

SUPPLEMENTAL INFORMATION

SUPPLEMENTAL FIGURES

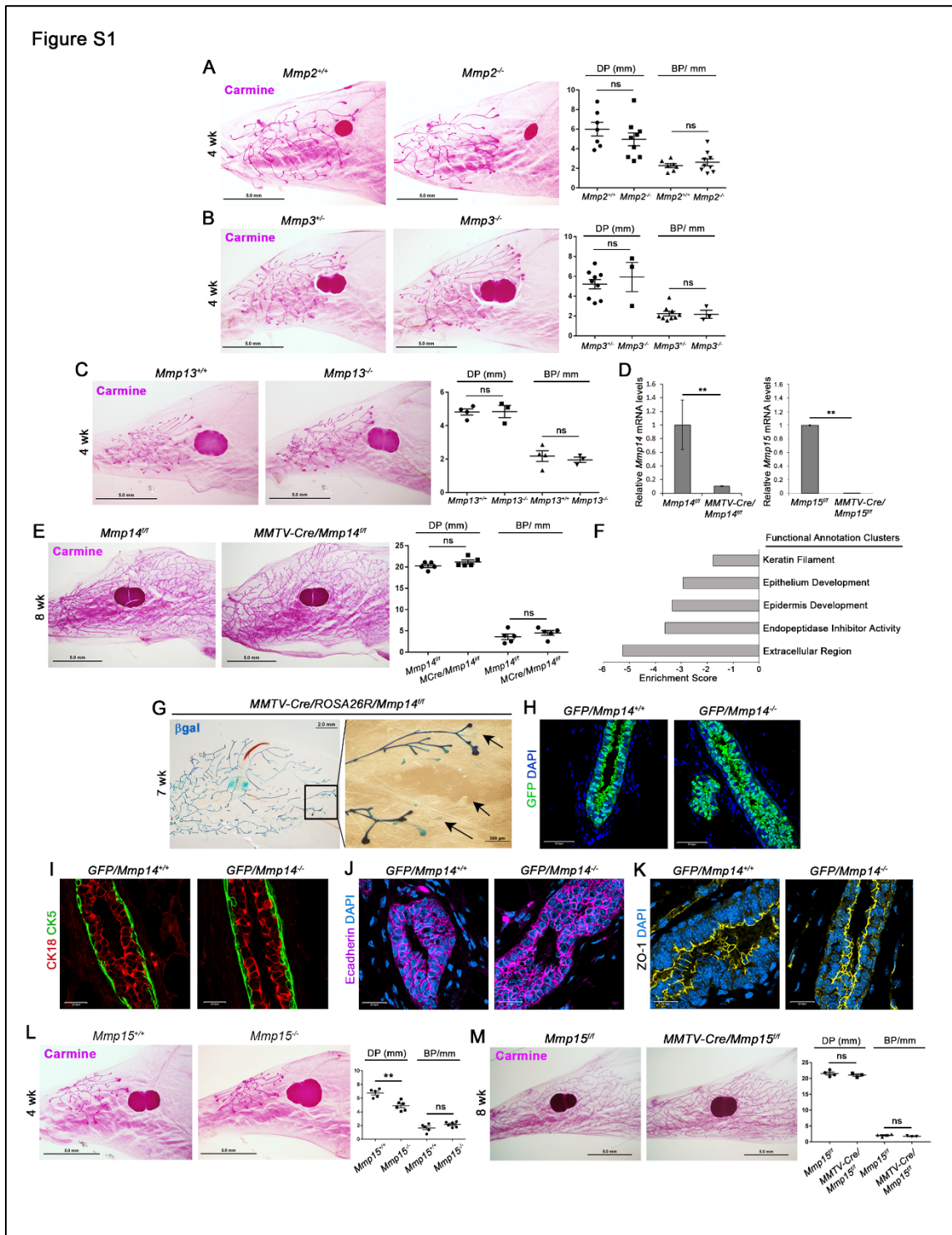


Figure S1. Postnatal mammary gland branching proceeds independently of MMP2, MMP3 and MMP13 as well as epithelial cell-derived MT1-MMP and MT2-MMP *in vivo*, Related to Figure 1.

