SUPPLEMENTAL MATERIAL

Table S1 Isolation data

Species	Strain	Isolation site	$Treatment^{\dagger}$	Treated abroad †
Pseudomonas aeruginosa	CCUG 70744	Sputum	Meropenem	-
Escherichia coli	CCUG 70745	Rectum	-	-
Klebsiella pneumoniae	CCUG 70742	Urine	-	-
Proteus mirabilis	CCUG 70746	Rectum	-	Balkan
Acinetobacter baumannii	CCUG 70743	Wound	-	Middle east
Klebsiella pneumoniae	CCUG 70747	Wound	Cefotaxime	Balkan

⁺ No information marked with dash (-).

Table S2 Intrinsic resistance genes in the reference genomes excluded from further analysis

Strain	Gene	Function	Accession number
P. aeruginosa PAO1	aph(3')-IIb	Aminoglycoside phosphotransferase	X90856
	<i>bla</i> oxa-50-like	β-lactamase	AY306130
	Ыа _{РАО}	β-lactamase	FJ666065
	catB7	Chloramphenicol acetyltransferase	AF036933
	fosA	Enzymatic fosfomycin resistance	NZ_ACWU01000146
K. pneumoniae	<i>Ыа</i> зну-11	β-lactamase	EF035558
MGH 78578	fosA	Enzymatic fosfomycin resistance	NZ_AFBO01000747
	oqxAB	Quinolone efflux pump	EU370913
P. mirabilis HI4320	cat	Chloramphenicol acetyltransferase	M11587
	tet(J)	Tetracycline efflux pump	ACLE01000065
A. baumannii ATCC	bla _{ADC-25}	β-lactamase	EF016355
17978	<i>Ыа</i> оха-66	β-lactamase	FJ360530



Fig. S1 Comparison between experimentally generated optical DNA maps (red) and theoretical optical DNA maps based on assembled sequences (teal) for non-novel plasmids (**a**) pKpn70747_2 in *K. pneumoniae* CCUG 70747 (118 kb), (**b**) pEco70745_2 in *E. coli* CCUG 70745 (44 kb), and (**c**) pKpn70742_2 in *K. pneumoniae* CCUG 70742 (63 kb). Intensity (y-axis) refers to the normalized emission intensity along the DNA molecule (x-axis) which correlates coarsely with the underlying DNA sequence. (**d**) Histogram displaying the size distribution of individual plasmid molecules found in *P. mirabilis* CCUG 70746, demonstrating a high variability of the plasmid within the sample.

		-	
Reference strain	Function	Gene	Locus tag
P. aeruginosa	Quinolone targets	gyrA	PA3168
PAO1		gyrB	PA0004
		parC	PA4964
		parE	PA4967
	β-lactamase	ampC	PA4110
	Penicillin-hinding	dacB	ΡΔ3047
	nrotein	dacC	DV3000
	protein	ftcl	DA4419
		j LSI mrcP	PA4418
		nhnA	PA4700
		рора	PA4003
		рорс	PAZZ/Z
		pbpG	PA0869
		ponA	PA5045
	Drug efflux pumps	mexA	PA0425
		техВ	PA0426
		mexC	PA4599
		mexD	PA4598
		mexE	PA2493
		mexF	PA2494
		mexX	PA2019
		meyV	DA2018
		opri	FA2018
		opri	PA4597
		oprivi	PA0427
		oprN	PA2495
	Porin	oprD	PA0958
	Regulatory proteins	ampD	PA4522
		ampR	PA4109
		czcR	PA2523
		czcS	PA2524
		mexR	PA0424
		mexT	PA2492
		mex7	PA2020
		nalC	PA3721
		nalD	
		nuiD	PA3574
	<u> </u>	пјхв	PA4600
E. coli str. K-12	Quinolone targets	gyrA	b2231
substr. MG1655		gyrB	b3699
		parC	b3019
		parE	b3030
K. pneumoniae	Quinolone targets	gyrA	KPN_02640
MGH 78578		gyrB	KPN 03437
		parC	KPN 03444
		parE	KPN 04102
	Drug efflux numps	acrA	KPN 00444
	Brag ernax partips	acrB	KPN 00443
	Regulatory protoins	acrD	
	Regulatory proteins	uci n	KPN_00443
		murA	KPN_01624
		marĸ	KPN_01625
		ramA	KPN_00556
		ramR	KPN_00555
		soxR	KPN_04463
		soxS	KPN_04462
	Outer membrane protein	tolC	KPN_03449
P. mirabilis	Quinolone targets	qyrA	HMPREF0693 1372
HI4320	0	avrB	HMPREF0693 0114
		parC	HMPREF0693 2557
		pare	HMPREF0693 2558
A baumannii	Quipolopo targato		A15 2626
		gyrA	A15_2020
ATCC 1/9/8		gyrв	A15_0004
		parC	A15_0194
		parE	A1S_3359

