

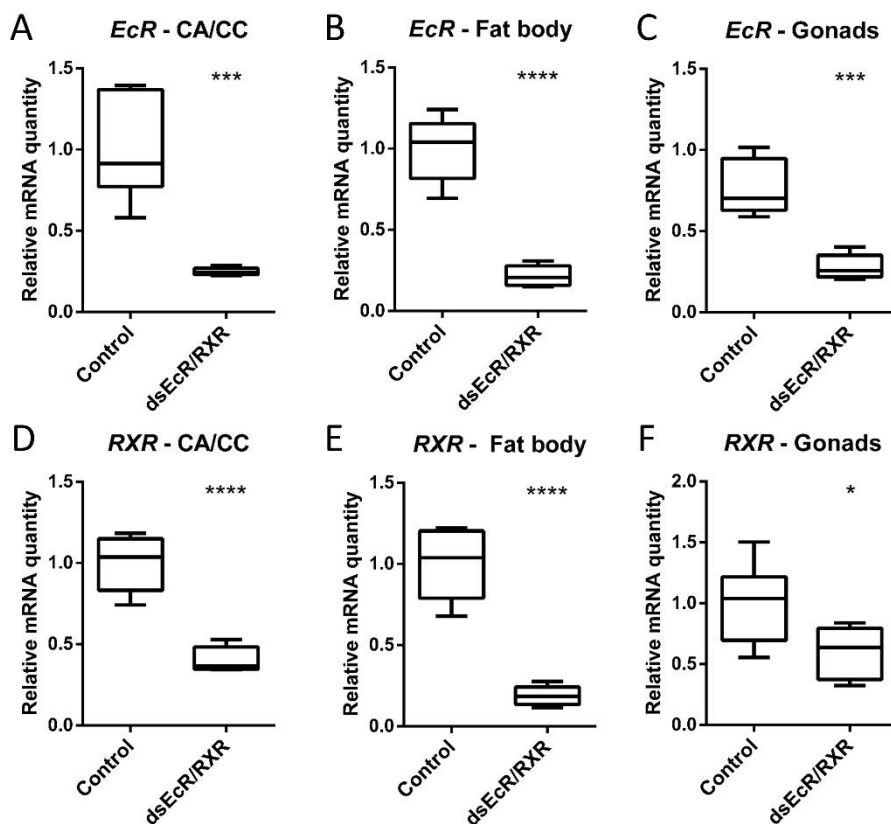
# The ecdysone receptor complex is essential for the reproductive success in the female desert locust, *Schistocerca gregaria*

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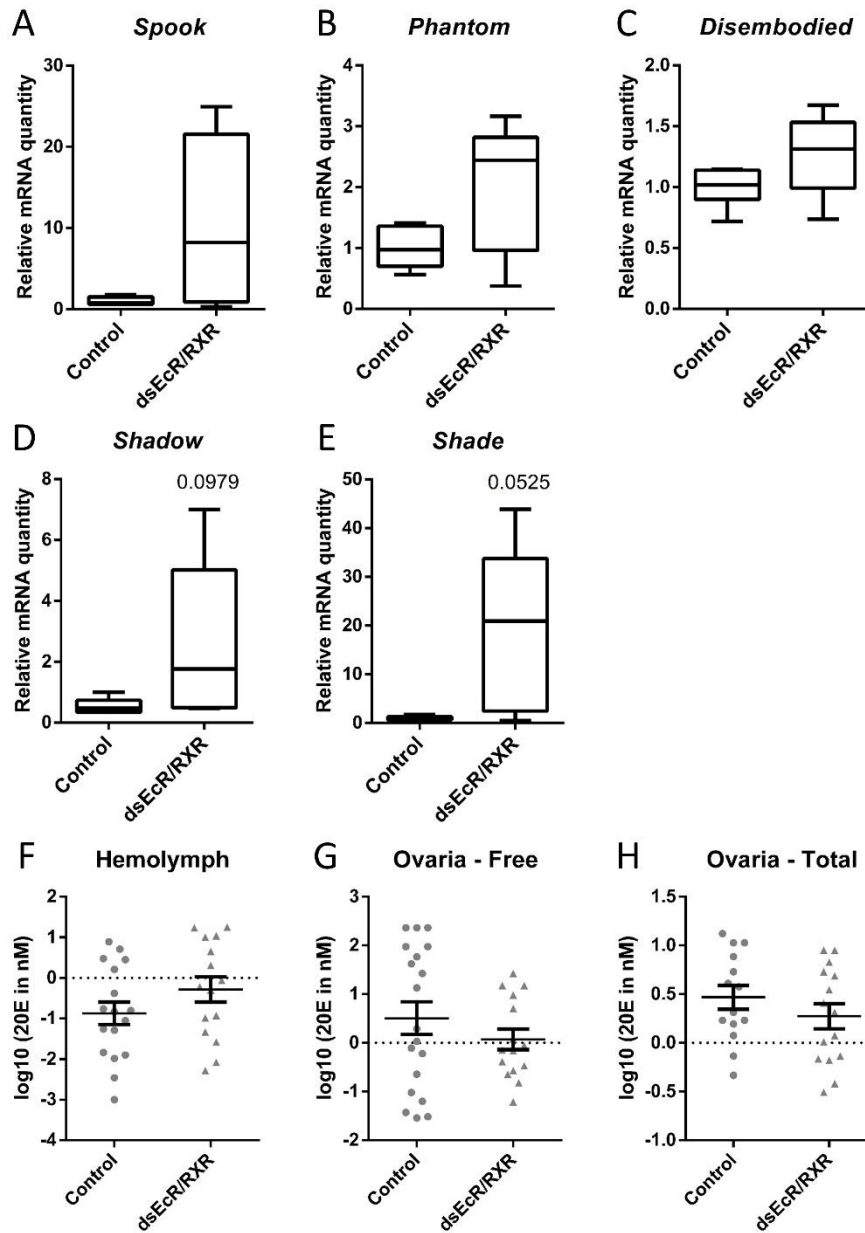
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## Supplementary figure 1



