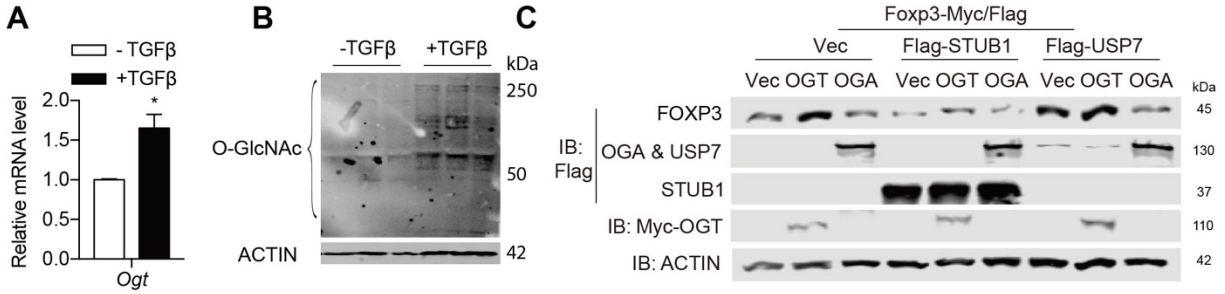
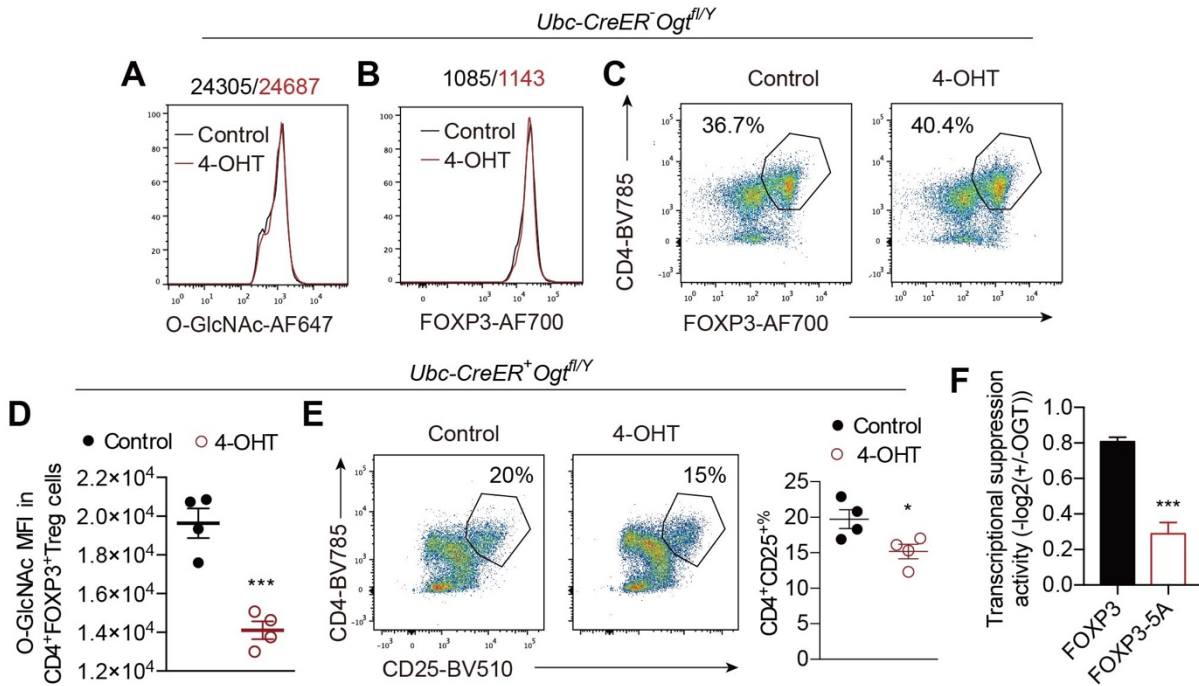


