

*Electronic Supplementary Information (ESI)*

## **An enzyme-activatable probe liberating AIEgen: on-site sensing and long-term tracking of $\beta$ -galactosidase in ovarian cancer cells**

Kaizhi Gu,<sup>‡a</sup> Wanshan Qiu,<sup>‡b</sup> Zhiqian Guo,<sup>\*a,c</sup> Chenxu Yan,<sup>a</sup> Shiqin Zhu,<sup>a</sup> Defan Yao,<sup>a</sup> Ping Shi,<sup>c</sup> He Tian,<sup>a</sup> and Wei-Hong Zhu<sup>a</sup>

<sup>a</sup>Shanghai Key Laboratory of Functional Materials Chemistry, Key Laboratory for Advanced Materials and Institute of Fine Chemicals, Joint International Research Laboratory of Precision Chemistry and Molecular Engineering, Feringa Nobel Prize Scientist Joint Research Center, School of Chemistry and Molecular Engineering, East China University of Science & Technology, Shanghai 200237, China.

<sup>b</sup>Department of Cardiothoracic Surgery, Children's Hospital of Fudan University, Shanghai 201102, China.

<sup>c</sup>State Key Laboratory of Bioreactor Engineering, East China University of Science & Technology, 130 Meilong Road, Shanghai 200237, China.

*E-mail: guozq@ecust.edu.cn;*

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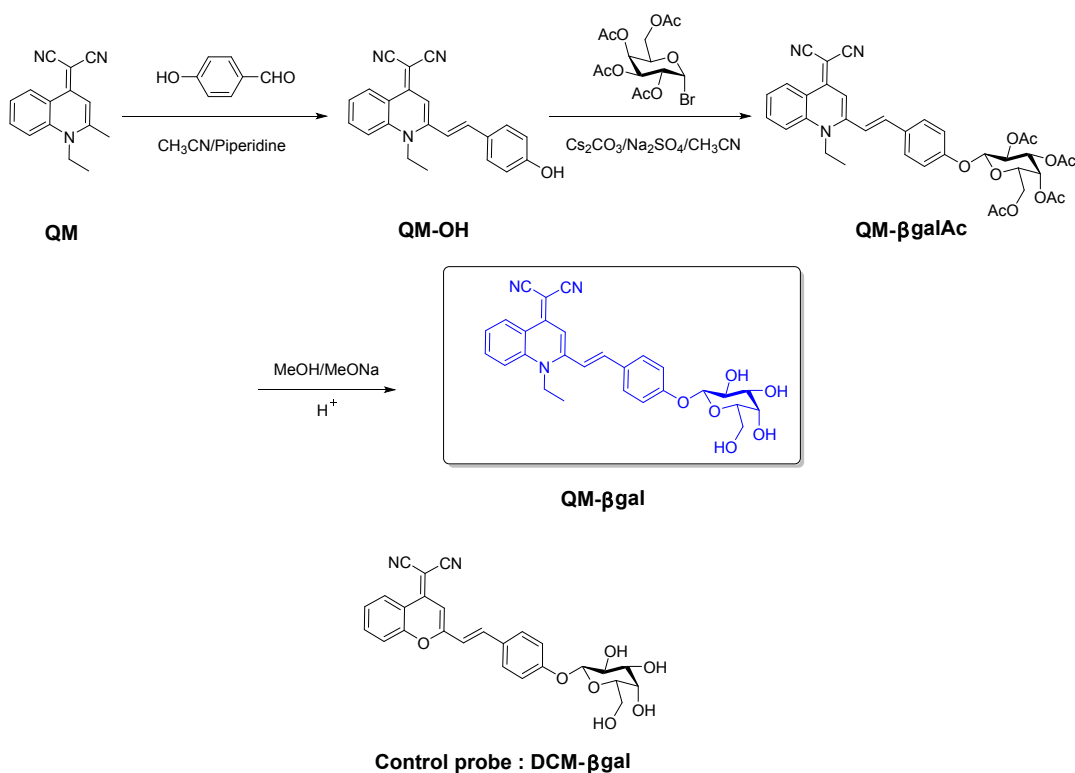
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## 1. Experimental section

### Materials and instruments

All solvents and chemicals, unless special stated, were purchased commercially in analytical grade and used without further purification.  $\beta$ -Galactosidase ( $\beta$ -gal) was supplied by J&K Scientific Ltd (Beijing, China).  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra in  $\text{DMSO}-d_6$  were obtained with a Bruker AvanceIII 400 MHz NMR spectrometer using TMS as an internal standard. High resolution mass spectrometry (HRMS) spectra were measured with a Waters LCT Premier XE spectrometer. UV-Vis absorption and fluorescence spectra were recorded on an Agilent Cary 60 spectrophotometer and Varian Cary Eclipse fluorescence spectrophotometer, respectively ( $10 \times 10$  mm quartz cuvette). Dynamic light scattering (DLS) experiments were conducted with Zetasizer Nano-ZS (Malvern Instruments, Worcestershire, UK), and scanning electron microscope (SEM) images were operated on a JEOL JSM-6360 scanning electron microscope. HPLC chromatograms were carried out using an Agilent 1100 series. Confocal fluorescence images were performed on confocal laser scanning microscope (CLSM, Nikon A1R).