- (A) Carmine staining of mammary gland whole mounts from 4 week-old *Mmp2*^{+/+} (n=7) and *Mmp2*^{-/-} (n=9) mice with quantification of ductal penetration (DP) (mm) and branch points per mm duct (BP/mm). Data are presented as mean ± SEM.
- (B) Carmine staining of whole mounts from 4 week-old *Mmp3*^{+/+} (n=9) and *Mmp3*^{-/-} (n=3) mice with quantification of DP (mm) and BP per mm duct. Data are presented as mean ± SEM.
- (C) Carmine staining of whole mounts from 4 week-old *Mmp13*^{+/+} (n=4) and *Mmp13*^{-/-} (n=3) mice with quantifications of DP (mm) and BP per mm duct. Data are presented as mean ± SEM.
- (A-C) Scale bar, 5.0 mm.
- (D) QPCRs for *Mmp14* and *Mmp15* in mammary epithelial cells harvested from 4 week-old *Mmp14*^{fl/fl} and *MMTV-Cre*^{+/-}/*Mmp14*^{fl/fl} mice, and from 4 week-old *Mmp15*^{fl/fl} and *MMTV-Cre*^{+/-}/*Mmp15*^{fl/fl}, as normalized to *Gapdh* (n=4, **p<0.001; mean ± SEM).
- (E) Carmine-staining of mammary gland whole mounts isolated from 8 week-old *Mmp14*^{fl/fl} and *MMTV-Cre*^{+/-}/*Mmp14*^{fl/fl} mice (scale bar, 5.0 mm) with quantification of DP (mm) and BP per mm duct from n=5 mice per genotype. Data are presented as mean ± SEM.
- (F) Chart of Functional Annotation Clusters and Gene ontology (GO) categories down-regulated in mammary epithelial cells harvested from 4 week-old *Mmp14*^{fl/fl} and *MMTV-Cre*^{+/-}/*Mmp14*^{fl/fl} mice (n=3 per genotype) with associated enrichment scores.
- (G) LacZ staining of *MMTV-Cre*^{+/-}/*Mmp14*^{fl/fl}/*Rosa26R*^{+/-loxP} mammary gland whole mount from 7 week-old mice (scale bar, 2.0 mm). A box circumscribes the region at higher magnification (scale bar, 500 μm). Arrows indicate the wild-type (Cre-negative) ducts.
- (H) GFP fluorescence with DAPI staining in mammary tissue cross-sections at 8 weeks post-transplantation of *Gfp*^{+/-}/*Mmp14*^{+/+} and *Gfp*^{+/-}/*Mmp14*^{-/-} ducts (scale bar, 50.0 μm).

- (I-K) Immunofluorescence for CK18 and CK5 (I), E-cadherin (J) and Zonula occludens (ZO)-1 with DAPI staining (K) in mammary tissue cross-sections at 8 weeks post-transplantation of *Gfp^{+/-}Mmp14^{+/-}* and *Gfp^{+/-}Mmp14^{-/-}* ducts (scale bar, 20.0 μ m).
- (L) Carmine-staining of mammary gland whole mounts isolated from 4 week-old *Mmp15^{+/+}* (n=5) and *Mmp15^{-/-}* (n=6) mice (scale bar, 5.0 mm) with quantification of DP (mm) and BP per mm duct. Data are presented as mean \pm SEM. (**p=0.0012)
- (M) Carmine-staining of mammary gland whole mounts isolated from 8 week-old *Mmp15^{ff}* (n=4) and *MMTV-Cre^{+/-}/Mmp15^{ff}* mice (n=3) (scale bar, 5.0 mm) with quantification of DP (mm) and BP per mm duct. Data are presented as mean \pm SEM.

