Supporting Information

Modular Synthesis of Cell Permeating 2-Ketoglutarate Esters

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Figure S1. Immunoblot confirming effects of 2-KG diester **3** on cellular HIF-1 α levels. HIF-1 α -ODD-luciferase HeLa cells were treated with either vehicle, (lane 1), known HIF-1 α stabilizer CoCl₂ (0.5 mM, lane 2) 2-KG diester **3** (1 mM, lane 3; 0.5 mM, lane 4). Lane 1 was aligned next to lane 2 for clarity; full blot data is available upon request. Green box indicates observation of HIF-1 α degradation via immunoblot, red box indicates stabilization of HIF-1 α , blue box indicates baseline (untreated) HIF-1 α levels. Bottom panel shows anti-histone-H3 (H3) blot as loading control.

			Concentra	tion dosed		
		1000 µM	500 µM	250 µM	125 µM	
Compound	2-KG	105.7	115.7	111.3	105.9	
		95.6	101.2	110.4	101.3	
		111.9	109.4	104.6	102.6	
	1	4.9	222.7	328.3	267.2	
		5.5	229.7	294.8	230.9	(Re
		7.4	259.9	319.6	252.4	lat
	2	119.1	119.3	101	101.5	tixe P
		133.4	106.1	106	103.5	erc e tc
		121.5	95.8	103.3	102.9	u en
	3	41.4	581.4	420.2	127.3	
		43.9	572.5	479.6	139.3	eat
		41.4	580.3	451.8	106.9	ed -1 c
	octyl-KG	26.4	159.7	161.4	128.2	8
		19.1	166.8	163.4	139.1	ntr
		13.8	155.6	157.3	120.6	ol)
	dimethyl-KG	118.3	120.3	119	108.2	
		123.2	114.4	117.7	110.1	Color Key
		120.3	106.5	115.8	114.6	HIF degraded
	TFMB-OH	144.9	102.3	93.2	88	HIF unaffected
		141.8	113.5	96.6	89.3	HIE stabilized
		139.3	107.1	86.6	87.1	

Figure S2. Concentration-dependent effects of 2-KG esters on HIF-luciferase activity. Values represent the percent HIF-luciferase activity relative to an untreated control. Values below 100% indicate less HIF-luciferase activity (anti-hypoxic, blue) values higher than 100% indicate greater HIF-luciferase activity (pro-hypoxic, red).



Figure S3. TFMB alcohol product produced by ester cleavage is a dose-dependent stabilizer of cellular HIFluciferase activity. Note the yaxis *begins* at 100% activity.

General Synthetic Procedures

Chemicals were purchased from commercial sources (Sigma-Aldrich, Alfa Aesar, and TCI America) and used without further purification unless otherwise noted. Nbromosuccinimide was recrystallized in water and vacuum dried immediately before use. Concentration of *n*-BuLi in hexanes was determined by titrating the reagent against diphenylacetic acid in anhydrous THF. Anhydrous solvents were prepared by passage over activated alumina. Thin-layer chromatography (TLC) was conducted with E. Merck silica gel 60 F254 precoated plates (0.25 mm) and visualized by exposure to UV light (254 nm) or chemical staining. Flash column chromatography was performed using normal phase or reverse phase on a CombiFlash® Rf 200i (Teledyne Isco Inc). ¹H NMR spectra were recorded at 400 or 500 MHz, and are reported relative to deuterated solvent signals. Data for ¹H NMR spectra are reported as follows: chemical shift (δ ppm), multiplicity, coupling constant (Hz), and integration. ¹³C NMR spectra were recorded at 100 or 125 MHz. Data for ¹³C NMR spectra are reported in terms of chemical shift. All NMR spectra were standardized to the NMR solvent signal as reported by Gottlieb.¹ Analytical LC/MS was performed using a Shimadzu LCMS-2020 Single Quadrupole utilizing a Kinetex 2.6 µm C18 100 Å (2.1 x 50 mm) column obtained from Phenomenex Inc. Runs employed a gradient of $0 \rightarrow 90\%$ MeCN/0.1% agueous formic acid over 4 minutes at a flow rate of 0.2 mL/min. High-resolution LC/MS analyses were conducted on a Thermo-Fisher LTQ-Orbitrap-XL hybrid mass spectrometer system with an Ion MAX API electrospray ion source in positive or negative ion mode. Separations were carried out on a narrow-bore (50 X 2.1 mm), Zorbax Rapid-Resolution, reversed-phase C18 (3.5 μ m) column with a flow rate of 250 μ L/min with a 10 min, 2-90% gradient of MeCN/H₂O containing 0.1% HCOOH.