### Synthesis of QM- $\beta$ gal



Scheme S1. Synthetic route of QM- $\beta$ gal

### Synthesis of QM-OH

QM (1.0 g, 4.25 mmol) and 4-hydroxybenzaldehyde (623 mg, 5.11 mmol) were dissolved in acetonitrile (30 mL) with piperidine (1.0 mL) under argon protection at room temperature. The mixture then was refluxed for 10 h. The solvent was removed by filtration, and the crude product was purified by recrystallization to afford the desired product QM-OH (827 mg, 2.44 mmol): yield 57%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>, ppm):  $\delta$  = 1.40 (t, *J* = 8.0 Hz, 3 H), 4.56 (q, *J* = 6.8 Hz, 2 H), 6.84 (d, *J* = 8.4 Hz, 2 H), 7.02 (s, 1 H), 7.29 (d, *J* = 16.0 Hz, 1 H), 7.36 (d, *J* = 15.6 Hz, 1 H), 7.61 (t, *J* = 7.6 Hz, 1 H), 7.67 (d, *J* = 8.4 Hz, 2 H), 7.92 (t, *J* = 7.6 Hz, 1 H), 8.08 (d, *J* = 8.8 Hz, 1 H), 8.92 (d, *J* = 8.4 Hz, 1 H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>, ppm):  $\delta$  = 13.63, 24.63, 26.28, 43.71, 46.24, 46.53, 106.42, 115.82, 116.58, 118.07, 120.60, 124.85, 125.09, 126.04, 130.04, 133.60, 137.84, 140.06, 149.68, 152.05, 159.80. High-resolution mass spectrometry (ESI negative ion mode for [M - H]<sup>-</sup>): Calcd. for C<sub>22</sub>H<sub>16</sub>N<sub>3</sub>O: 338.1293; found: 338.1299.

### Synthesis of QM- $\beta$ galAc

QM-OH (100 mg, 0.29 mmol) and tetra-O-acetyl- $\alpha$ -D-galactopyranosyl-1-bromide (200 mg, 0.48 mmol) were dissolved in acetonitrile (15 mL) with Cs<sub>2</sub>CO<sub>3</sub> (479 mg, 1.47 mmol) and Na<sub>2</sub>SO<sub>4</sub> (171.8 mg, 1.21 mmol) under argon protection at room temperature. The mixture then was stirred at room temperature for 4 h. After filtration, the solvent was removed under reduced pressure. The residue was taken up in sat.NH<sub>4</sub>Cl, and extracted with CH<sub>2</sub>Cl<sub>2</sub>. Next, the solution was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and the solvent was removed by evaporation again. Finally, the crude product was purified by silica gel chromatography with dichloromethane/methanol (100:1) to afford the desired product QM- $\beta$ galAc (93 mg, 0.14 mmol) as yellow solid: yield = 48%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>, ppm):  $\delta$  = 1.46 (t, *J* = 6.8 Hz, 3 H), 2.02 (s, 3 H), 2.09 (s, 3 H), 2.12 (s, 3 H), 2.22 (s, 3 H), 4.17 (d, *J* = 6.0 Hz, 2 H), 4.54 (t, *J* = 6.2 Hz, 1 H), 4.64 (d, *J* = 6.8 Hz, 2 H), 5.28-5.32 (m, 1 H), 5.35-5.36 (m, 1 H), 5.39-5.42 (m, 1 H), 5.65(d, *J*=7.6 Hz, 1 H), 7.08 (s, 1 H), 7.13 (d, *J* = 8.8 Hz, 2 H), 7.48 (d, *J* = 16 Hz, 1 H), 7.53 (d, *J* = 15.6 Hz, 1 H), 7.69 (t, *J* = 7.6 Hz, 1 H), 7.90 (d, *J* = 8.8 Hz, 2 H), 8.00 (t, *J* = 7.6 Hz, 1 H), 8.16 (d, *J* = 8.8 Hz, 1 H), 9.00 (d, *J* = 8.4 Hz, 1 H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>, ppm):  $\delta$  = 13.63, 20.32, 20.37, 20.43, 20.49, 43.83, 46.86, 61.29, 63.34, 67.19, 68.26, 70.10, 70.44, 97.25, 106.74, 116.51, 118.08, 119.41, 120.58, 124.93, 125.10, 129.76, 129.98, 133.68, 137.79, 138.93, 149.24, 152.23, 157.52, 169.22, 169.54, 169.84, 169.95. High-resolution mass spectrometry (ESI positive ion mode for [M + Na]<sup>+</sup>): Calcd. for C<sub>36</sub>H<sub>35</sub>N<sub>3</sub>O<sub>10</sub>Na: 692.2220; found: 692.2217.

