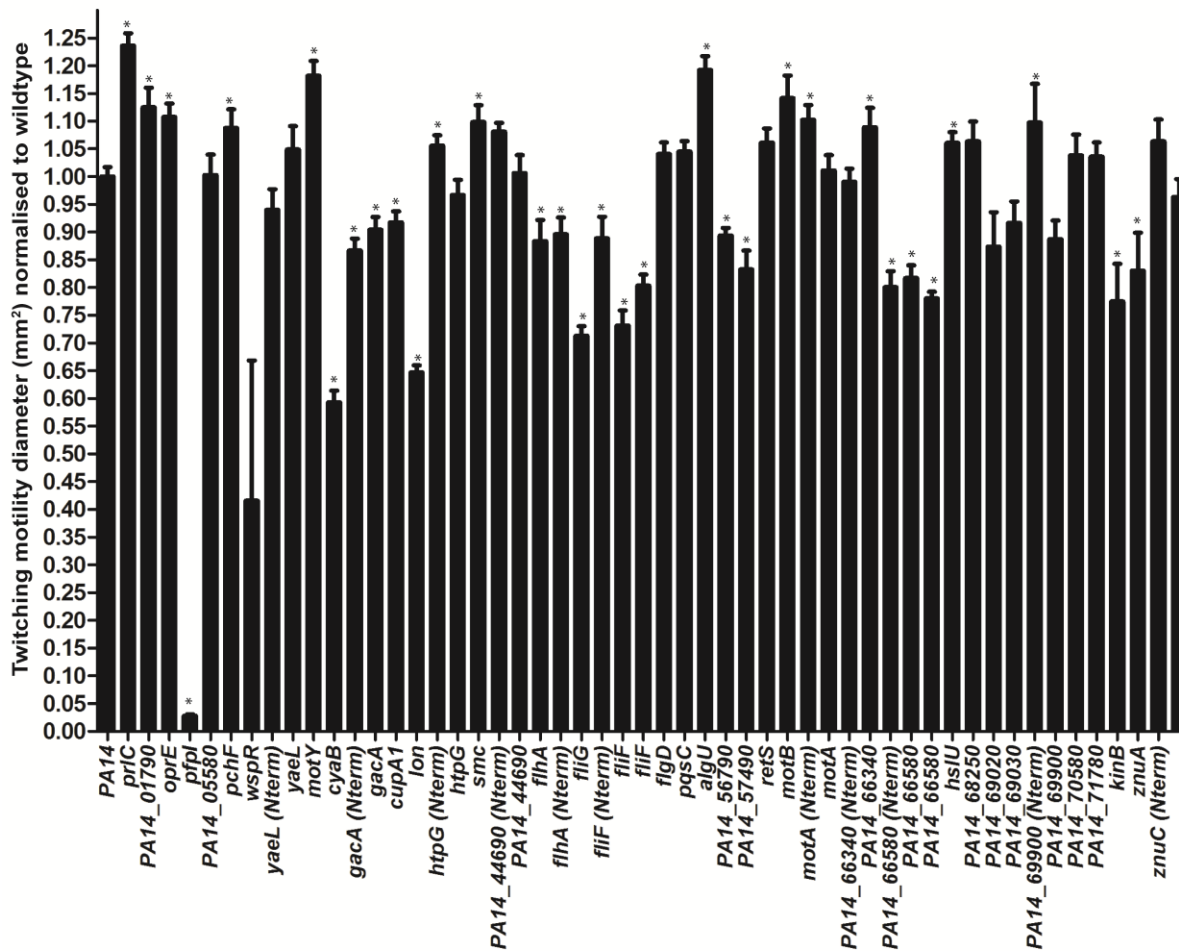
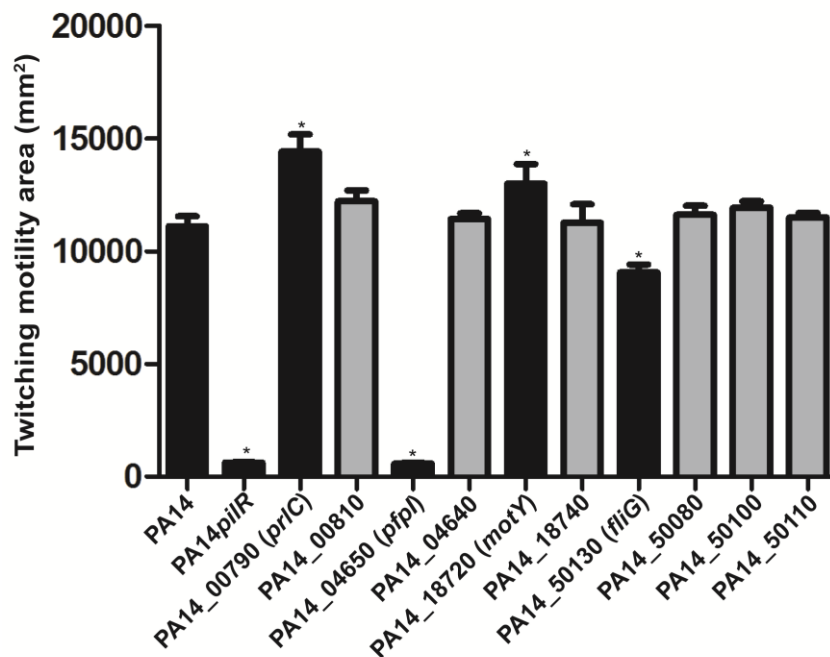