Synthesis of 2-KG Esters

(a) 3-(trifluoromethyl)benzyl 1,3-dithiane-2-carboxylate (4)



1,3-dithiane-2-carboxylic acid (1.64 g. 10.0 mmol), 3-trifluoromethylbenzyl alcohol (1.60 g, 9.1 mmol), and Oxyma Pure (1.44 g, 10.1 mmol) were dissolved in DMF (50 mL) with stirring. This mixture was cooled to 0 °C and *N*,*N'*-dicyclohexylcarbodiimide (DCC) was added (2.30 g, 11.1 mmol) in a single portion. After 1 hour, the ice bath was removed and the reaction was allowed to warm to room temperature and stir overnight. The solvent was removed *in vacuo*, and the subsequent oil was re-dissolved in dichloromethane (50 mL) and washed with 5% NaHCO₃ (2 x 75 mL). The organic layer was dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography (0 \rightarrow 50% EtOAc:hexanes) to yield the product **4** as an off-white solid (2.28 g, 78%). ¹H NMR (400 MHz, CDCl₃) δ 7.68 (s, 1H), 7.61 (dd, *J* = 12.4, 7.7 Hz, 2H), 7.52 (t, *J* = 7.6 Hz, 1H), 5.29 (s, 2H), 3.48-3.36 (m,

2H), 2.68-2.57 (m, 2H), 2.24-1.98 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 169.42, 136.46, 131.14, 129.15, 125.18, 125.15, 124.63, 124.60, 66.21, 39.57, 25.81, 24.92. HRMS (ESI+): Calcd for C₁₃H₁₄F₃O₂S₂: 322.0309. Measured: 323.0382

(b) Tert-butyl 1,3-dithiane-2-carboxylate (5)



Tert-butyl ester **5** was prepared by a method adapted from Wright and coworkers.² Briefly, concentrated H₂SO₄ (0.055 mL, 1.0 mmol) was added to vigorously stirring anhydrous MgSO₄ (0.481 g, 4.0 mmol), in dichloromethane (4 mL). The reaction mixture was stirred for 15 minutes at room temperature, followed by sequential addition of 1,3dithiane-2-carboxylic acid (164 mg. 1.0 mmol) and tert-butyl alcohol (0.48 mL, 5.0 mmol). The reaction mixture was stirred for 18 hours, quenched with 7.5 mL of 5% NaHCO₃, and stirred until homogenous. The organic phase was washed with brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The product was purified by flash chromatography (0 \rightarrow 50% EtOAc:hexanes) to yield the product **5** as an light yellow solid (1.78 g, 81%). ¹H NMR (400 MHz, CDCl₃) δ 4.11 (s, 1H), 3.47-3.37 (m, 2H), 2.67-2.57 (m, 2H), 2.21-2.10 (m, 1H), 2.07-1.98 (m, 1H), 1.53 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 168.95, 82.31, 41.40, 27.94, 26.05, 25.16. HRMS (ESI+): Calcd for C₉H₁₆O₂S₂: 220.0592. Measured: 221.0664

(c) 3-(trifluoromethyl)benzyl acrylate (7)



Acrylic acid (0.43 mL, 6.27 mmol), 3-trifluoromethylbenzyl alcohol (1.24 g, 7.0 mmol), dimethyl aminopyridine (DMAP) (0.153 g, 1.25 mmol) and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (1.34 g, 7.0 mmol) were dissolved in DMF (7.5 mL), heated to 40 °C, and stirred overnight. The solvent was removed *in vacuo*, and the resultant oil was dissolved in dichloromethane (20 mL) and washed with 5% NaHCO₃ (2 x 10 mL The organic layer was dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography (0 \rightarrow 50% EtOAc:hexanes) to yield the product **7** as an oil (2.28 g, 78%). ¹H NMR (400 MHz, CDCl₃) δ 7.69-7.48 (m, 4H), 6.50 (dd, *J* = 17.4, 1.4 Hz, 1H), 6.21 (dd, *J* = 17.3, 10.4 Hz, 1H), 5.92 (dd, *J* = 10.5, 1.4 Hz, 1H), 5.27 (s, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 165.79, 136.87, 131.59, 131.37, 129.08, 127.96, 125.09, 125.05, 124.82, 124.78, 65.40.