Figure S2

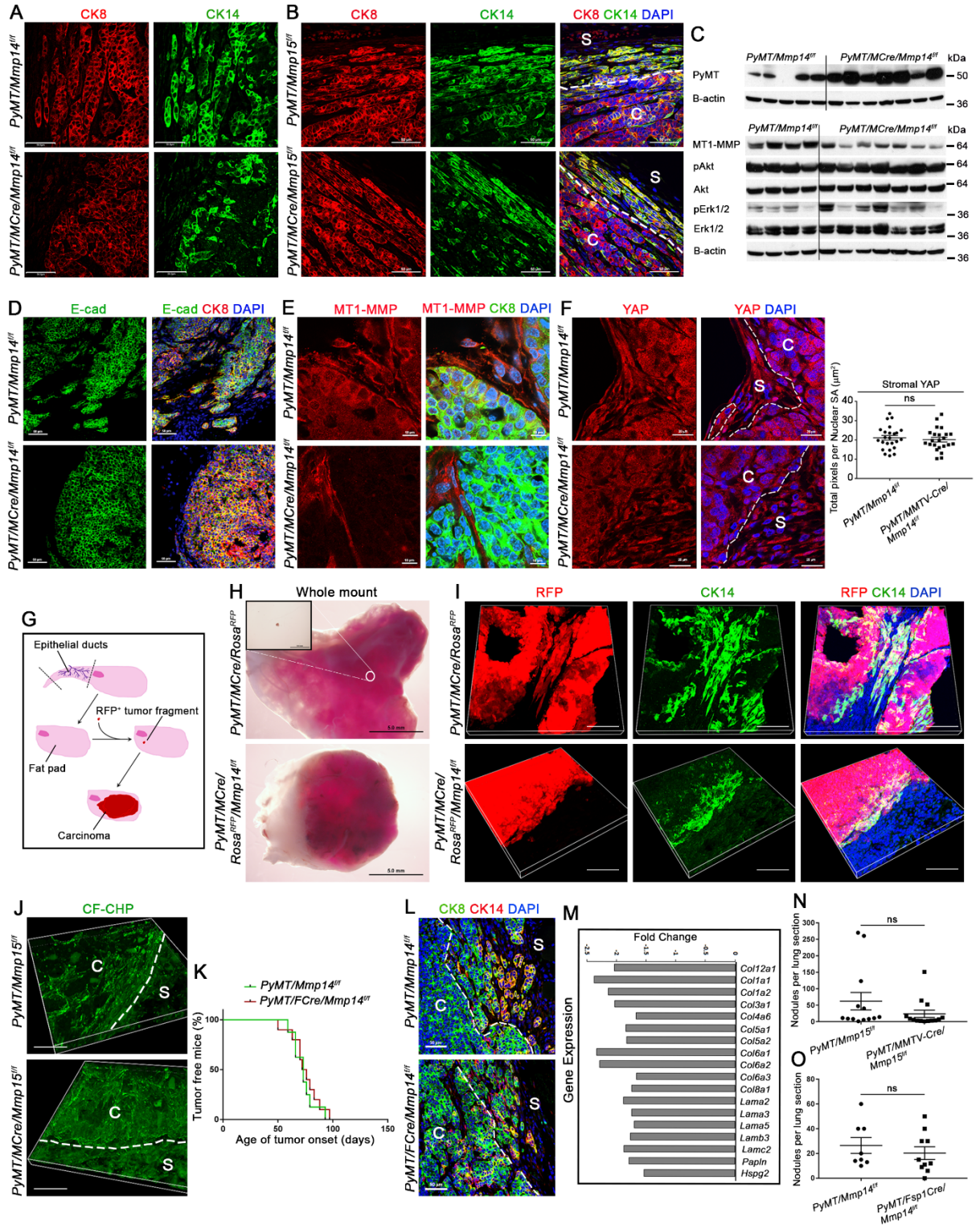


Figure S2. Carcinoma cell-derived MT1-MMP controls local invasion and metastasis independently of MT2-MMP or stromal cell-derived MT1-MMP, Related to Figure 2.

- (A) CK8 and CK14 immunofluorescence in mammary tumors harvested from 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{ff}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{ff}* mice (scale bar, 50.0 μ m).
- (B) CK8 and CK14 immunofluorescence with DAPI staining in mammary tumor cross-sections from 3-4 month-old *MMTV-PyMT^{+/-}/Mmp15^{ff}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp15^{ff}* mice (scale bar, 50.0 μ m). Dotted lines mark the tumor-stromal interface. "S" indicates the stroma and "C" indicates the carcinoma in each section.
- (C) Western blots for PyMT, MT1-MMP, pAkt, Akt, pErk1/2 and Erk1/2 in mammary tumors harvested from 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{ff}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{ff}* mice, as normalized to β -actin.
- (D) Immunofluorescence of E-cadherin with CK8 and DAPI staining at the invasive edge of mammary tumor cross-sections from 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{ff}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{ff}* mice (scale bar, 50 μ m).
- (E) MT1-MMP immunofluorescence in 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{ff}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{ff}* mammary tumors (scale bar, 10 μ m).
- (F) Stromal cell YAP immunofluorescence in 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{ff}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{ff}* mammary tumors. Dotted line marks the tumor-stromal interface (scale bar, 20 μ m). Quantification of the total pixels of YAP immunofluorescence per stromal cell nuclear surface area (μ m²) in n=4 tumors and n>20 fields per genotype. Data are presented as mean \pm SEM.
- (G) Cartoon of mammary tumor fragment transplantation.
- (H) Whole mounts of RFP⁺ tumors derived from *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Rosa^{RFP}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Rosa^{RFP}/Mmp14^{ff}* mammary tumor fragments at 7 weeks post-transplantation (scale bar, 5.0 mm). Inset shows tumor fragment before transplant

(scale bar, 5.0 mm). Circle drawn on tumor at 7 weeks indicates initial fragment size for comparison. Note the corrugated boundaries of the wild-type tumor mass relative to the conditional knockout.