Table S3 Chromosomal genes screened for mutations conferring resistance

Antibiotic class	Antibiotic	P. aeruginosa		E. coli		K. pneumoniae		P. mirabilis		A. baumannii		K. pneumoniae	
		CCUG 70744*		CCUG 707	745*	CCUG 70742*		CCUG 707466*		CCUG 70743*		CCUG 70747*	
Carbapenems	Ertapenem	>32	iR	>32	R	4	R	>32	R	>32	iR	>32	R
	Imipenem	>32	R	>32	R	0.5	S	>32	R	>32	R	>32	R
	Meropenem	>32	R	>32	R	0.5	S	32	R	>32	R	>32	R
Cephalosporins	Cefuroxime	>256	iR	>256	R	>256	R	>256	R	>256	iR	>256	R
	Cefotaxime	>32	iR	>32	R	>32	R	>32	R	>32	iR	>256	R
	Ceftazidime	256	R	>256	R	32	R	>256	R	>256	-	>256	R
	Ceftriaxone	>32	iR	>32	R	>32	R	>32	R	>32	iR	>32	R
Monobactams	Aztreonam	96 1		>256	R	64	R	2	Ι	>256	-	>256	R
Penicillins	Amoxicillin	>256	iR	>256	R	>256	R	>256	R	>256	iR	>256	R
	Ampicillin	>256	iR	>256	R	>256	iR	>256	R	>256	iR	>256	iR
	Amoxicillin/clavulanic acid	>256/2	iR	>256/2	R	>256/2	R	>256/2	R	>256/2	iR	>256/2	R
	Piperacillin/Tazobactam	>256/4	R	>256/4	R	256/4	R	32/4	R	>256/4	-	>256/4	R
Aminoglycosides	Amikacin	16	I	16	Ι	6	S	>256	R	64	R	32	R
	Gentamicin	4	S	1	S	2	S	>256	R	64	R	2	S
	Tobramycin	1	S	16	R	16	R	>256	R	64	R	16	R
Fluoroquinolones	Ciprofloxacin	1	I	>32	R	>32	R	>32	R	>32	R	>32	R
	Levofloxacin	8	R	>32	R	2	I	32	R	>32	R	32	R
	Moxifloxacin	8	R	>32	R	4	R	32	R	>32	-	>32	R
Sulfonamides	Trimethoprim/sulfamethoxazole	>32/608	iR	>32/608	R	>32/608	R	>32/608	R	>32/608	R	>32/608	R
Tetracyclines	Tetracycline	64	iR	>256	R	192	R	32	iR	>256	-	4	R
	Tigecycline	16	iR	1	S	4	R	32	R	8	-	1	S
Others	Colistin	0.5	S	0.25	S	2	S	>256	iR	1	S	2	S
	Fosfomycin	>256	-	0.25	S	32	S	8	S	1024	iR	64	R
	Chloramphenicol	>256	iR	64	R	4	S	256	R	>256	-	16	R

Table S4 MICs and EUCAST clinical resistance classifications

*S = susceptible, I = intermediate, R = resistant, iR = intrinsically resistant. Units are mg I^{-1} .