(d) 3-(trifluoromethyl)benzyl 2-(3-(tert-butoxy)-3-oxopropyl)-1,3-dithiane-2carboxylate (8)



TFMB ester 4 (1.44 g, 4.5 mmol) was dissolved under argon in anhydrous diethyl ether (5 mL) and cooled to -78 °C. A solution of *n*-BuLi in hexanes (1.97 mL, 4.5 mmol) was quickly added and the reaction mixture was allowed to stir for 45 minutes at 0 °C. This solution was transferred via cannula to a cooled (-78 °C) suspension of Cul (0.456 g, 2.39 mmol) in ether,² which was allowed to stir for an additional hour. This mixture was warmed to -40 °C and a solution of tert-butyl acrylate 6 (0.22 mL, 1.50 mmol) in ether (8 mL) was added. The reaction was allowed to warm to room temperature over 2.5 hours, quenched with saturated NH₄CI (25 mL) for 15 minutes, and filtered through Celite. The mixture was extracted with ethyl acetate, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography $(0 \rightarrow 50\%$ EtOAc:hexanes) to yield the product 8 as an oil (0.342 g, 63%). ¹H NMR (400 MHz, CDCl₃) δ 7.68 (s, 1H), 7.61 (t, *J* = 7.3 Hz, 2H), 7.52 (t, *J* = 7.7 Hz, 1H), 5.30 (s, 2H), 3.28-3.16 (m, 2H), 2.73-2.61 (m, 2H), 2.54 - 2.46 (m, 2H), 2.42 -2.33 (m, 2H), 2.18-2.07 (m, 1H), 1.93-1.79 (m, 1H), 1.45 (s, 9H). ¹³C NMR (125 MHz, CDCl₃) δ 171.66, 170.22, 136.67, 131.09, 129.17, 125.15, 125.12, 124.54, 124.51, 80.66, 66.40, 51.61, 33.30, 30.73, 28.06, 27.64, 24.31. HRMS (ESI+):Calcd for C₂₀H₂₅F₃O₂S₂: 450.1146. Measured: 451.1219

(e) 5-tert-butyl 1-(3-(trifluoromethyl)benzyl) 2-oxopentanedioate (9)



Diester **8** (71.9 mg, 0.16 mmol) was dissolved in acetone (0.8 mL) and added dropwise to a stirring, ice-cold solution of *N*-bromosuccinimide (228 mg, 1.3 mmol) in 95:5 acetone:water (4 mL).³ Once the starting material had been consumed as judged by TLC analysis (typically ~5-15 minutes), the reaction was quenched with saturated sodium sulfite (5 mL) and 1:1 hexane:dichloromethane (5 mL). The solution was washed with 5% NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography (0 \rightarrow 50% EtOAc:hexanes) to yield the product **9** as an oil (36.7 mg, 64%). ¹H NMR (400 MHz, CDCl₃) δ 7.69 (s, 1H), 7.62 (t, *J* = 7.3 Hz, 2H), 7.53 (t, *J* = 7.7 Hz, 1H), 5.35 (s, 2H), 3.13

(t, J = 6.5 Hz, 2H), 2.63 (t, J = 6.5 Hz, 2H), 1.44 (s, 9H). ¹³C NMR (125 MHz, CDCl₃) δ 192.20, 171.12, 160.19, 135.39, 131.89, 129.30, 125.66, 125.63, 125.36, 81.19, 67.02, 34.32, 29.00, 28.00. HRMS (ESI+): Calcd for C₁₇H₁₉F₃O₅: 360.1185. Measured: 378.1525 (corresponds to hydrate), 383.1078 (M+Na⁺)

(f) 4,5-dioxo-5-((3-(trifluoromethyl)benzyl)oxy)pentatonic acid (1)