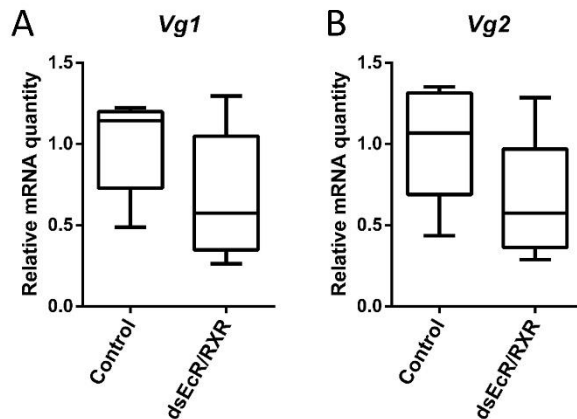
**Supplementary figure 1: Efficiency of the RNAi-mediated knockdown of the ecdysone receptor complex (*SchgrEcR/SchgrRXR*) in 12-day-old adult female *S. gregaria*.** Locusts were injected as described in materials and methods and dissected on day 12 of the adult stage. Relative transcript levels of (A-C) *SchgrEcR* and (D-F) *SchgrRXR* were measured in (A/D) the CA/CC complex, (B/E) the fat body and (C/F) the ovaries of 12-day-old female locusts, using qRT-PCR. The data represent box plots (min to max) of five independent pools of three locusts, run in duplicate and normalized to *CG13220* and  *$\alpha$ -tubulin1A* transcript levels for the ovaries, *CG13220*, *ubiquitin conjugating enzyme 10 (Ubi)* and *ribosomal protein 49 (RP49)* transcript levels for the fat body, and  *$\beta$ -actin* and *EF1 $\alpha$*  transcript levels for the CA/CC complex. Significant differences ( $p < 0.05$ ,  $p < 0.001$  and  $p < 0.0001$ ) are indicated by (an) asterisk(s) (\*, \*\*\* and \*\*\*\* respectively) (two-sided Welch's *t*-test on log-transformed data).

