

SUPPLEMENTARY MATERIAL

Gene expression, proteome and calcium signaling alterations in immortalized hippocampal astrocytes from an Alzheimer's disease mouse model.

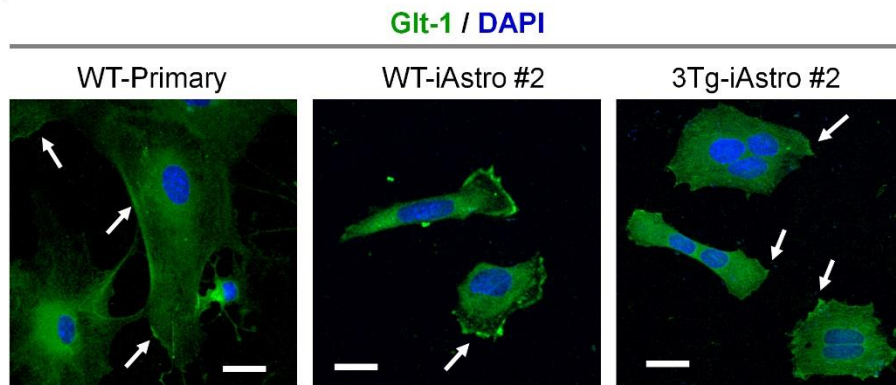
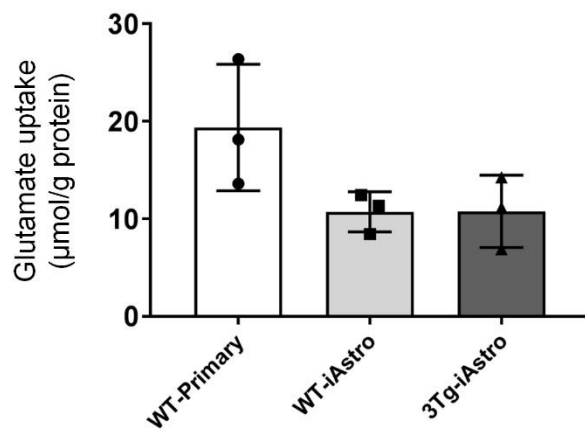
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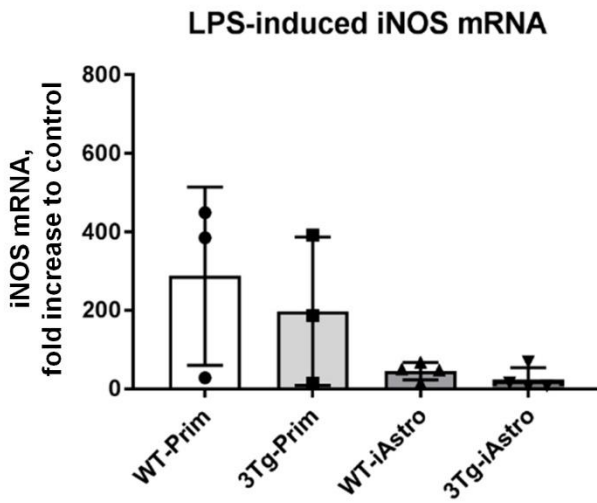
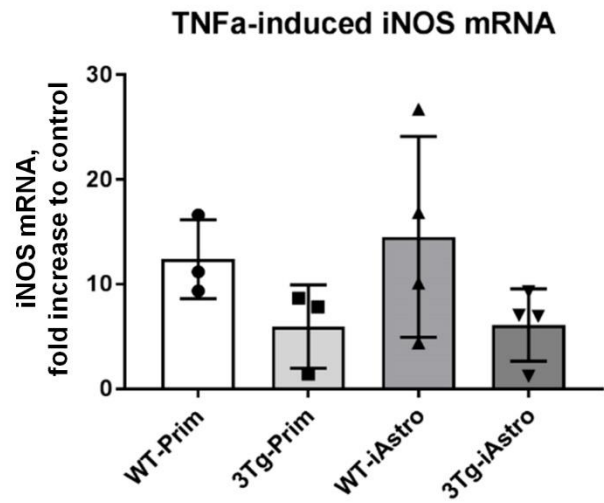
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Oligonucleotide primers used for quantitative real-time PCR.