### Synthesis of QM- $\beta$ gal

QM- $\beta$ galAc (88 mg, 0.13 mmol) was added MeONa (70 mg, 1.3 mmol) in methanol (5 mL) and the mixture was stirred at room temperature for 3 h. Then the reaction mixture was neutralized with Amberlite IR-120 plus (H<sup>+</sup>). After the Amberlite IR-120 plus (H<sup>+</sup>) was filtered off, the solvent was removed by evaporation. Finally, the crude product was purified by silica gel chromatography with dichloromethane/methanol (40:1) to afford the desired product QM- $\beta$ gal (41 mg, 0.08 mmol) as yellow solid: yield = 62%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>, ppm):  $\delta$  = 1.41 (t, *J* = 6.4 Hz, 3 H), 3.44-3.63 (m, 4 H), 3.72 (s, 2 H), 4.57 (t, *J* = 6.8 Hz, 2 H), 4.68 (s, 2 H), 4.93 (d, *J* = 8.4 Hz, 2 H), 5.22 (d, *J* = 4.4 Hz, 1 H), 7.03 (s, 1 H), 7.11 (d, *J* = 8.4 Hz, 2 H), 7.42 (s, 2 H), 7.62 (t, *J* = 8.0 Hz, 1 H), 7.79 (d, *J* = 8.4 Hz, 2 H), 7.93 (t, *J* = 8.0 Hz, 1 H), 8.10 (d, *J* = 8.8 Hz, 1 H), 8.94 (d, *J* = 8.4 Hz, 1 H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>, ppm):  $\delta$  = 13.63, 13.92, 28.97, 31.11, 31.25, 43.82, 46.63, 60.32, 68.08, 70.21, 73.24, 75.55, 100.56, 106.67, 116.44, 118.09, 118.56, 120.59, 124.92, 125.09, 128.61, 129.62, 133.69, 137.81, 139.33, 149.41, 152.20, 158.76. High-resolution mass spectrometry (ESI positive ion mode for [M + H]<sup>+</sup>): Calcd. for C<sub>28</sub>H<sub>28</sub>N<sub>3</sub>O<sub>6</sub>: 502.1978; found: 502.1976.

## Cell experiment

### Cell lines

Human embryonic kidney 293T cells was supplied by the Institute of Cell Biology (Shanghai, China). 293T cells were cultured at 37 °C under a humidified 5% CO<sub>2</sub> atmosphere in DMEM (GIBCO/Invitrogen, Camarillo, CA, USA), which were supplemented with 10% fetal bovine serum (FBS, Biological Industry, Kibbutz Beit Haemek, Israel) and 1% penicillin-streptomycin (10,000 U mL<sup>-1</sup> penicillin and 10 mg/ml streptomycin, Solarbio life science, Beijing, China).

Human ovarian adenocarcinoma cells (SKOV-3 cells) was supplied by the Institute of Cell Biology (Shanghai, China). SKOV-3 cells were cultured at 37 °C under a humidified 5% CO<sub>2</sub> atmosphere in McCoy's 5A (GIBCO/Invitrogen, Camarillo, CA, USA), which were supplemented with 10% fetal bovine serum (FBS, Biological Industry, Kibbutz Beit Haemek, Israel) and 1% penicillin-streptomycin (10,000 U mL<sup>-1</sup> penicillin and 10 mg/ml streptomycin, Solarbio life science, Beijing, China).

### *In vitro* cytotoxicity assay

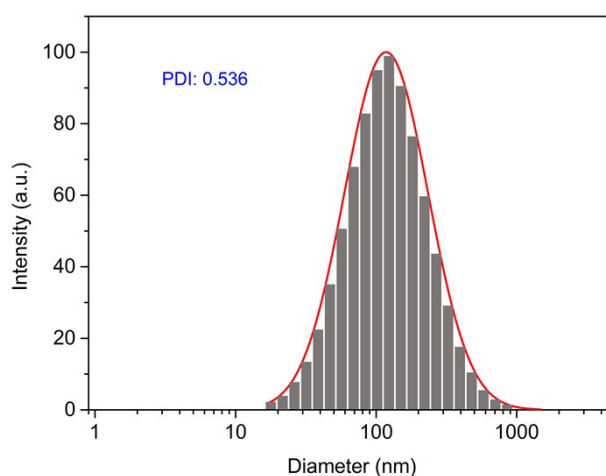
The cytotoxicity of QM- $\beta$ gal or QM-OH in both cancer and normal cell lines was evaluated by a MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. Briefly, cells were seeded into 96-

well plates at a density of  $1 \times 10^4$  cells/well and were cultured at 37 °C under a humidified 5% CO<sub>2</sub> atmosphere for 12 h. Then, the cells were exposed to the various concentrations (1, 2, 5, 10 μM) of QM-βgal or QM-OH, and for negative control group, 100.0 μL of culture medium were added. After incubation at 37 °C under a humidified 5% CO<sub>2</sub> atmosphere for 24 h, MTT solution (5 mg/mL in PBS, 10 μL) was added to the media and incubated for another 4 h, and the absorbance at 490 nm was measured with a Multimode Plate Reader (BioTek, USA). The relative cell viability (%) was calculated by the following formula: viability (%) = mean absorbance value of the treatment group-blank/mean absorbance value of the control-blank × 100.