- (I) 3-D reconstructions of RFP fluorescence and CK14 immunofluorescence with DAPI staining in tumors derived from *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Rosa^{RFP}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Rosa^{RFP}/Mmp14^{fl/fl}* mammary tumor fragments at 7 weeks post-transplantation (scale bar, 50 μ m).
- (J) 3-D reconstructions of CF-CHP immunofluorescence in mammary tumors harvested from 3-4 month-old *MMTV-PyMT^{+/-}/Mmp15^{fl/fl}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp15^{fl/fl}* mice. Dotted lines mark the tumor-stromal interface (scale bar, 30 μ m).
- (K) Kaplan-Meier plots depicting age of tumor onset (days) for *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* (n=12) versus *MMTV-PyMT^{+/-}/Fsp1-Cre^{+/-}/Mmp14^{fl/fl}* (n=13) mice.
- (L) CK8 and CK14 immunofluorescence with DAPI staining in mammary tumor cross-sections from 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* and *MMTV-PyMT^{+/-}/Fsp1-Cre^{+/-}/Mmp14^{fl/fl}* mice (scale bar, 50 μ m). Dotted lines mark the tumor-stromal interface.
- (M) Changes in ECM transcriptomes in 3-4 month-old *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{fl/fl}* mammary tumors (n=4 per genotype).
- (N) Quantification of lung nodules per cross-section in lungs harvested from *MMTV-PyMT^{+/-}/Mmp15^{fl/fl}* (n=13) and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp15^{fl/fl}* (n=14) mice. Data are presented as mean \pm SEM.
- (O) Quantification of lung nodules per cross-section in lungs harvested from *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* (n=8) and *MMTV-PyMT^{+/-}/Fsp1-Cre^{+/-}/Mmp14^{fl/fl}* (n=10) mice. Data are presented as mean \pm SEM.

