Supplementary Information Document

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Supplementary Tables and Figures

Entry	Sequence	Enzyme	k _m (μM)	k _{cat} (s⁻¹)	k _{cat} /k _m (s ⁻¹ M ⁻¹)
1	NAC ₂₀ KKNT	PPT1 APT1 APT2	7.9 ± 0.9 6.2 ± 0.8 9.4 ± 0.9	0.28 ± 0.01 0.38 ± 0.02 0.199 ± 0.006	35,000 61,000 21,000
2	GAC ₂₀ KKNT	PPT1 APT1 APT2	2.0 ± 0.1 2.7 ± 0.3 5.8 ± 0.6	0.28 ± 0.04 0.46 ± 0.01 0.106 ± 0.004	140,000 170,000 18,000
3	WAC ₂₀ KKNT	PPT1 APT1 APT2	12.0 ± 2.0 8.9 ± 2.0 7.9 ± 0.7	0.0063 ± 0.0004 0.027 ± 0.003 0.012 ± 0.004	530 3,300 1,500
4	ASC ₂₀ KKNT	PPT1 APT1 APT2	4.4 ± 0.5 1.2 ± 0.2 4.8 ± 0.4	0.43 ± 0.02 0.56 ± 0.02 0.29 ± 0.009	98,000 470,000 60,000
5	GSC ₂₀ KKNT	PPT1 APT1 APT2	8.2 ± 0.6 1.7 ± 0.3 5.6 ± 0.4	0.41 ± 0.009 0.66 ± 0.02 0.266 ± 0.006	50,000 390,000 48,000
6	WYC ₂₀ KKNT	PPT1 APT1 APT2	4.3 ± 0.6 8.4 ± 1.0 10.0 ± 2.0	0.0078 ± 0.0003 0.024 ± 0.002 0.0045 ± 0.0003	1,500 2,900 450
7	ASC ₂₀ KRNT	PPT1 APT1 APT2	5.7 ± 0.6 4.9 ± 0.7 5.6 ± 0.7	0.90 ± 0.03 0.95 ± 0.04 0.227 ± 0.009	160,000 190,000 41,000
8	GAC ₂₀ KRNT	PPT1 APT1 APT2	7.5 ± 0.7 5.9 ± 0.4 16.0 ± 2.0	0.77 ± 0.02 0.64 ± 0.01 0.056 ± 0.003	100,000 110,000 3,500
9		APT1	4.4 ± 0.6	0.044 ± 0.002	10,000
10		APT1	14.0 ± 1.0	0.03 ± 0.001	2,100
11		APT1 APT2	155 ± 84 91 ± 73	0.003 ± 0.001 0.003 ± 0.001	19 33
12 13	GAC ₂₀ AANT AHC ₂₀ DRNT	APT1 PPT1 APT1	9.0 ± 3.0 6.7 ± 0.6 4.8 ± 0.7	0.016 ± 0.002 0.054 ± 0.001 0.163 ± 0.007	1,800 8,060 34,000
		APT2	5.3 ± 0.5	0.00259 ± 0.00008	490
14	PAC ₂₀ EANT	APT1	23 ± 8	0.0022 ± 0.0003	97
15	SYC ₂₀ IANT	APT1	23 ± 4	0.040 ± 0.003	1,700
16	ADC ₂₀ SRNT	PPT1 APT1	11 ± 6 7 ± 1	0.012 ± 0.002 0.031 ± 0.002	1,090 4,400 170
17	гос ₂₀ кант	APT1	4∠ ± ∠5 59 ± 19	0.007 ± 0.002 0.036 ± 0.006	610
18	FRC ₂₀ KANT	PPT1 APT1 APT2	2.3 ± 0.4 1.7 ± 0.1 2.7 ± 0.3	0.032 ± 0.002 0.226 ± 0.006 0.062 ± 0.003	14,000 130,000 23,000
	1		1	1	

Table S1. Catalytic properties of APTs, for the hydrolysis of fluorogenic peptides, Relatedto figures 3 and 5. Amino acid substitutions at positions P2, P1, P1' and P2' are in red letters.