### Cells imaging

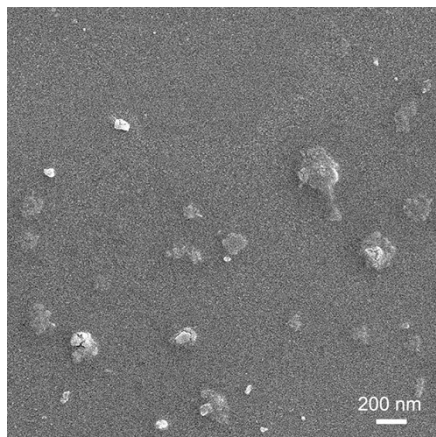
Cells were seeded onto glass-bottom Petri dishes in culture medium (1.5 mL) and allowed to adhere for 12 h before imaging. Probe QM-βgal at a final concentration of 10 μM (containing 0.1% DMSO) were added into culture medium and incubated for different time at 37 °C under a humidified 5% CO<sub>2</sub> atmosphere. Cells imaging was captured by using a confocal laser scanning microscope (CLSM, Nikon A1R system, Japan) with a 60× oil immersion objective lens. The fluorescence signals of cells incubated with probes were collected at 500–650 nm under excitation wavelength at 404 nm.

## 2. Hydrodynamic diameter of QM-OH aggregates



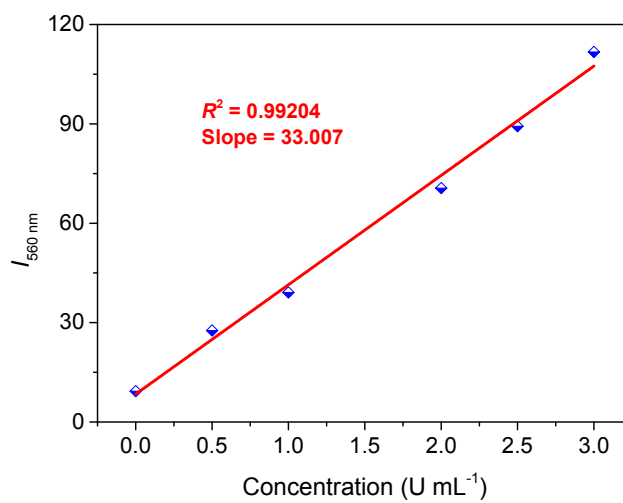
**Fig. S1** Hydrodynamic diameter of QM-OH (10 μM) in a mixture of water/DMSO (v/v = 95/5) obtained from dynamic light scattering (DLS).

### 3. SEM image of QM-OH aggregates



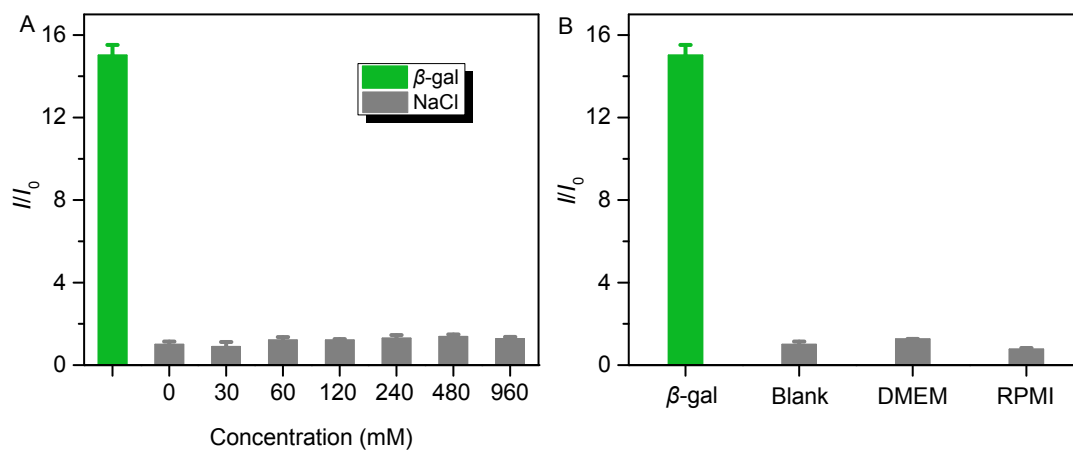
**Fig. S2** SEM image of aggregates formed by QM-OH (10  $\mu\text{M}$ ) in a mixture of water/DMSO (v/v = 95/5).

### 4. The detection limit of QM- $\beta$ gal



**Fig. S3** A linear correlation between fluorescence intensity at 560 nm ( $I_{560 \text{ nm}}$ ) and concentration of  $\beta$ -gal. Note: The detection limit was calculated to be  $1.0 \times 10^{-3} \text{ U mL}^{-1}$  ( $3\sigma/\text{slope}$ ).

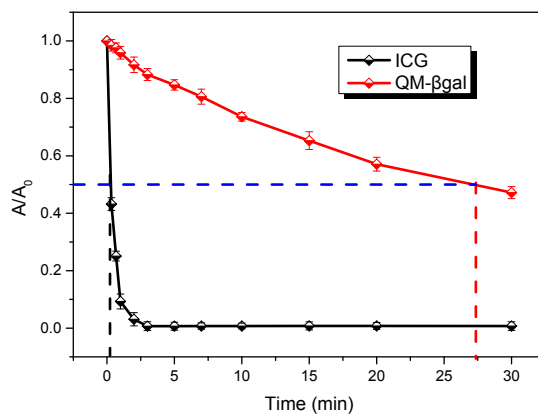
## 5. Effects of ionic strength and culture medium