**Liu et al.**

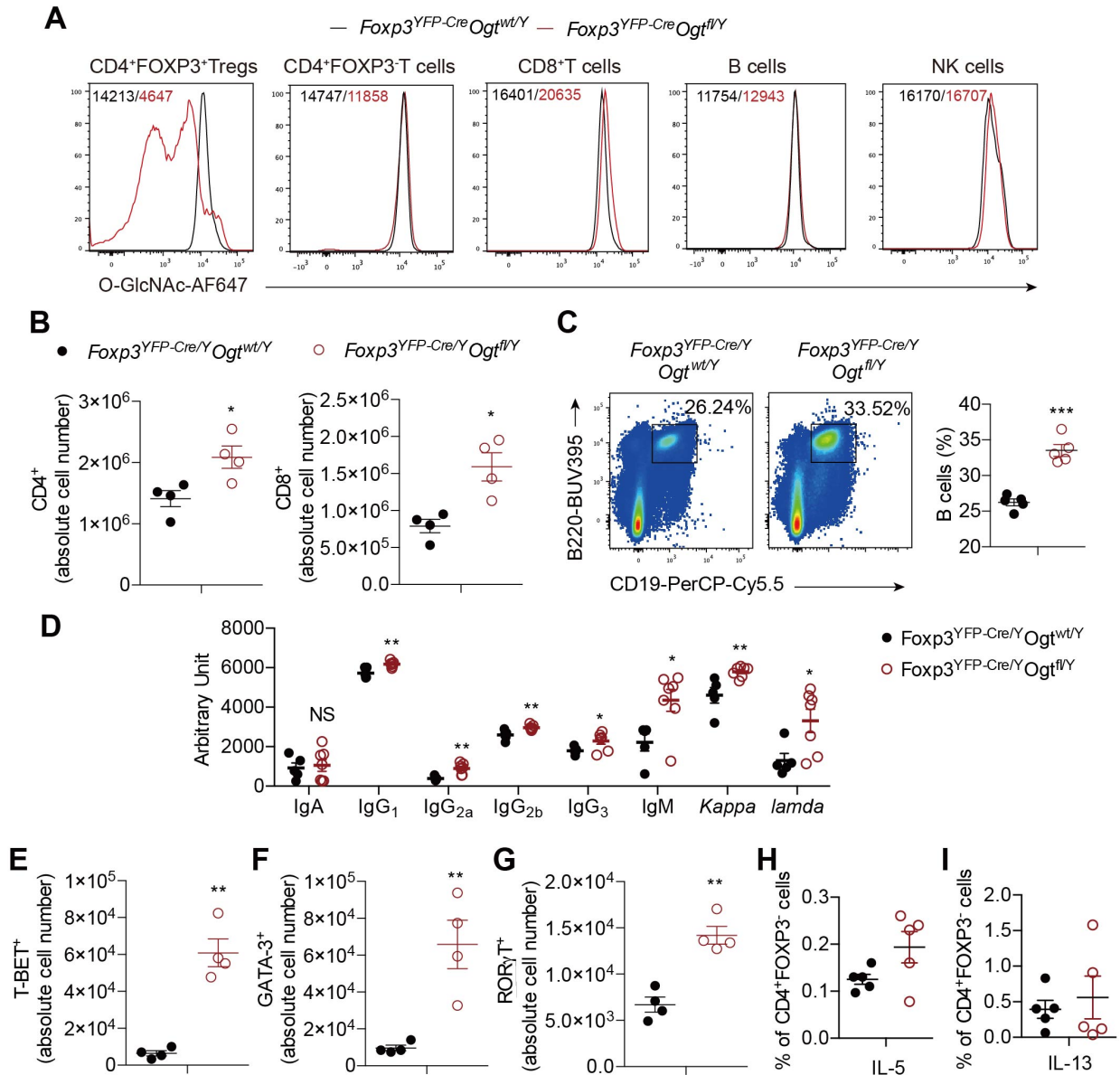
**The lineage stability and suppressive program of regulatory T cells require protein O-GlcNAcylation**



**Supplementary Fig. 1** (A, B) CD4<sup>+</sup>CD25<sup>-</sup> naïve T cells isolated from the LNs and spleen of wildtype mice were activated for 5-day with anti-CD3/CD28 beads in the presence of TGF-β to generate iTreg cells ex vivo. mRNA levels of *Ogt* (A, n = 3) and global protein O-GlcNAcylation (B) were measured. (C) Expression of FOXP3 protein in HEK 293 cells co-transfected with OGT or OGA and STUB1 or USP7. Data are shown as mean ± s.e.m. \*, p<0.05 by unpaired student's t-test.