Table S5.	Closest	matching	plasmids

Strain	Plasmid	Length	Closest match in GenBank	Length	Accession	Isolation	Alignment	Gaps	Same
		(bp)		(bp)	number	county	identity		ARGs
<i>E. coli</i> CCUG 70745	pEco70745_1	107 323	<i>Escherichia coli</i> isolate Co6114 plasmid pC06114_1	129 085	CP016035.1	Germany	67 %	19 %	No
	pEco70745_2	44 006	<i>Escherichia coli</i> strain OM26 plasmid pOM26-1	45 122	KP776609.1	Oman	98 %	3 %	Yes
	pEco70745_3	3 608	Salmonella enterica subsp. enterica serovar Typhimurium var. 5- str. CFSAN001921 plasmid unnamed	3 609	CP006052.1	USA	98 %	0 %	-
	pEco70745_4	1 934	<i>Escherichia coli</i> strain G3/10 plasmid pSYM3	1 934	KM107839.1	Germany	100 %	0 %	-
	pEco70745_5	1 780	<i>Escherichia coli</i> strain EC08 plasmid pEC08-1 RepA gene	1 980	JX238440.1	Sweden	93 %	0 %	-
	pEco70745_6	1 459	<i>Escherichia coli</i> strain ST648 plasmid pEC648_7	1 459	CP008721.1	China	100 %	0 %	-
<i>K. pneumoniae</i> CCUG 70742	pKpn70742_1	65 276	Klebsiella pneumoniae strain ATCC BAA- 2146 plasmid pHg	85 161	CP006662.2	USA	67 %	26 %	No
	pKpn70742_2	63 075	<i>Enterobacter cloacae</i> strain INSRA17313- 1 plasmid pUR17313-1	63 584	KP061858.1	Portugal	97 %	3 %	Yes
	pKpn70742_3	4 426	<i>Klebsiella pneumoniae</i> strain 0773 plasmid pKpn114, complete sequence	4 211	EU932690.1	China	96 %	7 %	-
	pKpn70742_4	1 780	<i>Escherichia coli strain</i> EC08 plasmid pEC08-1 RepA gene	1780	JX238440.1	Sweden	93 %	0 %	-
<i>P. mirabilis</i> CCUG 707466	pPmi70746_1	191 769	Salmonella enterica subsp. enterica serovar Corvallis plasmid pRH-1238	187 683	KR091911.1	Germany	78 %	20 %	No
<i>A. baumannii</i> CCUG 70743	pAba70743_1	10 880	<i>Acinetobacter baumannii</i> plasmid pAB120	10 879	JX069966.1	Lithuania	100 %	0 %	Yes
K. pneumoniae	pKpn70747 1	186 310	Klebsiella pneumoniae JM45 plasmid p1	317 154	CP006657.1	China	52 %	41 %	No
CCUG 70747	pKpn70747_2	117 859	<i>Klebsiella pneumoniae</i> strain Kpn-1870 plasmid pGR-1870	116 047	KF874498.1	Greece	89 %	7 %	Yes
	pKpn70747_3	32 773	Salmonella enterica subsp. enterica serovar Typhi linear plasmid pBSSB1	27 037	AM419040.1	Indonesia	73 %	21 %	-

Antibiotic resistance genes (ARGs).



Fig. S2 (a) Multiple sequence alignment of all phenicol resistance proteins in the ResFinder database and the *cat*-like protein encoded by the chromosome of *E. coli* CCUG 70745. The putative phenicol resistance gene (protein id PRJNA401330:CLH66_17905) is most similar to the group of CatB enzymes. The alignment was done using Clustal Omega and the figure depicts a neighbour joining tree using the BLOSUM62 distance matrix.