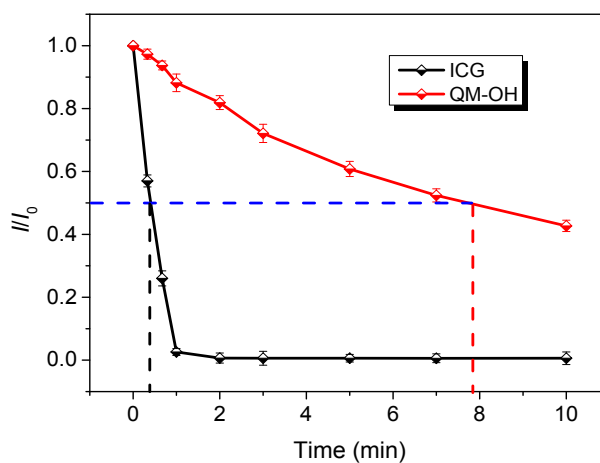
**Fig. S4** Fluorescence response of QM-βgal (10 μM) to (A) varied concentrations of NaCl (0-960 mM) and (B) cell culture medium (DMEM and RPMI) for 30 min,  $\lambda_{\text{ex}} = 434$  nm.  $I/I_0$  represents the fluorescence intensity ratio at 560 nm, and  $I_0$  is the fluorescence intensity of free QM-βgal.



## 6. The photostability of QM-βgal QM-OH

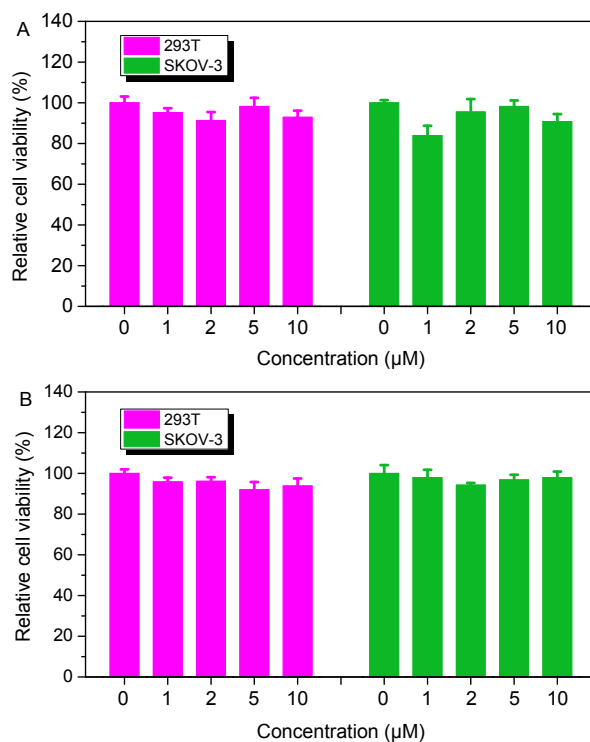


**Fig. S5** Time-dependent absorption of ICG (10  $\mu$ M, monitored at 780 nm), QM-βgal (10  $\mu$ M, monitored at 434 nm) in a mixture of water/DMSO ( $v/v = 95/5$ ) under continuous illumination.



**Fig. S6** Time-dependent fluorescence intensity of ICG (10  $\mu$ M, monitored at 812 nm, and  $\lambda_{ex} = 780$  nm), QM-OH (10  $\mu$ M, monitored at 560 nm, and  $\lambda_{ex} = 434$  nm) in a mixture of water/DMSO ( $v/v = 95/5$ ) under continuous illumination.

## 7. *In vitro* cytotoxicity of QM-βgal or QM-OH



**Fig. S7** Relative cell viability of 293T or SKOV-3 cells *in vitro* after incubation with (A) QM-βgal and (B) QM-OH at various concentrations for 24 h.

## 8. Pearson's correlation coefficient in colocalization experiments

**Table S1** Pearson's correlation coefficient of various organelles in colocalization experiments

Organelle	Golgi body	Lysosome	Endoplasmic reticulum	Mitochondria
Pearson's correlation coefficient	0.629	0.335	0.835	0.933