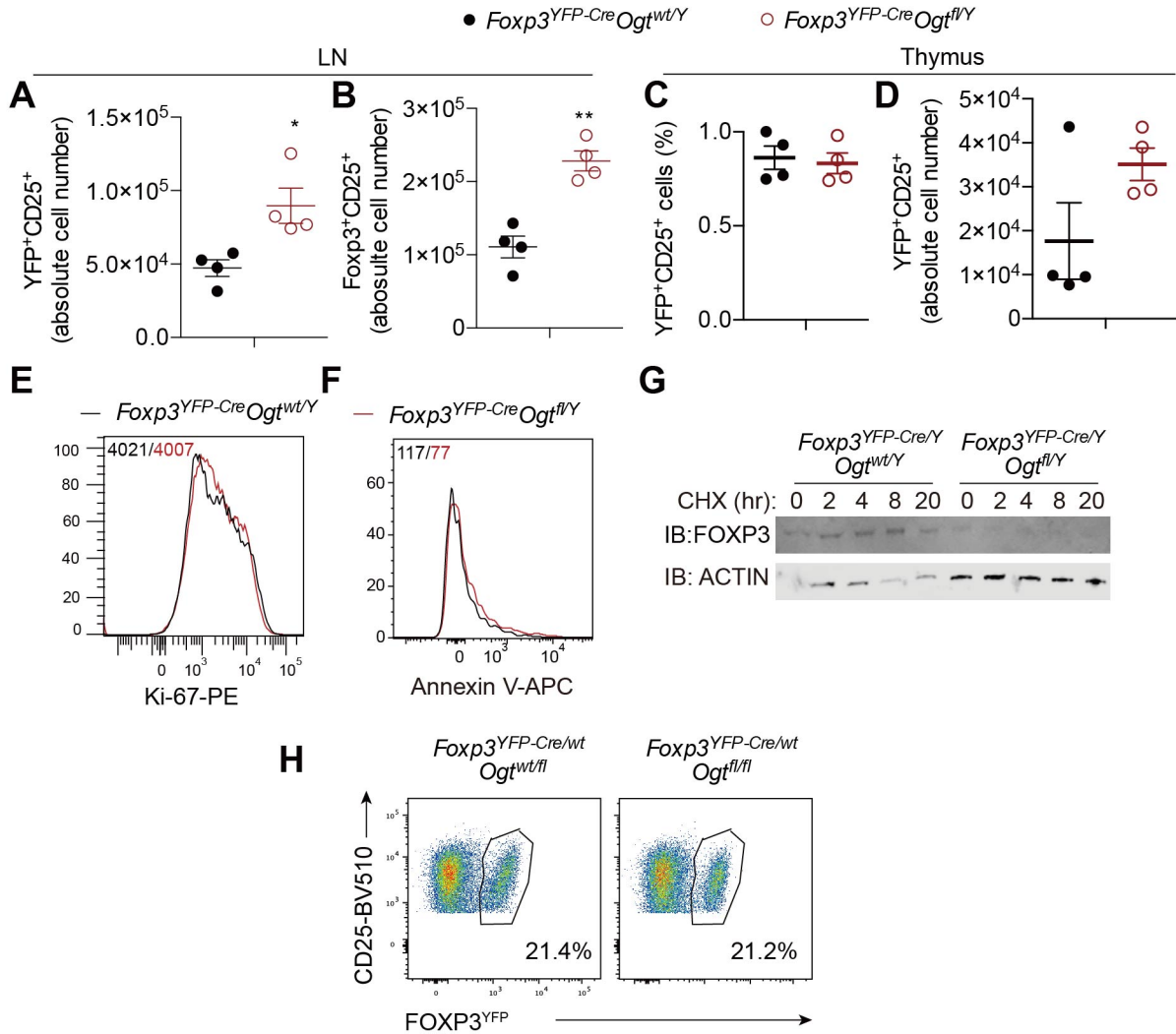


**Supplementary Fig. 2** (A-C) Isolated Treg cells from *Ubc-CreER<sup>+</sup>Ogt<sup>fl/y</sup>* mice were treated for 3-day 4-OHT ex vivo. Ethanol was used as a control. MFI of O-GlcNAcylation (A) and FOXP3 (B) among CD4<sup>+</sup>FOXP3<sup>+</sup> Treg cells. Representative flow cytometry of CD4<sup>+</sup>FOXP3<sup>+</sup> Treg cells were shown in (C). (D) MFI of O-GlcNAcylation in CD4<sup>+</sup>FOXP3<sup>+</sup> Treg cells from *Ubc-CreER<sup>+</sup>Ogt<sup>fl/y</sup>* mice that were treated with ethanol or 4-OHT, n = 4. (E) Representative flow cytometry and quantification of the frequencies of CD4<sup>+</sup>CD25<sup>+</sup> Treg cells from *Ubc-CreER<sup>+</sup>Ogt<sup>fl/y</sup>* mice that were treated with ethanol or 4-OHT, n = 4. (F) Forkhead responsive element (FHRE)-luciferase assay in HEK 293 cells transfected with FOXP3 or FOXP3-5A in the presence/absence of OGT. pS1-Rluc was co-transfected to control transfection efficiency. OGT-induced activation of FOXP3 suppressive activity on FHRE was plotted (n = 6). Data are shown as mean ± s.e.m. \*, p<0.05, \*\*\*, p<0.001 by unpaired student's t-test.