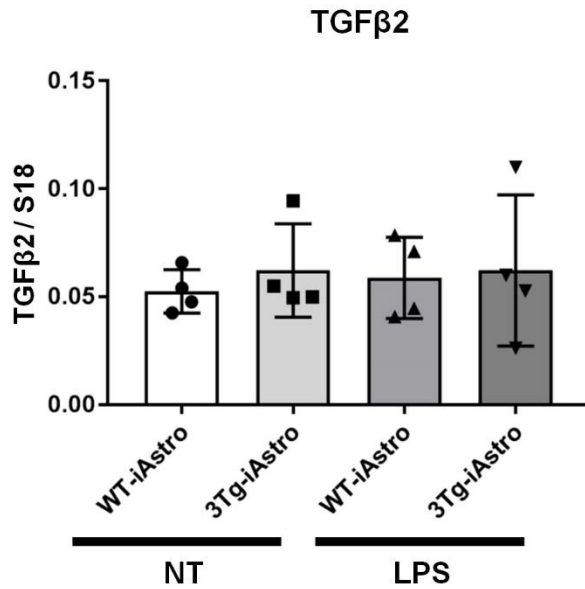
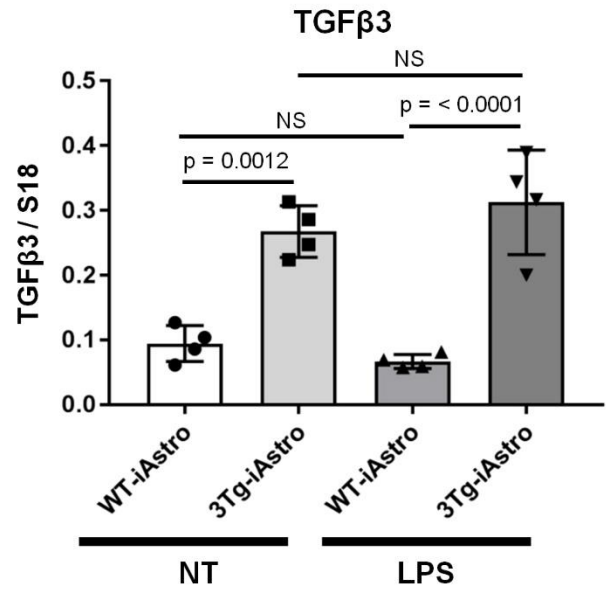
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Trim12a	NM_023835.2	Forward Reverse	TCATTGAAGAGGTGGCCCAG ACCGAACATTCTGCACATCT
Ccl27a (Pesky)	NM_001048179.1	Forward Reverse	TCTCCAACAAGCCAGAGACT GCTTGGGAGTGGCTGTCTAT
Car9	NM_139305.2	Forward Reverse	GGAGTCCCTTGGGTTAGAGG TGGGGCCAGAGTAGGGTG
Ptchd2	NM_001083342.1	Forward Reverse	CACAGCCTGCAGAACAATGT TTGGAGATGTACACGGTGCT
Slc2a4	NM_009204.2	Forward Reverse	TAAAACAAGATGCCGTCGGG CCAAACTGAAGGGAGCCAAG
P2ry2	NM_008773.3	Forward Reverse	ATCCTCACCACCTCAAGAGC GGCAACAGCACGTACTIONGAA
Panx1	NM_019482.2	Forward Reverse	TCACATGTATTGCCGTGGGT CTCGGGGAGAAGCAGCTTAT
Cacna2d3	NM_009785.1	Forward Reverse	TGCCTGTGAACATCAGTCTGA GCCATATGAGAGACGGATCCC
Fbln2	NM_007992.2	Forward Reverse	GGGCACTCACGATTGTAGCT GCAGGTCTGTACACACTCA
Col2a1	NM_001113515.2	Forward Reverse	GTGAGCCATGATCCGCCTC GGTTCTCCTTTCTGCCCTT
Padi2	NM_008812.2	Forward Reverse	GGGAAAATATGCTGCGGGAAC CACCTTCCGAGTGCTTCAGG
Pcdh17	NM_001013753.2	Forward Reverse	CTAATTCGAGCGAGACCCCT CTCTCAGGCTGGCTCTTTCT
Samd4	NM_001037221.2	Forward Reverse	TTGTGCCTCAGTATGACGGA AACCCTGAAGATGGCTGACA
Col4a6	NM_053185.2	Forward Reverse	CAAGCATGCACCCTGGATTG AGACTCCATGGCAATCTCGG
P2ry1	NM_008772	Forward Reverse	TCAAGCAGAATGGAGACACG CTCACTCAGGTGGCACACAC
iNOS	NM_010927.4	Forward Reverse	GCCACCAACAATGGCAACA CGTACCGGATGAGCTGTGAATT
Tgfb2	NM_009367.3	Forward Reverse	AGGCAGAGTTCAGGGTCTTC CCTTGCTATCGATGTAGCGC
Tgfb3	NM_009368.3	Forward Reverse	AGGAGACCTCGGAGTCTGAG CACTGAGGACACATTGAAACGA
S18	NM_213557	Forward Reverse	TGCGAGTACTCAACACCAACA CTGCTTTCCTCAACACCACA