Figure S3

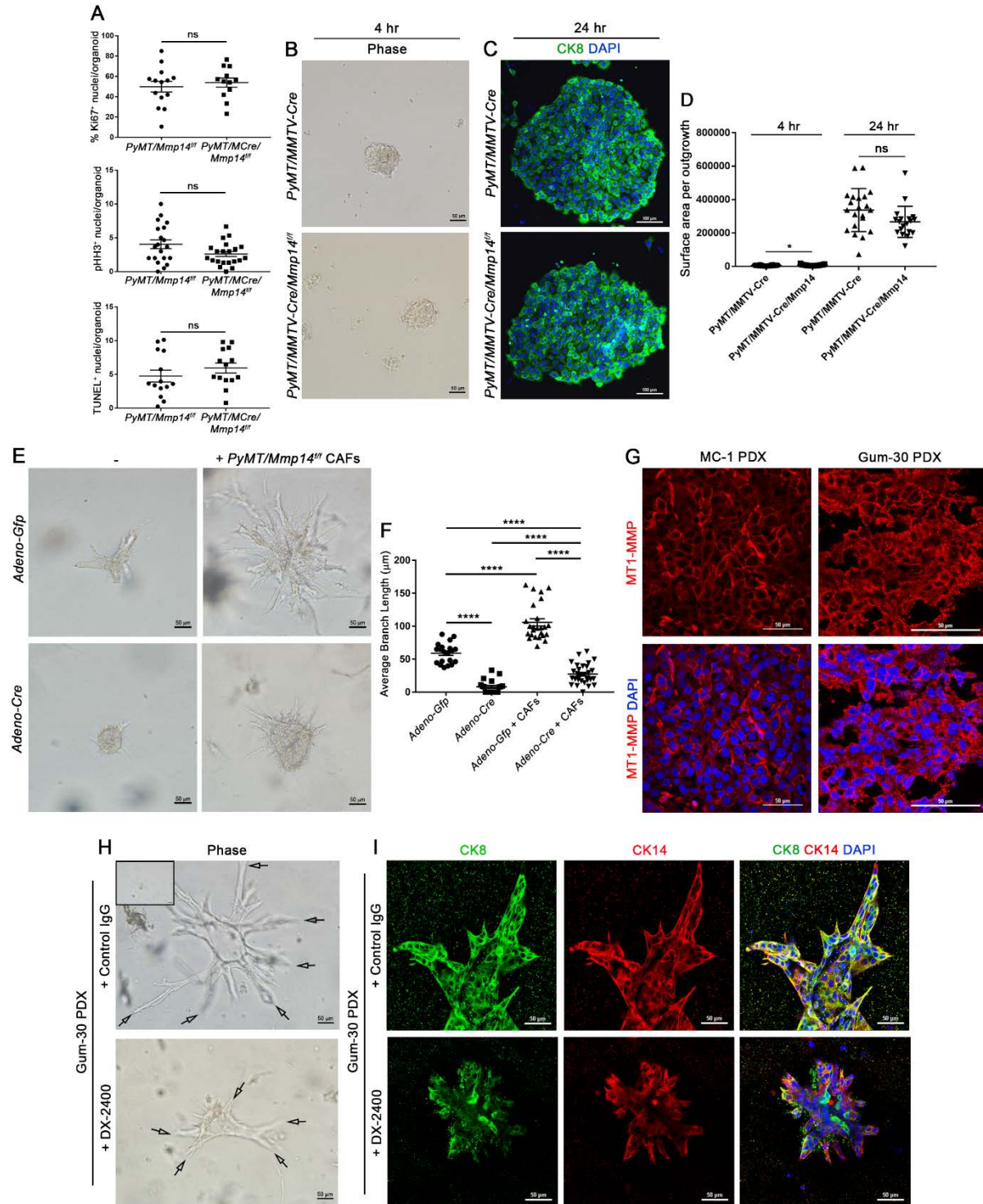


Figure S3. MT1-MMP directs both mouse and human breast carcinoma invasion programs *ex vivo*, Related to Figure 3.

- (A) Quantification of the percentage of Ki67-positive nuclei per organoid ($n \geq 12$ per genotype) (top panel), the number of phospho-histone H3 (pHH3)-positive nuclei per organoid ($n \geq 20$ per genotype) (middle panel), and the number of TUNEL-positive nuclei per organoid ($n = 14$ per genotype) (lower panel) in *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{fl/fl}* mammary carcinoma-derived organoids after 3 days of culture within 3-D type I collagen with FGF-2. Data are presented as mean \pm SEM.
- (B) Bright-field micrographs of *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{fl/fl}* mammary carcinoma-derived organoids cultured atop a 3-D type I collagen hydrogel with FGF-2 at 4 hours.
- (C) CK8 immunofluorescence with DAPI counter-staining in *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{fl/fl}* mammary carcinoma-derived organoids after 24 hours of culture atop a 3-D type I collagen hydrogel with FGF-2.
- (D) Quantification of *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{fl/fl}* mammary carcinoma-derived organoid surface area at 4 hours and following outgrowth at 24 hours ($n \geq 30$ at 4 hours and $n \geq 20$ at 24 hours from representative experiment). (* $p = 0.0141$ with *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* at $5813 \pm 433.5 \mu\text{m}^2$ and *MMTV-PyMT^{+/-}/MMTV-Cre^{+/-}/Mmp14^{fl/fl}* at $7622 \pm 570.9 \mu\text{m}^2$).
- (E) Bright-field micrographs of mammary carcinoma-derived organoids harvested from 3 month-old *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* mice and transduced with either Adenoviral-GFP or Adenoviral-Cre prior to embedding in 3-D type I collagen for 3 days with FGF-2 in the presence or absence of *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* tumor-derived CAFs (scale bar, 50 μm).
- (F) Quantifications of tumor organoid invasion, as represented by the average branch length (μm) of the three longest branches (**** $p < 0.0001$; $n \geq 19$ for organoid alone conditions and $n \geq 25$ for co-culture conditions). Data are presented as mean \pm SEM.

- (G) MT1-MMP immunofluorescence with DAPI staining in cross-sections from intact human patient-derived xenograft (PDX) tumors MC-1 and Gum30 (scale bar, 50 μm).
- (H) Bright-field micrographs of organoids harvested from PDX Gum30 tumors and embedded within 3-D type I collagen for 4 days with FGF-2 in the presence of the anti-MT1-MMP antibody (DX-2400) or a control IgG antibody (scale bar, 50 μm). Inset shows bright-field micrograph of Gum30 organoid at time 0. Arrows mark terminal ends of branching structures. Organoid branch length is reduced from $150.5 \pm 7.7 \mu\text{m}$ with 8.8 ± 0.9 branches/organoid in the presence of control IgG (n=18) to $65.0 \pm 6.1 \mu\text{m}$ with 4.9 ± 0.6 branches/organoid in the presence of DX-2400 (n=15) ($p \leq 0.001$, mean \pm SEM).
- (I) CK8 and CK14 immunofluorescence with DAPI staining in organoids harvested from PDX Gum30 tumors and embedded within 3-D type I collagen for 4 days with FGF-2 in the presence of DX-2400 antibody or control IgG antibody (scale bar, 50 μm).