Selective removal of the tert-butyl ester was accomplished following a method adapted from Mehta and coworkers.⁴ Briefly, diester **9** (21.3 mg, 0.059 mmol) was dissolved in dichloromethane (0.5 mL), and a mixture of TFA (0.77 mmol) and triethylsilane (0.15 mmol) were directly added. The reaction was stirred for 15 minutes at room temperature, and directly loaded onto a disposable Combi*Flash* FR 200i column for flash chromatography purification using a $0 \rightarrow 10\%$ CH₂Cl₂:MeOH gradient. Concentration of eluted product under reduced pressure results in isolation of a mixture of the ketone and hydrate form of **1** (15.5 mg, 92%; estimated at ~60% ketone by ¹H-NMR). Note: ¹H NMR and¹³C NMR values for both the ketone and hydrate are reported. ¹H NMR (500 MHz, CDCl₃) δ 7.72-7.51 (m, 8H), 5.42-5.31 (m, 4H), 4.67 (s, 1H), 3.21 (t, *J* = 6.4 Hz, 2H), 2.90-2.70 (m, 6H), 2.40- 2.34 (m, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 191.58, 175.24, 174.21, 168.22, 159.99, 135.25, 134.92, 131.89, 131.71, 131.49, 131.35, 131.23, 129.54, 129.34, 125.96 (q, *J* = 3.8 Hz), 125.72 (q, *J* = 3.8 Hz), 125.38 (q, *J* = 5Hz), 125.23 (q, *J* = 3.8 Hz), 125.20, 125.17, 124.81, 101.17, 68.13, 67.16, 34.01, 30.66, 28.20, 26.91. HRMS (ESI-): Calcd for C₁₃H₁₁F₃O₅: 304.0559. Measured: 303.0485

(g) Tert-butyl 2-(3-oxo-3-((3-(trifluoromethyl)benzyl)oxy)propyl)-1,3-dithiane-2carboxylate (10)



Tert-butyl ester **5** (232 mg, 1.1 mmol) was dissolved under argon in anhydrous diethyl ether (1.3 mL) and cooled to -78 °C. A solution of *n*-BuLi in hexanes (0.45 mL, 1.1 mmol) was quickly added and the reaction mixture was allowed to stir for 45 minutes at 0 °C. This solution was transferred via cannula to a cooled (-78 °C) suspension of Cul (107 mg, 0.56 mmol) in ether (2.5 mL),² which was allowed to stir for an additional hour. This

mixture was warmed to -40 °C and a solution of TFMB acrylate **7** (81.1 mg, 0.35 mmol) in ether was added. The reaction was allowed to warm to room temperature over 2.5 hours, quenched with saturated NH₄Cl (5 mL) for 15 minutes, and filtered through Celite. The mixture was extracted with ethyl acetate, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography (0 \rightarrow 50% EtOAc:hexanes) to yield the product **10** as an oil (0.107 g, 68%). ¹H NMR (400 MHz, CDCl₃) δ 7.67-7.46 (m, 4H), 5.19 (s, 2H), 3.37-3.25 (m, 2H), 2.75-2.61 (m, 4H), 2.42-2.31 (m, 2H), 2.20-2.09 (m, 1H), 1.93-1.80 (m, 1H), 1.53 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 172.39, 169.46, 136.91, 131.37, 129.05, 125.02, 124.98, 124.82, 124.78, 82.64, 65.48, 51.95, 33.25, 29.66, 27.95, 27.55, 24.49. HRMS (ESI+): Calcd for C₂₀H₂₅F₃O₄S₂: 450.1146. Measured: 451.1221



Diester **10** (207 mg, 0.46 mmol) was dissolved in acetone (2.5 mL) and added dropwise to a stirring, ice-cold solution of *N*-bromosuccinimide (655 mg, 3.7 mmol) in 95:5 acetone:water (12 mL).³ Once the starting material had been consumed as judged by TLC analysis (typically ~5-15 minutes), the reaction was quenched with saturated sodium sulfite (5 mL) and 1:1 hexane:dichloromethane (5 mL). The solution was washed with 5% NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography (0→40% EtOAc:hexanes) to yield the product **9** as an oil (154 mg, 93%). ¹H NMR (500 MHz, CDCl₃) δ 7.64-7.50 (m, 4H), 5.20 (s, 2H), 3.17 (t, *J* = 6.5, 6.5 Hz, 2H), 2.74 (t, *J* = 6.5, 6.5 Hz, 2H), 1.57 (s, 9H). ¹³C NMR (125 MHz, CDCl₃) δ 193.46, 171.83, 159.79, 136.69, 131.36, 129.11, 125.11, 125.08, 124.81, 124.78, 84.26, 65.74, 34.00, 27.78, 27.54. HRMS (ESI+): Calcd for C₁₇H₁₉F₃O₅: 360.1185. Measured: 378.1525 (corresponds to hydrate), 383.1078 (M+Na⁺)