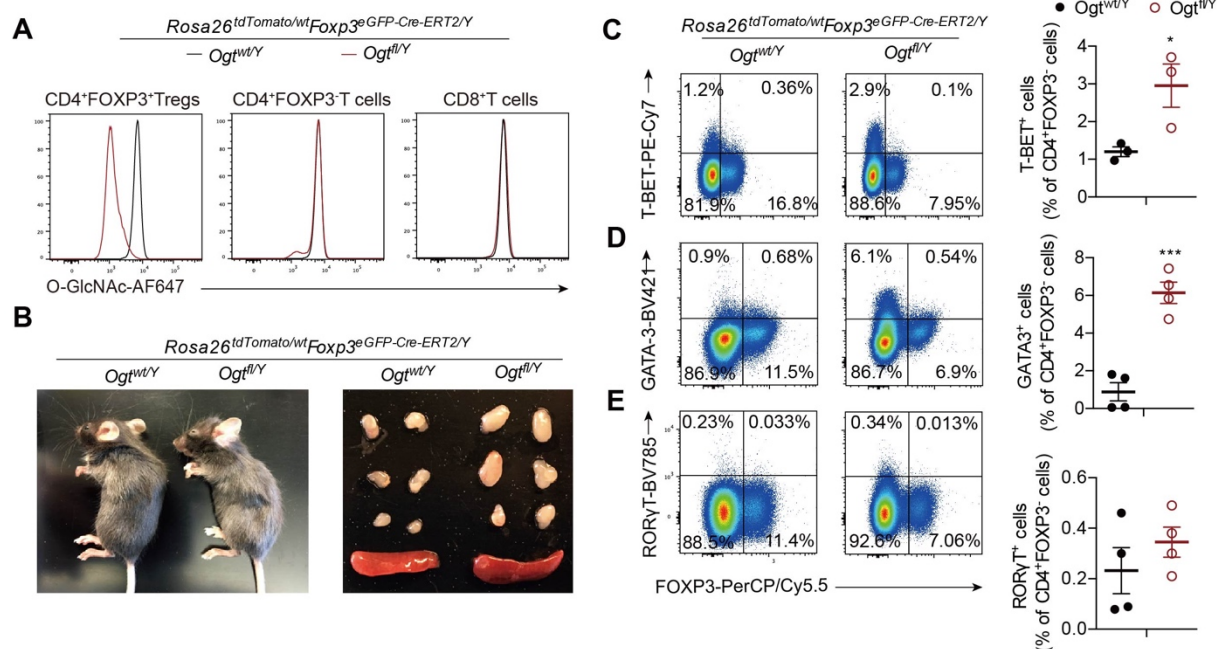


**Supplementary Fig. 3** (A) MFI of O-GlcNAcylation in indicated cell populations from *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>wt/Y</sup>* and *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>fl/Y</sup>* mice. (B) Total number of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the LNs from 2-week-old *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>wt/Y</sup>* and *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>fl/Y</sup>* mice, n = 4 each group. (C) Representative flow cytometry plot and quantification of the frequencies showing CD19<sup>+</sup>B220<sup>+</sup> B cells among single cells in the LNs of 2-week-old *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>wt/Y</sup>* and *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>fl/Y</sup>* mice (n = 5). (D) Levels of IgA, IgG<sub>1</sub>, IgG<sub>2a</sub>, IgG<sub>2b</sub>, IgG<sub>3</sub>, IgM, Kappa and lamda in sera of *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>wt/Y</sup>* (n = 5) and *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>fl/Y</sup>* (n = 7) mice. (E-G) Total numbers of T-BET<sup>+</sup> (E), GATA-3<sup>+</sup> (F) and RORγT<sup>+</sup> (G) cells among CD4<sup>+</sup> T cells in the LNs from 2-week-old *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>wt/Y</sup>* and *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>fl/Y</sup>* mice, n=4 each group. (H, I) Frequencies of IL-5<sup>+</sup> (H) and IL-13<sup>+</sup> (I) cells in CD4<sup>+</sup>FOXP3<sup>-</sup> T cells stimulated with PMA/Ionomycin in the LNs of 2-week-old *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>wt/Y</sup>* and *Foxp3<sup>YFP-Cre/Y</sup>Ogt<sup>fl/Y</sup>* mice, n = 5. Data are shown as mean ± s.e.m. \*, p<0.05; \*\*, p<0.01; \*\*\*, p<0.001 by unpaired student's *t*-test.