Figure S1. Characterization of fluorogenic peptides, Related to figures 3 and 5. HPLC purified peptides were characterized by LCMS, UV traces are measured at a wavelength of 220 nm. $[M+H]^+$ parent ions as well as $[M+2H]^{2+}$ were compared to calculated masses.



Figure S2. Characterization of fluorogenic peptides, Related to figures 3 and 5. HPLC purified peptides were characterized by LCMS, UV traces are measured at a wavelength of 220 nm. $[M+H]^+$ parent ions as well as $[M+2H]^{2+}$ were compared to calculated masses.



Figure S3. Depalmitoylase activity in cell lysates can be interrogated with selective inhibitors, Related to figure 1. Cell lysates (20 μ g per reaction) were pre-incubated with inhibitors (10 μ M) or DMSO as control, and then QStE (10 μ M) was added to measure residual activity (Ex = 410 nm, Em = 450 nm) over the course of 60 minutes.







Figure S5. FP-Rho competition labeling confirms selective inhibition of APT1 and APT2, Related to figure 6. Tumor tissue lysate ($20 \ \mu g$) was incubated with DMSO as control or inhibitors at 10 μ M for 30 minutes on ice, then 1 μ M of FP-Rho was added and samples were incubated for additional 20 min on ice. Lysates were resolved by SDS-PAGE, and the gel was scanned for fluorescence (532-nm laser, 610-nm filter, PMT800). Western blot for GAPDH is shown for loading control.



Figure S6. APT2 exhibits selectivity towards a Scribble-derived peptide sequence, Related to figure 5. The peptide WRCNRVH was synthesized based on the sequence of human Scribble. Activity of the recombinant depalmitoylases (50 nM) for the hydrolysis of the peptide (1 μ M) was measured over 30 minutes. Error bars represent the S.D. of three replicates.

1) Supplementary Synthesis schemes



Scheme S1. Synthesis of DABCYL-azide, Related to Compound Synthesis methods in STAR Method Section. Reaction conditions; a) LiN₃, H₂O, 12 hr. b) NHS, EDC, DMF, 16 hr. c) 1, DIPEA, DMF, 4 hr.



Scheme S2. Synthesis of 7-Hydroxycoumarin-3-carboxylic acid N-succinimidyl ester, Related to Compound Synthesis methods in STAR Method Section. Reaction conditions; a) Diethyl malonate, EtOH, piperidine, 12 hr. b) NaOH, H₂O, 1 hr. c) NHS, EDC, DMF, 3 hr.



Scheme S3. Synthesis of QStE. Related to Compound Synthesis methods in STAR Method Section. Reaction conditions; a) HBTU, Morpholine, DMF, 30 min. b) 20% piperidine, DMF, 30 min. c) 6, DIPEA, DMF, 4 hr. d) 1. 5% TFA, DCM, 2 hr. 2. 17-ODYA, (COCI)₂, DMF, DCM 3. TFA, 3 hr. e) 3, CuSO₄, L-ascorbic acid, DMF, 40 ^oC, 2 hr.



Scheme S4. Synthesis of QSE. Related to Compound Synthesis methods in STAR Method Section. Reaction conditions; a) HBTU, Morpholine, DMF, 30 min. b) 20% piperidine, DMF, 30 min. c) 6, DIPEA, DMF, 4 hr. d) 1. 50% TFA, DCM, 2 hr. 2. 17-ODYA, (COCI)₂, DMF, DCM 3. TFA, reflux, 4 hr. e) 3, CuSO₄, L-ascorbic acid, DMF, 40 ^oC, 2 hr.



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Scheme S5. Synthesis of 17-ODYA, Related to Compound Synthesis methods in STAR Method Section. Reaction conditions; a) Br₂, KOH, DMSO, 100 ^oC, 1hr. b) LiAlH₄, diethyl ether, 4 hr. c) KH, APA, 16 hr. d) PDC, DMF, 12 hr.



Scheme S6. Synthesis of cysteine building block (C₂₀), Related to Compound Synthesis methods in STAR Method Section. Reaction conditions; a) 1. TFA, TIS, DCM 2. CHCl₃, 17-ODYA, SOCl₂, reflux, 1hr. 3. 40 $^{\circ}$ C, 4 hr. b) Cu(II)SO₄, Ascorbic acid, **3**, DMF, 40 $^{\circ}$ C, 2 hr.