Diester **11** (68.6 mg, 0.19 mmol) was dissolved in dichloromethane (0.5 mL), and a mixture of TFA (0.19 mL, 2.49 mmol) and triethylsilane (0.076 mL, 0.48 mmol) were directly added.⁴ The reaction was stirred for 30 minutes, concentrated under reduced pressure, and purified by flash chromatography (0 \rightarrow 10% CH₂Cl₂:MeOH) to yield **2** as an oil (51.4 mg, 89%). ¹H NMR (400 MHz, CDCl₃) δ 7.54 (s, 2H), 7.48-7.38 (m, 2H), 5.11 (s, 2H), 3.21 (t, *J* = 6.8, 5.8 Hz, 2H), 2.74 (t, *J* = 6.8, 5.8 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 194.16, 171.50, 158.48, 136.42, 131.41, 129.16, 125.25, 125.22, 124.87,

124.84, 66.01, 32.19, 27.82. HRMS (ESI+): Calcd for $C_{13}H_{11}F_3O_5$: 304.0559. Measured: 349.0270 (M+2Na⁺)

(j) 3-(trifluoromethyl)benzyl 2-(3-oxo-3-((3-(trifluoromethyl)benzyl)oxy)propyl)-1,3-dithiane-2-carboxylate (12)



TFMB ester 4 (670 g, 2.1 mmol) was dissolved under argon in anhydrous diethyl ether (1 mL) and cooled to -78 °C. A solution of n-BuLi in hexanes (0.83 mL, 2.1 mmol) was quickly added and the reaction mixture was allowed to stir for 45 minutes at 0 °C. This solution was transferred via cannula to a cooled (-78 °C) suspension of Cul (211 mg, 1.1 mmol) in ether (5 mL),² which was allowed to stir for an additional hour. This mixture was warmed to -40 °C and a solution of TFMB acrylate 7 (160 mg, 0.7 mmol) in ether was added. The reaction was allowed to warm to room temperature over 2.5 hours, quenched with saturated NH₄CI (10 mL) for 15 minutes, and filtered through Celite. The mixture was extracted with ethyl acetate, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography $(0 \rightarrow 50\%$ EtOAc:hexanes) to yield the product **12** as an oil (0.182 g, 47%). ¹H NMR (400 MHz, CDCl₃) δ 7.70-7.46 (m, 8H), 5.29 (s, 2H), 5.18 (s, 2H), 3.24 (t, J = 16.0 Hz, 2H), 2.76-2.58 (m, 4H), 2.45 (t, J = 8.0 Hz, 2H), 2.19-2.05 (m, 1H), 1.93-1.76 (m, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 172.15, 170.12, 136.79, 136.56, 131.40, 131.10, 129.18, 129.06, 125.19, 125.16, 125.06, 125.03, 124.85, 124.82, 124.55, 124.52, 66.45, 65.56, 51.26, 32.99, 29.50, 27.58, 24.17. HRMS (ESI+): Calcd for C₂₄H₂₂F₆O₄S₂: 552.0864. Measured: 553.0930

(k) Bis(3-(trifluoromethyl)benzyl) 2-oxopentanedioate (3)



Diester **12** (55.5 mg, 0.10 mmol) was dissolved in acetone (0.5 mL) and added dropwise to a stirring, ice-cold solution of *N*-bromosuccinimide (142 mg, 0.8 mmol) in 95:5 acetone:water (2.5 mL).³ Once the starting material had been consumed as judged by TLC analysis (typically ~5-10 minutes), the reaction was diluted with saturated sodium sulfite (5 mL) and 1:1 hexane:dichloromethane (5 mL). The solution was washed with 5% NaHCO₃, water, and brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resultant oil was purified by flash chromatography (0 \rightarrow 50%