## 9. Characterization of compounds

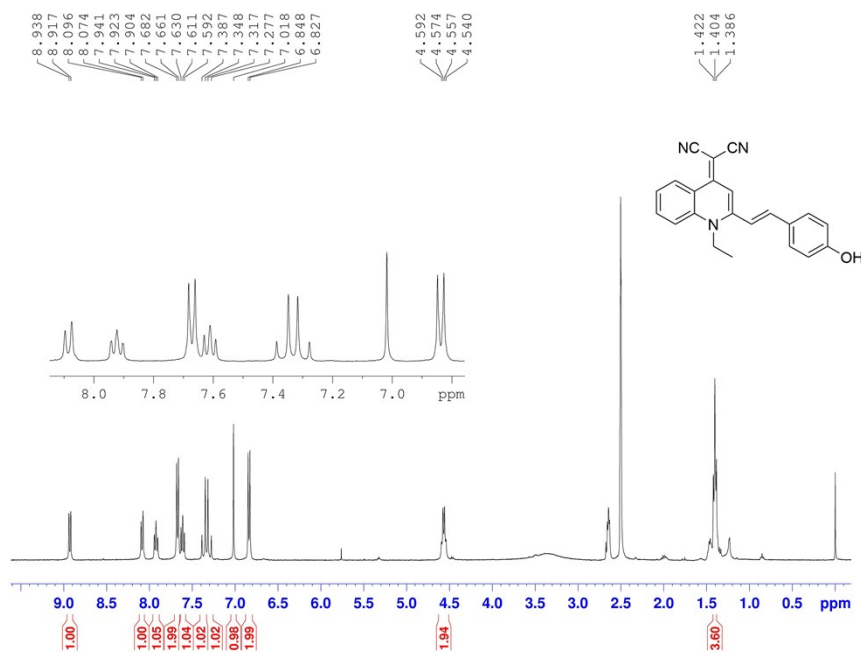


Fig. S8 <sup>1</sup>H NMR spectrum of QM-OH in DMSO-*d*<sub>6</sub>.

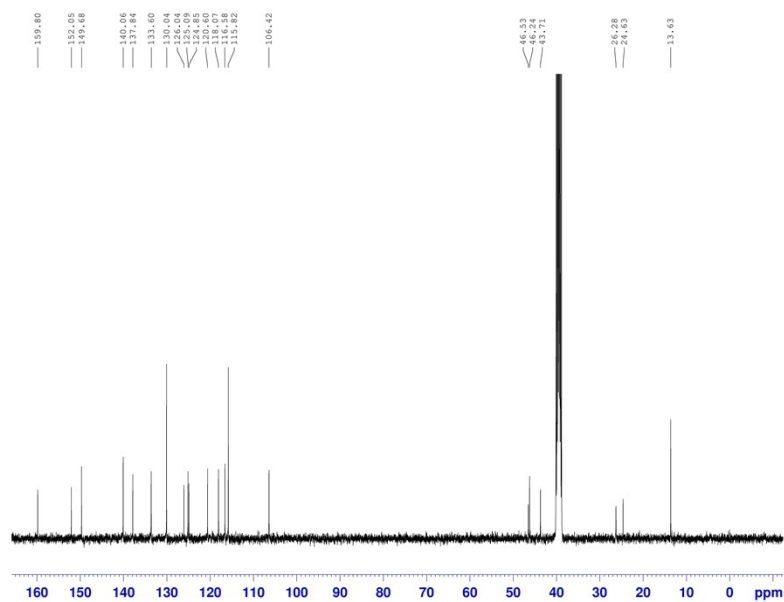
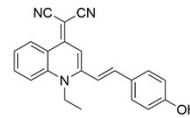


Fig. S9 <sup>13</sup>C NMR spectrum of QM-OH in DMSO-*d*<sub>6</sub>.

Elemental Composition Report

Single Mass Analysis

Tolerance = 50.0 PPM / DBE: min = -1.5, max = 100.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

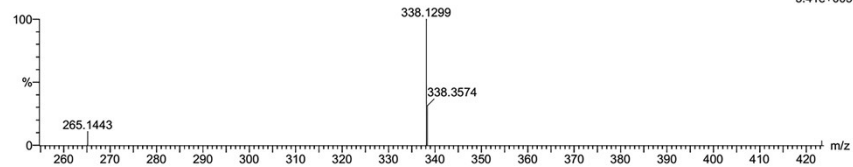


Monoisotopic Mass, Even Electron Ions  
 4 formula(e) evaluated with 1 results within limits (up to 1 closest results for each mass)  
 Elements Used:  
 C: 0-22 H: 0-16 N: 0-3 O: 0-1  
 WH-ZHU

ECUST institute of Fine Chem

18-Sep-2016  
 20:42:27  
 1: TOF MS ES-  
 3.41e+005

ZW-GKZ-0918 4 (0.153) Cm (2:45)



Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
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Fig. S10 HRMS spectrum of QM-OH.

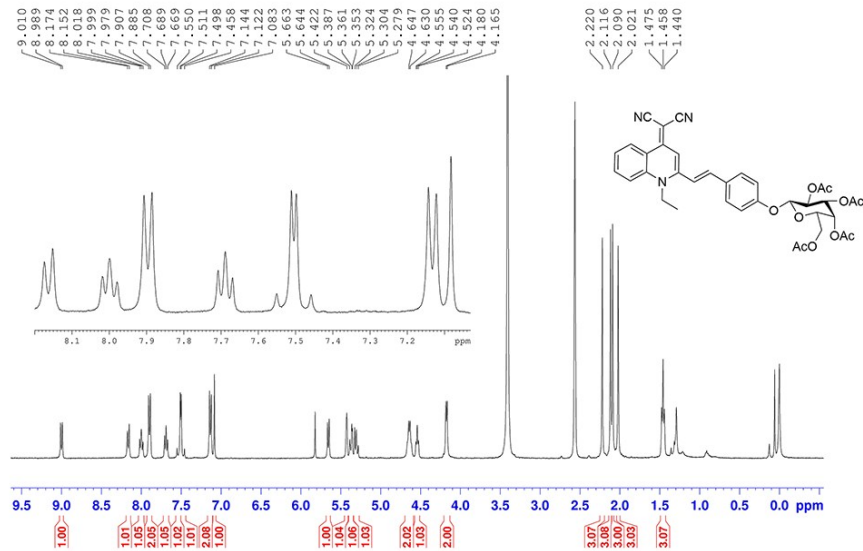


Fig. S11 <sup>1</sup>H NMR spectrum of QM-βgalAc in DMSO-d<sub>6</sub>.