Figure S4. Stromal cell-derived MT1-MMP controls mammary gland branching in an organ autonomous fashion independent of stromal cell-derived growth factors, macrophages or endothelial cells, Related to Figure 5.

- (A) Schematic of whole mammary gland transplant procedure.
- (B) Carmine-stained whole mounts of transplanted donor mammary glands from *Mmp14^{fl/fl}* or *Dermo1-Cre^{+/-}/Mmp14^{fl/fl}* mice at 10 days and 5 weeks post-transplantation (scale bar, 2.0 mm). “LN” indicates donor inguinal lymph node.
- (C) Bright-field micrographs of mammary epithelial organoids harvested from 6 week-old *Mmp14^{fl/fl}* and *Dermo1-Cre^{+/-}/Mmp14^{fl/fl}* mice and cultured within Matrigel with FGF-2 with quantification of branch number per organoid from n=12 fields per genotype (scale bar, 50 μ m). Data are presented as mean \pm SEM. Inset shows bright-field micrograph of organoid at time 0.
- (D) Gene ontology (GO) categories differentially expressed in 4 week-old *Dermo1-Cre^{+/-}/Mmp14^{fl/fl}* and wild-type mammary tissue (n=3 per genotype).
- (E) Bright-field micrographs of mammary epithelial organoids harvested from 6 week-old *Mmp14^{fl/fl}* mice and cultured within Matrigel alone under basal conditions (without FGF-2) (left panel), in the presence of 100 *Mmp14^{fl/fl}* fibroblasts per organoid (middle panel), or in the presence of 100 *Dermo1-Cre^{+/-}/Mmp14^{fl/fl}* fibroblasts per organoid (right panel) (scale bar, 50 μ m) with quantification of branch number per organoid (**p=0.0002, ****p<0.0001).
- (F) Senescence-associated β -galactosidase (SA- β gal) staining (pH 6.0) of mammary gland cross-sections from 4 week-old *Mmp14^{fl/fl}* and *Dermo1-Cre^{+/-}/Mmp14^{fl/fl}* mice with Eosin counter-staining (scale bar, 200 μ m).
- (G) F4/80 with α -smooth muscle actin (α SMA) immunofluorescence to delineate the myoepithelial cell compartment and DAPI staining (scale bar, 50 μ m) in *Mmp14^{fl/fl}* and *Dermo1-Cre^{+/-}/Mmp14^{fl/fl}* mammary gland cross-sections.

(H) CD31 (vascular endothelial cells) and Lyve1 (lymphatic endothelial cells) immunofluorescence with DAPI staining in *Mmp14^{ff}* and *Dermo1-Cre^{+/-}/Mmp14^{ff}* mammary gland cross-sections (scale bar, 100 μ m).

