain lysates. IP, immunoprecipitation. (B) Colocalization images of BDNF-Alexa Fluor 633 with Hook1-Halo in axon of hippocampal neuron. Arrows show Hook1 puncta colocalized with BDNF. Bar, 2 μm. (C) Line scan through axon in B. Arrows point to colocalized Hook1 and BDNF. (D) BDNF-Qdot motility fractionated into retrograde, anterograde, and nonmotile events per neuron. Bar graphs show mean ± SEM, two-way ANOVA (\*\*\*,  $P < 0.0001$ ; ns,  $P = 0.9931$ ). Mock:  $n = 53$  neurons; Hk1 KD:  $n = 44$  neurons. Some data are repeated from Fig. 2 E.

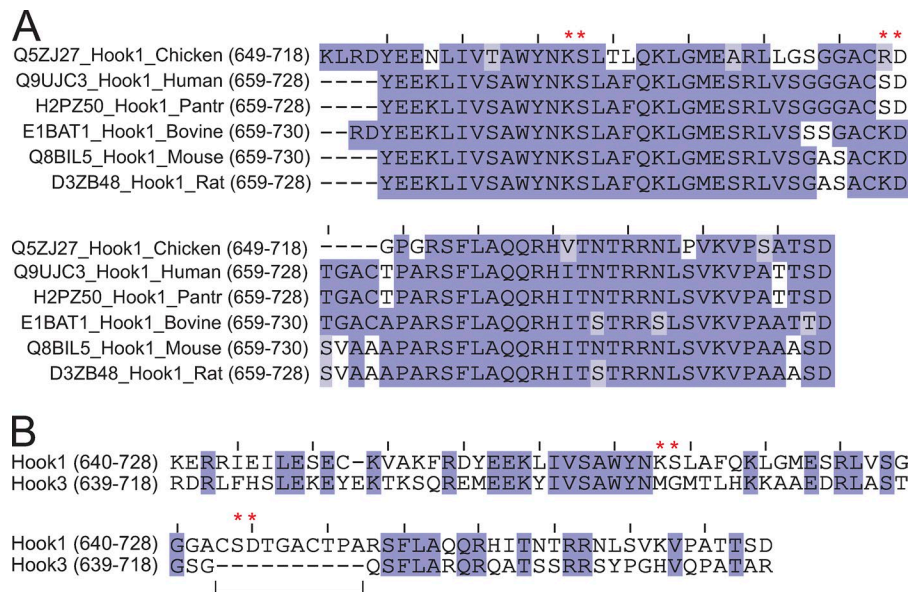
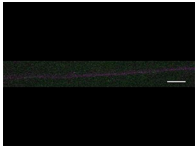
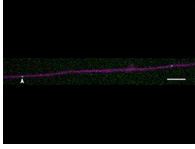


Figure S3. **Sequence analysis of the Hook1 and Hook3 C terminus.** (A) Sequence alignments of Hook1 C termini from different species as noted. Blossum62 coloring; red asterisks indicate mutated residues. (B) Sequence alignment of Hook1 and Hook3 C termini. Blossum62 coloring; red asterisks indicate mutated residues.



Video 1. **BDNF-Qdot motility in control neurons grown in microfluidic chambers.** GFP fill in magenta and Qdots in green, soma to the left. Bar, 5  $\mu\text{m}$ ; four frames per second with playback at 7.5 $\times$  real time.



Video 2. **BDNF-Qdot motility in Hk1 KD neurons grown in microfluidic chambers.** GFP fill in magenta and Qdots in green, soma to the left. Bar, 5  $\mu\text{m}$ ; four frames per second with playback at 7.5 $\times$  real time.