SUPPORTING INFORMATION

Enzyme Architecture: Breaking Down the Catalytic Cage that Activates Orotidine 5'- Monophosphate Decarboxylase for Catalysis.

Archie C. Reyes,[†] David C. Plache,† Astrid P. Koudelka,† Tina L. Amyes,† John A. Gerlt,‡ and John P. Richard†,*

†Department of Chemistry, University at Buffalo, SUNY, Buffalo, New York 14260-3000. ‡ Department of Chemistry and Biochemistry, University of Illinois, Urbana, Illinois 61801.

* Author to whom correspondence should be addressed: *Email:* jrichard@buffalo.edu

RESULTS

The kinetic parameters from Figure 2A were determined using the Michaelis-Menten equation, with the assumption that $[OMP] \gg [E \cdot OMP]$, where $[OMP]$ and $[E \cdot OMP]$ are the concentration of free OMP and the Michaelis complex, respectively. Table S1 shows the values of [E•OMP] calculated for Figure 2A using the kinetic parameters obtained from the fit of the kinetic data to the Michaelis-Menten equation, and the initial enzyme concentration $[E]_0$ and substrate concentration $[OMP]_0$. Table S1 shows for all cases that $[E_•OMP]$ is < 10 % of $[OMP]_0$. The uncertainty in the derived kinetic parameter (k_{cat}/K_m) introduced by the accumulation of E•OMP is along the lines of that introduced into steady-state kinetic treatments by the small decrease in initial velocity observed over the first *ca* 10% reaction of substrate.

^a [E•OMP] = $[(v/(v_{\text{max}})][E]_0$, where v_{max} is the observed activity when $[E\text{-OMP}] = [E]_0$.

Page S3

Figure S1 shows the linear plot of *v*/[E] against [FOMP] for *Ec*OMPDC-catalyzed decarboxylation of FOMP, with slope of $k_{cat}/K_m = 2.7 \times 10^6$ M⁻¹ s⁻¹. Figures S2A–D shows timecourses for the change in absorbance at 282 nm for decarboxylation of FOMP catalyzed by the S154A/Y217F (Figure S2A), S154A/R235A (Figure S2B), S154A/Q215A/Y217F (Figure S2C) and the S154A/Q215A/R235A (Figure S2D) mutants of *Sc*OMPDC. The solid lines show the fit of the experimental data for each Figure to the exponential decay for the pseudo first-order enzyme-catalyzed reaction of FOMP, to give the observed first order rate constant k_{obsd} (s⁻¹) for OMPDC-catalyzed decarboxylation. In each case the data are fit by a simple exponential decay, which shows that the reactions are first order in [FOMP] and that the starting concentration $[FORMP] = 0.06-0.15$ mM $<< K_m$ for the OMPDC-catalyzed decarboxylation reaction. Each of the experiments from Figures S2 were repeated at least two times, and the values of $k_{obsd}/[E]$ for a given mutant *Sc*OMPDC-catalyzed decarboxylation were reproducible to better than ±10%.

Figure S1. Dependence of $v/[E]$ (s⁻¹) for decarboxylation of FOMP catalyzed by wild-type *Ec*OMPDC on the concentration of FOMP for reactions at 25 °C, pH 7.1 (10 mM MOPS) and at $I = 0.105$ (NaCl).

Figure S2. Full time-course for the reaction of FOMP catalyzed by mutants of *Sc*OMPDC for reactions at 25 °C, pH 7.1 (10 mM MOPS) and at *I* = 0.105 (NaCl). Figure S1A: decarboxylation of 60 µM FOMP catalyzed by 3.0 µM of the S154A/Y217F mutant of *Sc*OMPDC. Figure S1B: decarboxylation of 60 µM FOMP catalyzed by 0.02 mM of the S154A/R235A mutant of *Sc*OMPDC. Figure S1C: decarboxylation of 60 µM FOMP catalyzed by 0.01 mM of the S154A/Q215A/Y217F mutant of *Sc*OMPDC. Figure S1D: decarboxylation of 150 µM FOMP catalyzed by 6 µM of the S154A/Q215A/R235A mutant of *Sc*OMPDC.