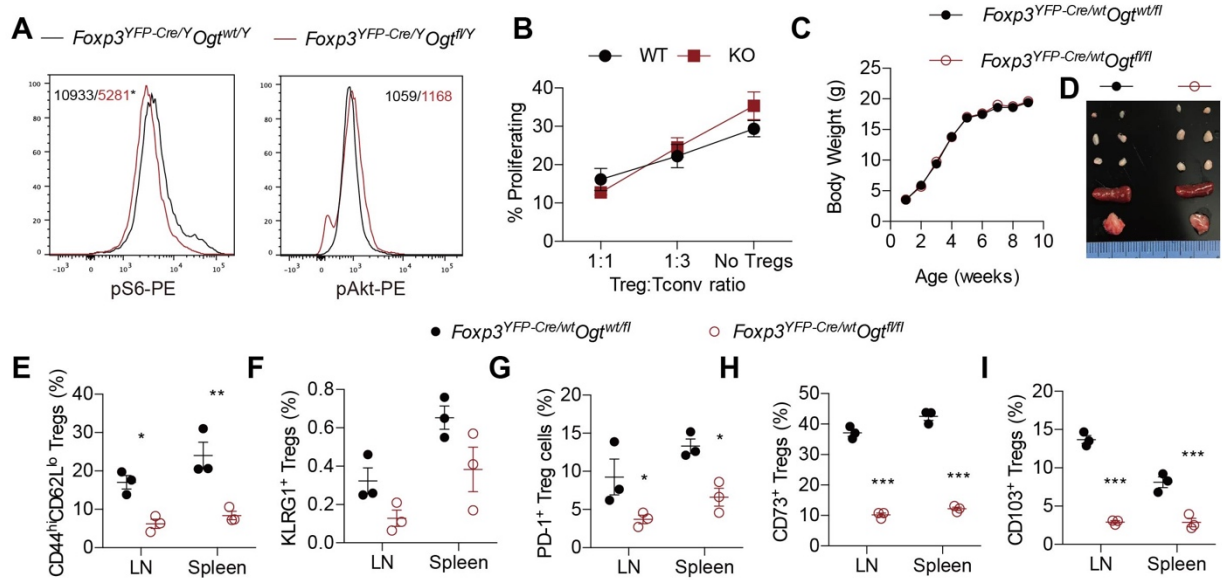




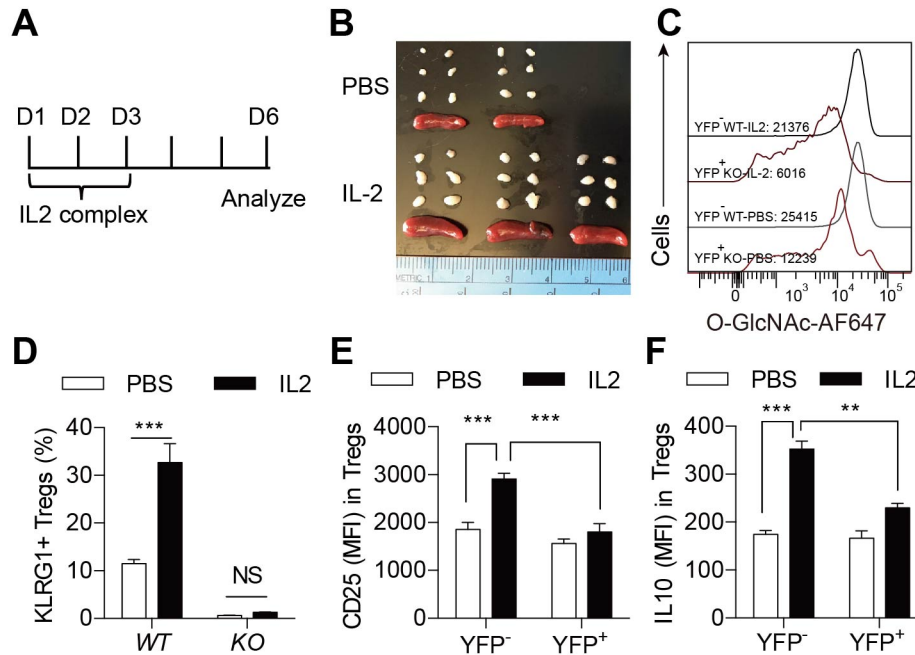
**Supplementary Fig. 4** (A, B) Total numbers of YFP<sup>+</sup>CD25<sup>+</sup> (A) and FOXP3<sup>+</sup>CD25<sup>+</sup> (B) Treg cells among CD4<sup>+</sup> T cells in the LNs from 2-week-old *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>wt/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>fl/Y</sup> mice, n = 4 each group. (C, D) Frequency (C) and total number (D) of YFP<sup>+</sup>CD25<sup>+</sup> Treg cells among CD4<sup>+</sup> T cells in the thymus from 2-week-old *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>wt/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>fl/Y</sup> mice, n = 4 each group. (E, F) MFI of Ki-67 (E) and Annexin V (F) among YFP<sup>+</sup>CD25<sup>+</sup> Treg cells in the LNs from 2-week-old *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>wt/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>fl/Y</sup> mice. (G) FOXP3 stability in Treg cells isolated from *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>wt/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>fl/Y</sup> mice was determined by CHX treatment. (H) Frequencies of YFP<sup>+</sup> Treg cells in the LNs from *Foxp3*<sup>YFP-Cre/wt</sup>*Ogt*<sup>wt/fl</sup> and *Foxp3*<sup>YFP-Cre/wt</sup>*Ogt*<sup>fl/fl</sup> female mice. Data are shown as mean ± s.e.m. \*, p<0.05; \*\*, p<0.01 by unpaired student's *t*-test.



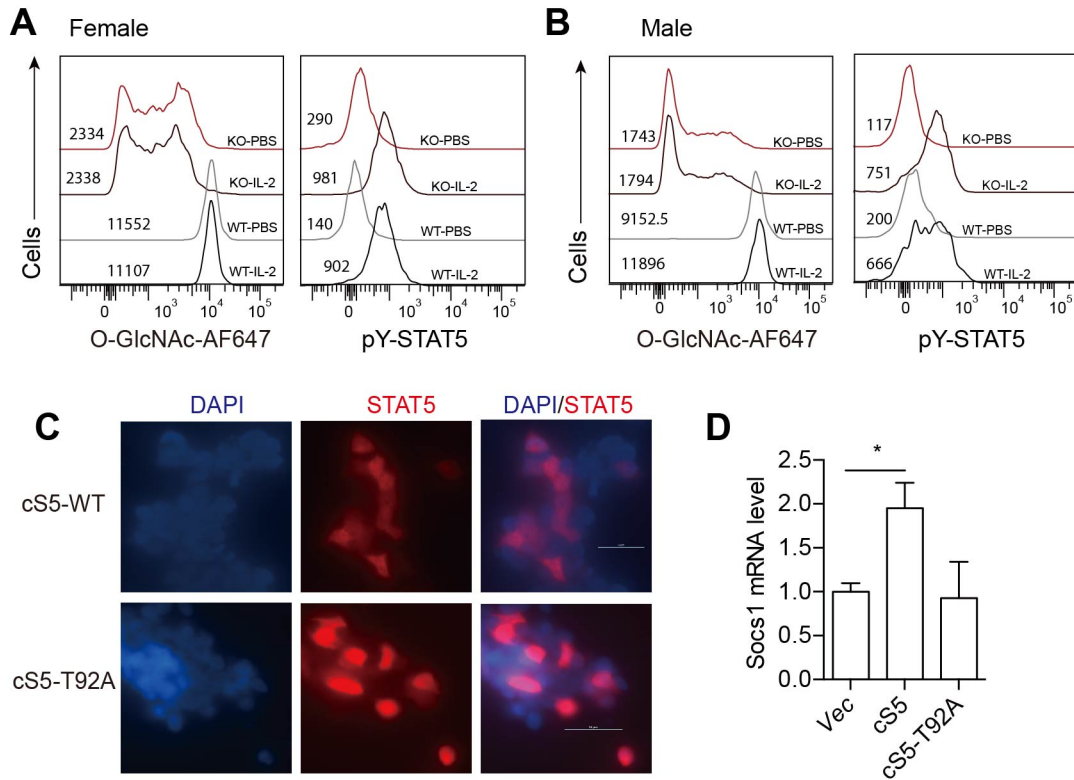
**Supplementary Fig. 5** (A) MFI of O-GlcNAcylation in indicated cell populations from *Foxp3<sup>eGFP-Cre-ERT2/Y</sup> Ogt<sup>wt/Y</sup> Rosa26<sup>tdTomato/wt</sup>* and *Foxp3<sup>eGFP-Cre-ERT2/Y</sup> Ogt<sup>fl/Y</sup> Rosa26<sup>tdTomato/wt</sup>* male mice. (B) Representative images of 6-week-old *Foxp3<sup>eGFP-Cre-ERT2/Y</sup> Ogt<sup>wt/Y</sup> Rosa26<sup>tdTomato/wt</sup>* and *Foxp3<sup>eGFP-Cre-ERT2/Y</sup> Ogt<sup>fl/Y</sup> Rosa26<sup>tdTomato/wt</sup>* male mice fed tamoxifen food for 3 weeks and peripheral LNs and spleen. (C-E) T-BET<sup>+</sup> FOXP3<sup>-</sup> cells (C), GATA3<sup>+</sup> FOXP3<sup>-</sup> cells (D) and RORγT<sup>+</sup> FOXP3<sup>-</sup> cells (E) among CD4<sup>+</sup> T cells in LNs from *Foxp3<sup>eGFP-Cre-ERT2/Y</sup> Ogt<sup>wt/Y</sup> Rosa26<sup>tdTomato/wt</sup>* and *Foxp3<sup>eGFP-Cre-ERT2/Y</sup> Ogt<sup>fl/Y</sup> Rosa26<sup>tdTomato/wt</sup>* male mice, n = 3-4. Data are shown as mean ± s.e.m. \*, p<0.05; \*\*\*, p<0.001 by unpaired student's *t*-test.