> PRJNA401330:CLH66_17905 MKIMKINNIDVMKENDIVFSNTAGFWSGPVEIECPAEIRQGVYQVDKIGAF SYMGGKSTHMLHIESIGRFCAIAGNVVAGAMEHPAHFLSPHPIMQGVFKWK ALDDFKLKNKDMLWKSRTFNNQINKDRFGKIKIGNDVWIGEGAFIRRGVKI GDGAIIASHSVVSKDVAPYSIVGGVPAKHIKFRFEDNIIEQLIEIKWWDYG LSALEGVDFTDINKAIKKIRENIDSGVAEKFTPNVIRINKDGVAEKI*

Fig. S2 (b) The predicted protein sequence of the putative novel phenicol inactivating enzyme in *E. coli* CCUG 70745.

AF462019_1	1	MNFFTSPFSGIPLDQQVTNPNIIVGKHSYYSGYYHGHSFDDCVRYLHPER	50
CLH66_17905	1	MKIMKINNIDVMKENDIVFSNTAGFWSGPVEIECPAEIRQGV	42
AF462019_1	51	DDVDKLVIGSFCSIGSGAVFMMAGNQGHRSDWI	83
CLH66_17905	43	<pre></pre>	89
AF462019_1	84	STFPGDTIIGHDV	115
CLH66_17905	90	.:: : : : : SPHPIMQGVFKWKALDDFKLKNKDMLWKSRTFNNQINKDRFGKIKIGNDV	139
AF462019_1 1	116	WIGTEAMIMPGVKIGHGAIIASRSVVTKDVAPYEVVGSNPAKHIKFRFSD	165
CLH66_17905 1	140	. .	189
AF462019_1 1	166	VEIAMLLEMAWWNWPESWLKESMQSLCSSDIEG	198
CLH66_17905 1	190	<pre></pre>	239
AF462019_1 1	199	LYLNWQSKART- 209	
CLH66_17905 2	240	:.: IRINKDGVAEKI 251	

Fig. S2 (c) A global protein alignment between the putative novel phenicol inactivating enzyme in *E. coli* CCUG 70745 (PRJNA401330:CLH66_17905) and the closest match in ResFinder, CatB9 (accession number AF62019) with 31.6% alignment identity, 42.6% similarity, and 24.7% gaps. Underlined region corresponds to the match against the conserved domain "Zenobiotic acyltransferase (XAT)" (see FIG. S2d).

		*	10 	*	20 	30 *) *	40 	*	50 I	60 * I	70 * I	80 * I)
CLH66_17905	48	IGAFS	YMGGK	STHML	HI-ESIG	RFCAIAG	NVVAGA	-ME HP AHI	FLSPHP:	IMQGV	FKWKALDD	F Klknk D MLI	√Ksrtfnnqi	125
Cdd:cd03349	4	VGDYS	YGSGP	DCDVG	GDKLSIG	KFCSIAP	GVKIGL	gGN HP TDI	NVSTYP	FYIFG	ge w eddak	FD D WP:	5 K	- 71
				90	100	1	10	120		130	14	a 1º	50	
			*	[,]	* .	*	.	*	*		*	····*····	*	
CLH66_17905	126	nkdr	fGKIK	IGNDVW	IGEGAF	IRRGVKIC	GDGAII	ASHSVVSK	(DVAPYS	SIVGG	/ PAK HIKF	RFEDNI IE QI	IEI KWWD Y	203
Cdd:cd03349	72		-GDVI	IGNDVW	IGHGAT	ILPGVTIC	DGA VI	AAGAVVTK	(DVPPY/	IVGG	PAKVIRY	RFDEETIERI	LAL KWWD W	144

FIG. S2 (d) Local alignment between the putative novel phenicol inactivating enzyme in *E. coli* CCUG 70745 (PRJNA401330:CLH66_17905) and the closest matching domain in the Conserved Domains Database (CDD): Zenobiotic acyltransferase (XAT) (Accession: cd03349; pssm-ID: 100040; domain length: 145; e-value: 6.30e-54). "The XAT class of hexapeptide acyltransferases is composed of a large number of microbial enzymes that catalyze the CoA-dependent acetylation of a variety of hydroxyl-bearing acceptors such as chloramphenicol and streptogramin, among others."