## Supplementary figure 2



**Supplementary figure 2: Effect of RNAi-mediated knockdown of the ecdysone receptor complex (*SchgrEcr/SchgrRXR*) on ecdysteroid synthesis in 12-day-old adult female *S. gregaria*.** Locusts were injected as described in materials and methods and dissected on day 12 of the adult stage. (A-E) Relative transcript levels of the *Halloween* genes, *SchgrSpo*, *SchgrPhm*, *SchgrDib*, *SchgrSad* and *SchgrShd* were measured in the ovaries of 12-day-old control and *dsEcr/RXR*-treated adult *S. gregaria*, using qRT-PCR. The data represent box plots (min to max) of five independent pools of three locusts, run in duplicate and normalized to *CG13220* and  *$\alpha$ -tubulin1A* transcript levels. (F-G) Ecdysteroid titres in the haemolymph and free and total ecdysteroid levels in the ovaries of 12-day-old control and *dsEcr/RXR*-treated female locusts. Ecdysteroid titres, expressed in ng 20E equivalents per  $\mu$ L haemolymph, and ecdysteroid levels, expressed in ng 20E equivalents per ovary, were measured with an EIA. The data represent mean  $\pm$  S.E.M. of individual animals ( $n = 15-19$ ), as well as the individual values (grey dots). No significant differences were observed.

### Supplementary figure 3



**Supplementary figure 3: Effect of RNAi-mediated knockdown of the ecdysone receptor complex (*SchgrEcR/SchgrRXR*) on vitellogenin synthesis in 12-day-old adult female *S. gregaria*.** Locusts were injected as described in materials and methods and dissected on day 12 of the adult stage. Relative transcript levels of *SchgrVg1* and *SchgrVg2* were measured in the fat body of 12-day-old control and dsEcR/RXR-treated adult *S. gregaria*, using qRT-PCR. The data represent box plots (min to max) of five independent pools of three locusts, run in duplicate and normalized to *CG13220*, *ubiquitin conjugating enzyme 10 (Ubi)* and *ribosomal protein 49 (RP49)* transcript levels.

## Supplementary table 1

Oligonucleotide sequences for primers used in qRT-PCR.

Reference genes	Forward primer	Reverse primer
<i>α-tubulin1A</i>	5'-TGACAATGAGGCCATCTATG-3'	5'-TGCTTCCATACCCAGGAATGA-3'
<i>CG13220</i>	5'-TGTTTCAGTTTTGGCTCTGTTCTGA-3'	5'-ACTGTTCTCCGGCAGAATGC-3'
<i>Ubi</i>	5'-GACTTTGAGGTGTGGCGTAG-3'	5'-GGATCACAACACAGAACGA-3'
<i>RP49</i>	5'-CGCTACAAGAAGCTTAAGAGGTCAT-3'	5'-CCTACGGCGCACTCTGTTG-3'
<i>β-actin</i>	5'-AATTACCATTGGTAACGAGCGATT-3'	5'-TGCTTCCATACCCAGGAATGA-3'
<i>EF1α</i>	5'-GATGCTCCAGGCCACAGAGA-3'	5'-TGCACAGTCGGCCTGTGAT-3'