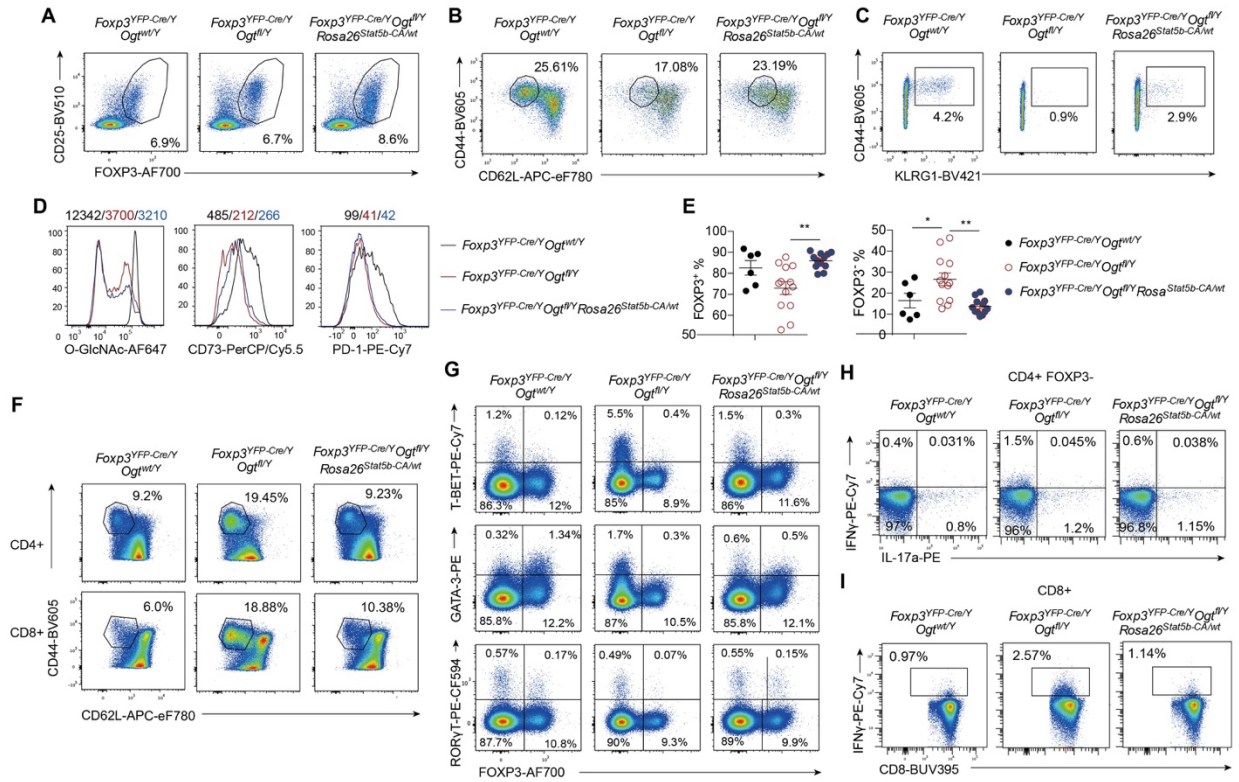
**Supplementary Fig. 6** (A) MFI of pS6 and pS473-AKT among CD4<sup>+</sup>FOXP3<sup>+</sup> Treg cells in LNs of 2-week-old *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>wt/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup>*Ogt*<sup>fl/Y</sup> mice. (B) Suppression activity of OGT-sufficient and OGT-deficient Treg cells, the percentages of T conventional cell proliferation were measured by flow cytometry. (C, D) Body weight (C, n = 12) and representative images of peripheral LNs, spleen and thymus (D) of *Foxp3*<sup>YFP-Cre/wt</sup>*Ogt*<sup>wt/fl</sup> and *Foxp3*<sup>YFP-Cre/wt</sup>*Ogt*<sup>fl/fl</sup> female mice. (E-I) Frequencies of indicated cell populations among YFP<sup>+</sup> Treg cells in the LNs and spleen from *Foxp3*<sup>YFP-Cre/wt</sup>*Ogt*<sup>wt/fl</sup> and *Foxp3*<sup>YFP-Cre/wt</sup>*Ogt*<sup>fl/fl</sup> female mice, n = 3 each group. Data are shown as mean ± s.e.m. \*, p<0.05; \*\*, p<0.01; \*\*\*, p<0.001 by unpaired student's t-test.