Figure S5

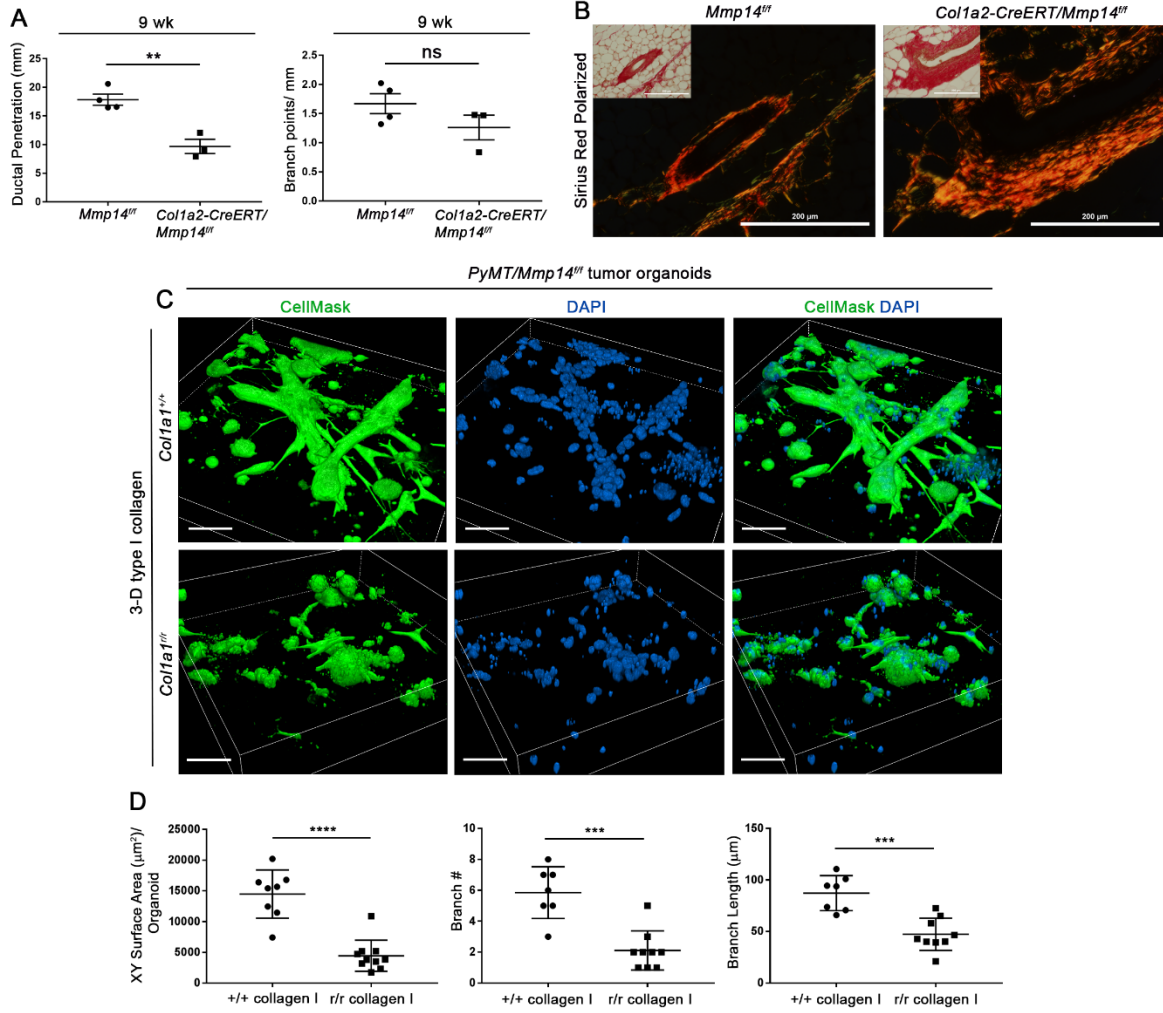


Figure S5. Postnatal mammary gland branching requires type I collagen remodeling by periductal fibroblasts while invading carcinoma cells directly remodel type I collagen, Related to Figure 6.

- (A) Quantification of ductal penetration (mm) and branch points per mm duct in 9 week-old mammary glands harvested from *Mmp14^{fl/fl}* (n=4) and *Col1a2-CreERT^{+/-}/Mmp14^{fl/fl}* (n=3) (**p=0.006).
- (B) Polarized light images of Sirius Red staining of 9 week-old *Mmp14^{fl/fl}* and *Col1a2-CreERT^{+/-}/Mmp14^{fl/fl}* glands (scale bar, 200 μ m). Insets show corresponding bright-field images.
- (C) 3-D reconstructions of *MMTV-PyMT^{+/-}/Mmp14^{fl/fl}* mammary tumor organoids within 3-D type I collagen extracted from *Col1a1^{+/+}* or *Col1a1^{fl/fl}* mouse tails and labeled with CellMask dye and DAPI after 4 days of culture with FGF-2. Carcinoma cells invade 3-D hydrogels of native, but not mutant, type I collagen (scale bar, 50 μ m).
- (D) Quantifications of XY surface area (μ m²), branch number and branch length (μ m) of organoids cultured within *Col1a1^{+/+}* or *Col1a1^{fl/fl}* type I collagen for 4 days with FGF-2 (****p<0.0001, ***p≤0.0005).

SUPPLEMENTAL TABLES

Table S1. Differential expression of epithelium-associated genes in array of *Mmp14*-targeted mammary epithelial cells, Related to Figure 1.

Gene	Fold change
<i>Glycam1</i>	-40.9
<i>Csn1s1</i>	-6.99
<i>Csn2</i>	-5.31
<i>Spr2a3</i>	-4.77
<i>Spr2a1</i>	-2.99
<i>Csn1s2a</i>	-2.39
<i>Epgn</i>	-2.07
<i>Wfdc21</i>	-2.06
<i>Krt1</i>	-2.03
<i>Krt10</i>	-1.95
<i>Spr2e</i>	-1.80
<i>Spr1b</i>	-1.78
<i>Lalba</i>	-1.77
<i>Muc13</i>	-1.56
<i>Krt6b</i>	-1.53
<i>Areg</i>	-1.52
<i>Spr1a</i>	-1.52

Table S1. Differential expression of epithelium-associated genes in Affymetrix microarray of mammary epithelial organoids isolated from *MMTV-Cre^{+/+}/Mmp14^{fl/fl}* female mice (n=2), as compared to *Mmp14^{fl/fl}* female littermates (n=3).