EtOAc:hexanes) to yield the product **3** as an oil (28.2 mg, 68%). ¹H NMR (400 MHz, CDCl₃) δ 7.70-7.48 (m, 8H), 5.35 (s, 2H), 5.19 (s, 2H), 3.23 (t, *J* = 6.4 Hz, 2H), 2.78 (t, *J* = 6.4 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 191.78, 171.64, 160.03, 136.57, 135.27, 131.87, 131.38, 129.32, 129.12, 125.70, 125.67, 125.36, 125.33, 125.16, 125.12, 124.83, 124.80, 122.72, 67.13, 65.85, 34.16, 27.60. HRMS (ESI+): Calcd for $C_{24}H_{22}F_6O_4S_2$: 462.0902. Measured: 463.0976

NMR Spectra for Final Products and Intermediates













































General cell culture materials and methods

HeLa cells containing a stable transfection of HIF-1 α oxygen dependent-degradation domain (HIF-1 α ODD) fused to luciferase (Luc) were obtained from Tsui-Fen Chou (UCLA).^{6,7} Cells were grown in DMEM supplemented with 2.5% FBS, 2 mM glutamine at 37 °C and 5% CO₂. 2.5 µg/mL puromycin was added to media to maintain HIF-Luc selection expression. For all luciferase experiments, cells were plated on sterile, white-walled 96-well clear bottom plates (Corning #3610) luciferase activity detected using D-luciferin (Pierce # 88291). SDS-PAGE and Western blotting was performed using Bis-Tris NuPAGE gels (4-12%), MES running buffer in Xcell SureLock MiniCells (Invitrogen), and anti-HIF1 α antibody (Cell Signaling # 3716) according to the manufacturer's instructions.

Cell-based assay for HIF-1 α -degradation

To assay 2-KG dependent changes in HIF-1 α levels, cells were seeded on 96-well plates (100 μ L, 10,000-20,000 cells/well) and allowed to adhere for 16 hours. Cells were dosed with vehicle or test compounds (50 μ l of a 2-4 mM stock in DMEM) and returned to the incubator. After 2 hours, cells were removed from the incubator, inspected for any signs of toxicity by microscopy, and D-luciferin (50 μ l of 1 mg/ml in PBS) was added using a multichannel pipette. After five minutes, luminescense intensity was quantitated on a Biotek Synergy 2 plate reader (Biotek) using a 0.1-ms integration time.

Ketoglutarate Ester Stablility Assay

The relative stabilities of 1-TFMB-KG (1) and 5-TFMB-KG (2) were determined in phosphate buffered saline (10 mM Na₂HPO₄, 1.8 mM KH₂PO₄ [pH 7.4] 137 mM NaCl, 2.7 mM KCl). Esters (2 mM) were incubated in PBS for 1 hour prior to analysis using a Biovision α -KG Assay (#K677-100) according to the manufacturer's instructions. The assay is based on enzymatic transamination of 2-KG to glutamate, which yields an equimolar amount of pyruvate, which is then consumed pyruvate oxidase to produce an equivalent of H₂O₂ and activate an oxidation-sensitive fluorophore (λ_{ex} = 535 nm, λ_{em} = 587 nm). A 2 mM solution of 2-ketoglutaric acid in PBS was used as a positive control representing 100% hydrolysis. Reported values represent the average of three measurements.

Treatment and Harvesting of Nuclear Extracts for HIF-1α Western Blot

Western blotting was used to verify the effects of 2-KG esters on HIF-1 α levels in HeLa cells. Briefly, HeLa cells were seeded on 6-well plates (2.5 mL, 10⁶ cells/well), allowed to adhere overnight (16 hrs) and treated with either vehicle, CoCl₂ (0.5 mM; positive control for HIF-1 α stabilization) or 2-KG diester **3** (0.5 mM, 1 mM). After 2 hours, cells were washed with ice cold PBS, harvested, and centrifuged (1400g, 5 min, 4°C). Supernatant (PBS) was removed and nuclear protein fractions were isolated using Thermo Scientific NE-PER kit (#PI-78835) according to the manufacturer's instructions. Protein concentrations were measured on a Qubit 2.0 fluorometer, and 15 µg of nuclear extract per well was found to give sufficient signal for detection of HIF1 α stabilization or degradation. Western blotting materials and antibodies are given in "General cell culture materials and methods" (above).

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