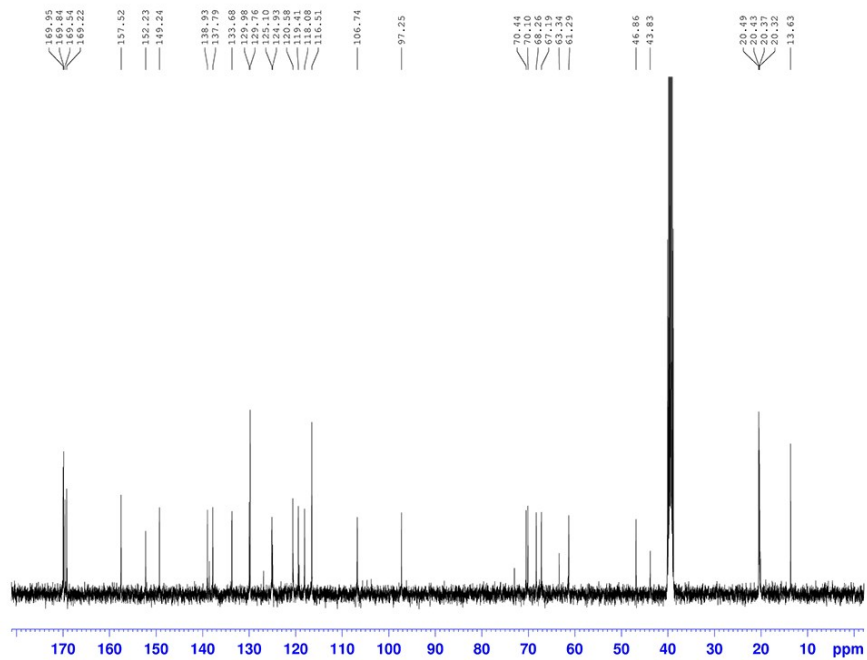


Fig. S12  $^{13}\text{C}$  NMR spectrum of QM- $\beta$ galAc in  $\text{DMSO-}d_6$ .