Target genes	Forward primer	Reverse primer
<i>SchgrEcR</i>	5'-AAGGTTGATAATGCGGAATATGC-3'	5'-GTGATGGGCGCTCTGAAAAT-3'
<i>SchgrRXR</i>	5'-AATGCCTCGCTATGGGAATG-3'	5'-TCCTTTGTTTCGCTGCCTTC-3'
<i>SchgrE74</i>	5'-AAAGGAAGAGCCGTGAAG-3'	5'-CCCTGTTCTGCTCCACTTAAT-3'
<i>SchgrE75</i>	5'-GCACACCAACAGCATGATCT-3'	5'-ACCACCGAGCAGAACAGC-3'
<i>SchgrBr-C</i>	5'-CAGATCATCAGCACCACAATTACA-3'	5'-CGTGCCCGTTTGTTCGTT-3'
<i>SchgrHR3</i>	5'-CACCAGTACAGGCAATGA-3'	5'-TCAGTTGCTAGCTCAAGAC-3'
<i>SchgrFTZ-F1</i>	5'-CCTACTCTAGCCGATCACTTCAATG-3'	5'-CAAATGTAATCGCTGAGGTCAAAT-3'
<i>SchgrJHAMT</i>	5'-CGGAGCAAAGGCAAGCA-3'	5'-CCACTTACCCTGGTTT-3'
<i>SchgrCYP15a1</i>	5'-AAAGCAACTTCATCATTACAGATG-3'	5'-CAGAGCCAGCCATGAACAAA-3'
<i>SchgrMet</i>	5'-GGTGCCTGAAGAGGAAGAAA-3'	5'-ATGGAGGTGATGAAGGAGAAAAG-3'
<i>SchgrKr-h1</i>	5'-CTCCAAGACGTTTCATCCAGAG-3'	5'-TGCTTGGAGCAGGTGAAG-3'
<i>SchgrInR</i>	5'TGGATGGCACCAGAAAGCTT-3'	5'-CACAGCACTACCCGTAGCT-3'
<i>SchgrIRP</i>	5'-CCGTGGCAACTACAACCCAT-3'	5'-TCCGCGTCCGACACATCT-3'
<i>SchgrNP1</i>	5'-CGCACGGACACGTCACA-3'	5'-GTGACACGTAGGCGAAAGGAA-3'
<i>SchgrNP2</i>	5'-CGCTGGCGGTGACACA-3'	5'-CAAACCTCACTGTACAAAATAATAGACTAGA-3'
<i>SchgrNP3</i>	5'-GCGGAATCGGCATGGA-3'	5'-TCACAGAGCAACCGGAACATT-3'
<i>SchgrNP4</i>	5'-TGCGGACTCTCCAGTGCTT-3'	5'-TGACACATTCATTCTCTTCTGACA-3'
<i>SchgrVKR</i>	5'-GCATCTTGGCATTGATTTGCTA-3'	5'-GGAATCTCCATTTGTCAAGAGTT-3'
<i>SchgrSpo</i>	5'-CAACATCTTACCAGCTACATGTG-3'	5'-GGGTCGTCGTAGTCGAAGGA-3'
<i>SchgrPhm</i>	5'-CGCAGAGCCCGACAAC-3'	5'-CGAACATGTCGGCCATGA-3'
<i>SchgrDib</i>	5'-CCCAGGCTGCTATCGAGACT -3'	5'-CGACGACCAGGCTATGTAGTT -3'
<i>SchgrSad</i>	5'-ATCGTGGCCGAGATTACGAA -3'	5'-AGCACCATCTCCGGATCCT -3'
<i>SchgrShd</i>	5'-CCGCCGTCATTGACTTCATA-3'	5'-GTGAGCTCCAAGCGTGG-3'
<i>SchgrVg1</i>	5'-CCGCTGAACATCACTGCAAT-3'	5'-ACTTGGGCCAAATGGATGAG-3'
<i>SchgrVg2</i>	5'-GCTACCCGCAATCTGTAATAACA-3'	5'-CGACTGTGAAAGGGCATTGA-3'

Abbreviations: *Ubi* = ubiquitin conjugating enzyme 10, *RP49* = ribosomal protein 49, *EF1α* = elongation factor 1 alpha, *EcR* = ecdysone receptor, *RXR* = retinoid-X-receptor, *Br-C* = Broad-Complex, *FTZ-F1* = fushi tarazu factor 1, *JHAMT* = juvenile hormone acid methyltransferase, *CYP15a1* = methyl farnesoate epoxidase, *Kr-h1* = Krüppel-homolog 1, *Met* = methoprene-tolerant receptor, *InR* = insuline receptor, *IRP* = insuline related peptide, *NP* = neuroparsin, *VKR* = venus kinase receptor, *Spo* = spook, *Phm* = phantom, *Dib* = disembodied, *Sad* = shadow, *Shd* = shade, *Vg* = vitellogenin

## Supplementary table 2

Oligonucleotide sequences for primers used in dsRNA construct design. Underlined sequences are the T7 promoter sequences.

Target genes	Forward primer	Reverse primer
<i>SchgrEcR</i>	5'- <u>GAAATTAATACGACTCACTATAGGGCC</u> ACGTGAGGTTTCGGCACATC-3'	5'- <u>GAAATTAATACGACTCACTATAGGGCC</u> GTTTCCCCCATAACCAGCCAG-3'
<i>SchgrRXR</i>	5'- <u>GAAATTAATACGACTCACTATAGGGCC</u> GCTCAATGGGTCCACAGTCA-3'	5'- <u>GAAATTAATACGACTCACTATAGGGCC</u> ACACCATAATGCTTCCCGCT-3'
<i>GFP</i>	5'- <u>TAATACGACTCACTATAGGGAGA</u> AAGGTGATGCTACATACGGAA-3'	5'- <u>TAATACGACTCACTATAGGGAGA</u> ATCCAGCAGCAGTTACAAAC-3'

Abbreviations: *EcR* = ecdysone receptor, *RXR* = retinoid-X-receptor, *GFP* = green fluorescent protein