**Supplementary Fig. 7** (A) Mice were injected with PBS or the IL-2 immune complex for 3 consecutive days and Treg cells were analyzed at day 6. (B, C) Representative images of peripheral LNs and Spleen(B) and MFI of O-GlcNAcylation among YFP<sup>-</sup>OGT-sufficient or YFP<sup>+</sup>OGT-deficient Treg cells (CD4<sup>+</sup>CD25<sup>+</sup>GITR<sup>+</sup>) (C) from *Foxp3*<sup>YFP-Cre/wt</sup> *Ogf*<sup>fl/fl</sup> female mice injected with PBS or IL2 complex. (D) Frequencies of CD44<sup>+</sup>KLRG1<sup>+</sup> cells among CD4<sup>+</sup>CD25<sup>+</sup>FOXP3<sup>+</sup> Treg cells in LNs and spleen from 2-week-old control and *Foxp3*<sup>YFP-Cre/Y</sup> *Ogf*<sup>fl/Y</sup> male mice injected with PBS or the IL-2 complex, n = 3. (E, F) MFI of CD25 (E) and IL10 (F) among YFP<sup>-</sup>OGT-sufficient or YFP<sup>+</sup>OGT-deficient Treg cells (CD4<sup>+</sup>CD25<sup>+</sup>GITR<sup>+</sup>) from *Foxp3*<sup>YFP-Cre/wt</sup> *Ogf*<sup>fl/fl</sup> female mice injected with PBS or IL2 complex, n = 2-5. Data are shown as mean ± s.e.m. \*\*, p<0.01; \*\*\*, p<0.001 by two-way ANOVA.

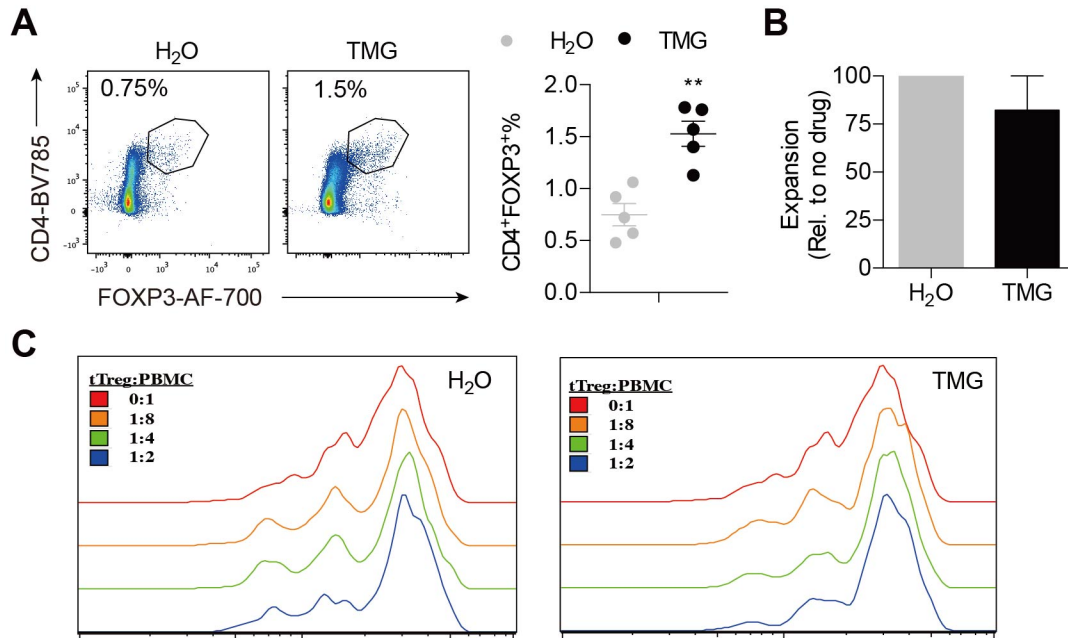


**Supplementary Fig. 8** (A, B) MFI of O-GlcNAcylation and pY-STAT5 among YFP<sup>+</sup>CD25<sup>+</sup> Treg cells in the LNs from *Foxp3*<sup>YFP-Cre/wt</sup> *Ogt*<sup>wt/fl</sup> and *Foxp3*<sup>YFP-Cre/wt</sup> *Ogt*<sup>fl/fl</sup> female mice (A) and *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>wt/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>fl/Y</sup> male mice (B) stimulated with IL-2 in vitro, PBS incubation was used as control. (C) Representative images of STAT5 nuclear translocation in HEK 293 cells transfected with cS5 or cS5-T92A. (D) mRNA levels of *Socs1* in OGT-deficient Treg cells after retrovirus infection as indicated, n = 3. Data are shown as mean ± s.e.m. \*, p<0.05 by one-way ANOVA.