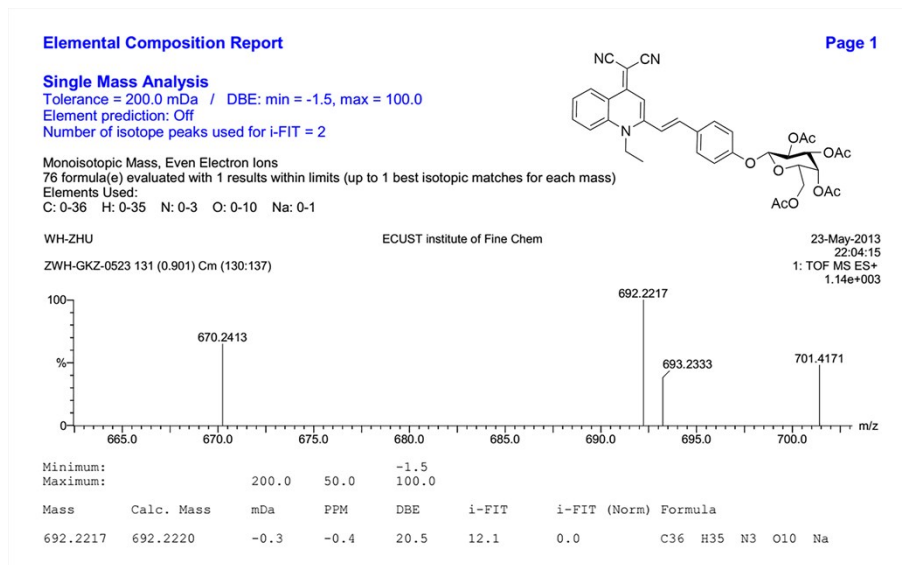
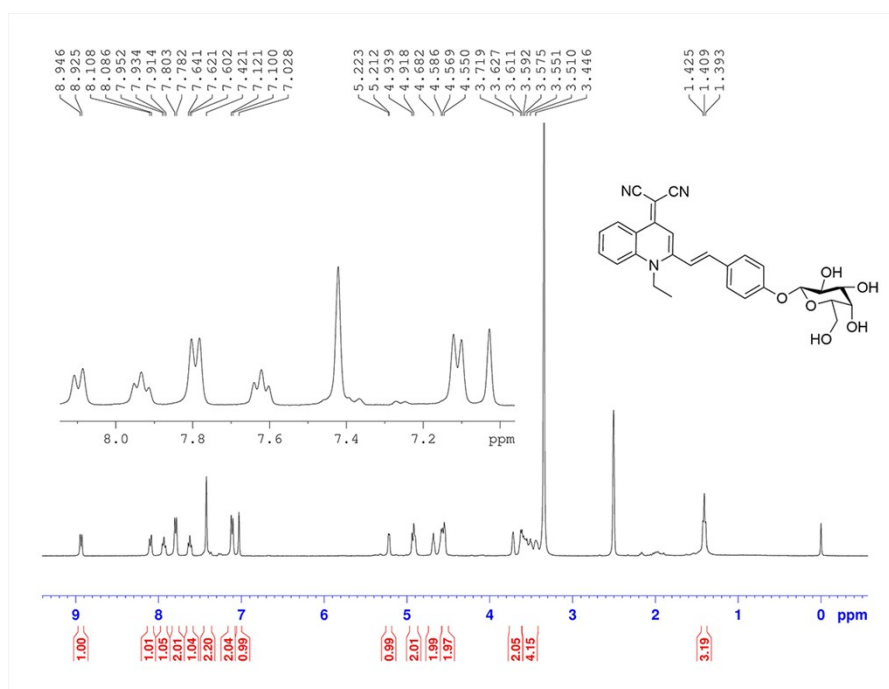
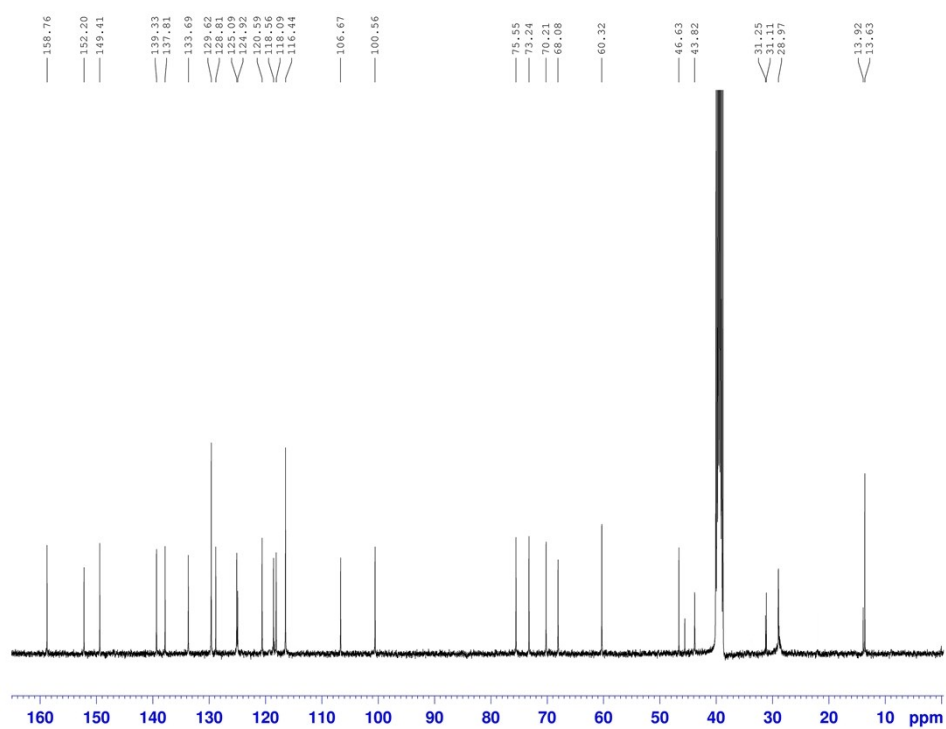


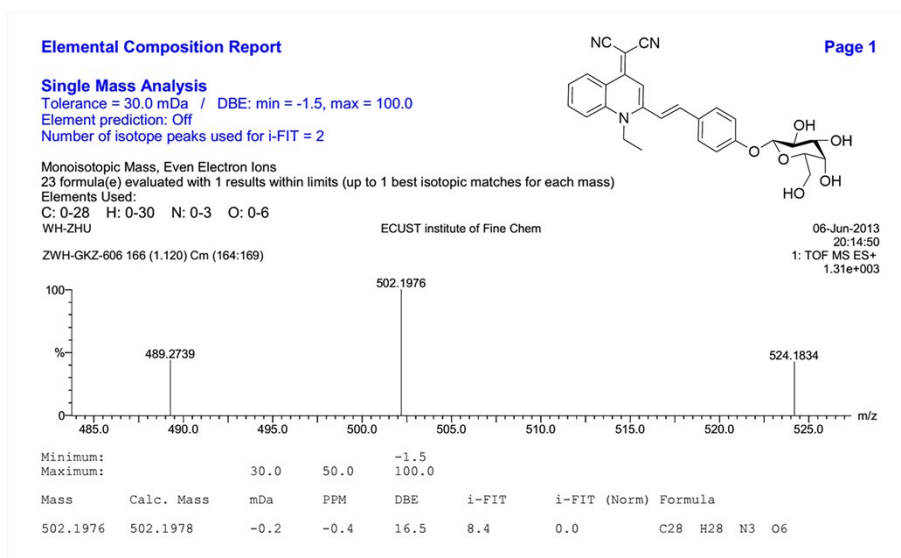
Fig. S13 HRMS spectrum of QM- $\beta$ galAc.



**Fig. S14** <sup>1</sup>H NMR spectrum of QM-βgal in DMSO-*d*<sub>6</sub>.



**Fig. S15** <sup>13</sup>C NMR spectrum of QM-βgal in DMSO-*d*<sub>6</sub>.



**Fig. S16** HRMS spectrum of QM- $\beta$ gal.