A**B**

Supplementary Figure 1. iAstro express membrane-localized GLT-1 and are capable of glutamate uptake. **A**, membrane localization of GLT-1 in primary WT astrocytes and in WT-iAstro#2 and 3Tg-iAstro#2. The images are representative of 3 independent experiments. Similar staining was observed in other independently generated iAstro lines. Bar, 20 µm. **B**, Glutamate uptake was measured as described in Methods section in WT-primary and WT- and 3Tg-iAstro cells. The data are expressed as mean ± SD of experiments performed in triplicate for 3 independent cultures of WT primary astrocytes and WT-iAstro#2 and 3Tg-iAstro#2. ANOVA, followed by Tukey's post-hoc test was used for statistical analysis. The differences were not significant at $p < 0.05$.

A**B**

Supplementary Figure 2. LPS and TNF α -induced iNOS mRNA in WT- and 3Tg primary and immortalized astrocytes. Induction of iNOS upon treatment of primary and immortalized astrocytes from WT and 3xTg-AD mice with LPS (**A**) (100 ng/ml for 3h) or TNF α (**B**) (20 ng/ml 6h). Data are expressed as mean \pm SD of iNOS mRNA fold increase as compared to WT cells of 3 independent cultures for primary astrocytes and 4 independently generated iAstro lines (WT-iAstro#2,#3,#5 and #6, and 3Tg-iAstro#2,#3,#4 and #6). ANOVA, followed by Tukey's post-hoc test was used for statistical analysis. The differences were not significant at $p < 0.05$.

A**B**

Supplementary Figure 3. TGFβ2 and TGFβ3 mRNA levels in WT-iAstro and 3Tg-iAstro. TGFβ2 (A) and TGFβ3 (B) mRNA levels were measured by real-time PCR in control non treated (NT) and LPS-treated (100 ng/ml, 3h) WT-iAstro and 3Tg-iAstro lines. Data are expressed as mean ± SD of $\Delta C(t)$ TGFβ/S18 mRNA of 4 independently generated iAstro lines (WT-iAstro#2,#3,#5 and #6, and 3Tg-iAstro#2,#3,#4 and #6). ANOVA, followed by Tukey's post-hoc test was used for statistical analysis. Where not indicated, the differences were not significant at $p < 0.05$.