Table S2. Genotyping PCR Primers, Related to the STAR Methods Section.

Allele	Forward (5' to 3')	Reverse (5' to 3')
<i>Mmp14^{KO}</i>	<i>Mmp14^{WT}</i> : CTAGGCCTGGAACAT TCTAACGATC <i>Mmp14^{KO}</i> : GTGCGAGGCCAGAGGC CACTTGTGT	<i>Mmp14^R</i> : CTTTGTGGGTGACCCTGACTTGC
<i>Mmp14^{lacZ}</i>	<i>Mmp14^{lacZ}(+)</i> : ACCTGCGTGCAATCCATCTTG <i>Mmp14^{lacZ}(-)</i> : TGAGGTGGAAAACACGACCAG	<i>Mmp14^{lacZ}</i> : ATGATGGCGGAGGGATCGTTAG

<i>Mmp15</i> ^{KO}	<i>Mmp15</i> ^{WT} : CCGCCACCAAGCCTC ACTGTCT <i>Mmp15</i> ^{KO} : CGCCACCAAGCCTCA CTGTCT	<i>Mmp15</i> ^{WT} : AAAGCCACCCACGCC ATCAAAC <i>Mmp15</i> ^{KO} : AATTGCTGGGGATGG AGGAAGGTA
<i>Mmp15</i> ^{lacZ}	GAGATGGCGCAACGCAATTAATG	TGCACGTCCCATTCTCATGC
<i>Mmp2</i> ^{KO}	<i>Mmp2</i> ^{WT} : GTGCTACTGCAGGATA AACTGATG <i>Mmp2</i> ^{KO} : GCGCCTACCGGTGGA TGTGGAATGTGTGCG	<i>Mmp2</i> ^{WT} : CCGGGACAGGAACGTA CTGGGTTC <i>Mmp2</i> ^{KO} : CCGGGACAGGAACGTA CTGGGTTC
<i>Mmp3</i> ^{KO}	<i>Mmp3</i> ^{WT} : ACCGGATTTGCCAAG ACAGAGTG	<i>Mmp3</i> ^{WT} : GCATCTCCATTAATCCC TGGTCC
<i>Mmp13</i> ^{KO}	<i>Mmp13</i> ^{WT} : TTGGCCACTCCCTAGGTCT	<i>Mmp13</i> ^{WT} : CTACCCAGACAAGCAGTTTGC
<i>Neo</i>	AGGATCTCCTGTCATCTCAC CTTGCTCCTG	AAGAACTCGTCAAGAAGG CGATAGAAGGCG
<i>Mmp14</i> ^{flox}	GTTGAGGCAGGAGGATTGTGAGTT	CCTGGAAAAGTGGGCGAGAAG
<i>iCre</i>	CCGTTTGCCGGTCGTGGG	CGAATATCCTGGCAGCGATC
<i>MMTV-Cre</i>	GGTTCT GATCTGAGCTCTGAGTG	CATCACTCGTTGCATCGACCGG
<i>PyMT</i>	Internal Control (oIMR8744): CAAATGTTGCTTGTCTGGTG Trangene (oIMR0384): GGAAGCAAGTACTTCACAAGGG	Internal Control (oIMR8745): GTCAGTCGAGTGCACAGTTT Transgene (oIMR0385): GGAAAGTCACTAGGAGCAGGG
<i>Fsp1-Cre</i>	ATGCTTCTGTCCGTTTGCCG	CAATGCGATGCAATTTCTC
<i>Col1a1</i> ^f	TGGTTCTGGAATGAGGATGG	TGCTCTGCTTCCTTAGTGC

Table S3. Quantitative real-time PCR primers, Related to the STAR Methods Section.

Gene	Forward (5' to 3')	Reverse (5' to 3')
<i>Gapdh</i>	TGAAGCAGGCATCTGAGGG	CGAAGGTGGAAGAGTGGGAG
<i>Arbp</i>	CACTGGTCTAGGACCCGAGAA	AGGGGGAGATGTTCAGCATGT
<i>Mmp14</i>	CTGCCATTGCCGCCATGCAAAA	TGGCGTGGCACTCTCCCATACT
<i>Mmp15</i>	ACATGTCCACCATGCGCTCT	TACCATGATGTCAGCCTCC
<i>Mmp2</i>	TCTGGAGCGAGGATACCCCAA	TTCCAGGAGTCTGCGATGAGC
<i>Mmp3</i>	G TTCCTGATGTTGGTGGCTT	AGCCTCTCCTTCAGAGATCC
<i>Mmp13</i>	CTTTTCCTCCTGGACCAA ACT	TCATGGGCAGCAACAATAAA