**Figure S1. (A)** The transposon mutant pool was inoculated in the centre of an inverted agar plate and incubated for 72 h at 37 °C. Mutants were separated based upon their ability to undergo twitching motility-mediated biofilm expansion away from the inoculation site. After 72 hrs mutants were harvested from the inner non-motile region (outlined in white) and the outer active twitching edge (outlined in black). Genomic DNA was extracted from each pool of mutants and sequenced using a mass parallel approach. **(B)** Growth rates in minimal media for 11 selected transposon mutants assayed for a submerged biofilm defect in Figure 1B. Growth rates were determined by incubation of transposon mutants at 37 °C for 19 h in M63 minimal media (same media used for submerged biofilm assays). There was no significant different between growth rates of transposon mutants compared to wildtype predicted by a one-way ANOVA with Dunnett's multiple comparison test. Data are represented as the mean  $\pm$  standard deviation for 2 independent replicates performed in triplicate.



**Figure S2.** Twitching motility of selected transposon mutant targets identified using TraDIS. Sub-surface twitching motility-mediated interstitial biofilm expansion at agar/plastic interface after 48 h incubation at 37 °C is presented as the average surface area in mm<sup>2</sup> ± standard deviation normalized against wildtype as obtained from 2 independent experiments performed in triplicate. Two-tailed student's t-test, \* (p<0.005) compared to wildtype.



**Figure S3.** Twitching motility of transposon mutants in genes downstream of *prlC*, *pfpl*, *fliG* and *motY*. Sub-surface twitching motility-mediated interstitial biofilm expansion at agar/plastic interface after 48 h incubation at 37 °C is presented as the mean surface area in mm<sup>2</sup> ± standard error of the mean as obtained from 2 independent experiments performed in triplicate. *prlC*, *pfpl*, *fliG* and *motY* are black bars and the transposon mutants in the operon downstream of each of these genes are grey bars. Two-tailed student's t-test, \* ( $p < 0.05$ ) compared to wildtype; ns for *PA14\_00810* (downstream of *prlC*) ( $p = 0.098$ ) compared to wildtype; ns for *PA14\_04640* (downstream of *pfpl*) ( $p = 0.546$ ) compared to wildtype; ns for *PA14\_50080* ( $p = 0.437$ ) or *PA14\_50100* ( $p = 0.381$ ) or *PA14\_50110* ( $p = 0.147$ ) (all downstream of *fliG*) compared to wildtype; and ns for *PA14\_18740* (downstream of *motY*) ( $p = 0.0828$ ) compared to wildtype.