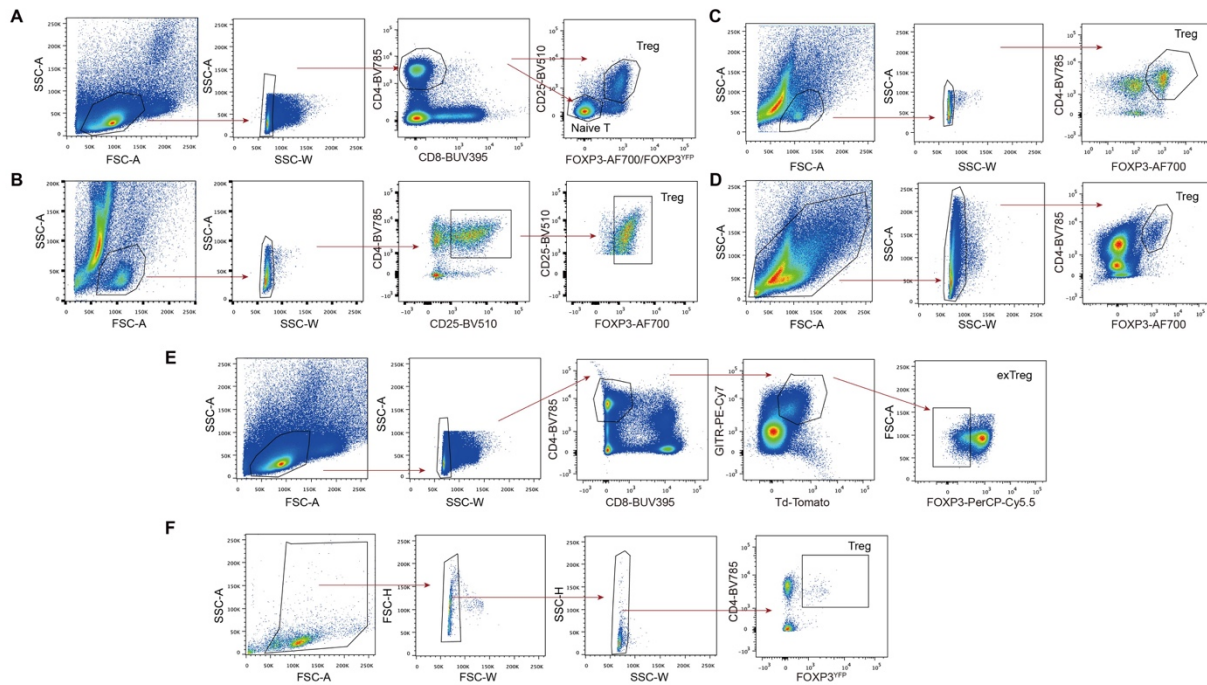


**Supplementary Fig. 9** (A) Flow cytometry of FOXP3<sup>+</sup>CD25<sup>+</sup> Treg cells among CD4<sup>+</sup> T cells in the LNs from *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>wt/Y</sup>, *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>fl/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>fl/Y</sup> *Rosa26*<sup>Stat5b-CA/wt</sup> mice. (B, C) Flow cytometry of CD44<sup>hi</sup>CD62L<sup>lo</sup> eTreg cells (B) and CD44<sup>+</sup>KLRG1<sup>+</sup> eTreg cells (C) among FOXP3<sup>+</sup>CD25<sup>+</sup> Treg cells in the LNs from *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>wt/Y</sup>, *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>fl/Y</sup> and *Foxp3*<sup>YFP-Cre/Y</sup> *Ogt*<sup>fl/Y</sup> *Rosa26*<sup>Stat5b-CA/wt</sup> mice. (D) MFI of O-GlcNAcylation, CD73 and PD-1 among FOXP3<sup>+</sup>CD25<sup>+</sup> Treg cells. (E) Frequencies of FOXP3<sup>+</sup> and FOXP3<sup>-</sup> cells among CD4<sup>+</sup>CD25<sup>+</sup>YFP<sup>+</sup> Treg cells in the LNs, at least n = 6 each group. (F) Flow cytometry of CD44<sup>hi</sup>CD62L<sup>lo</sup> cells among CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the LNs. (G) Flow cytometry of T-BET<sup>+</sup>, GATA3<sup>+</sup> and RORγT<sup>+</sup> population among CD4<sup>+</sup>FOXP3<sup>-</sup> cells in the LNs. (H, I) Flow cytometry of IFNγ-expressing cells among CD4<sup>+</sup>FOXP3<sup>-</sup> (H) and CD8<sup>+</sup> T cells (I) in the LNs. Data are shown as mean ± s.e.m. \*, p<0.05; \*\*, p<0.01 by one-way ANOVA.



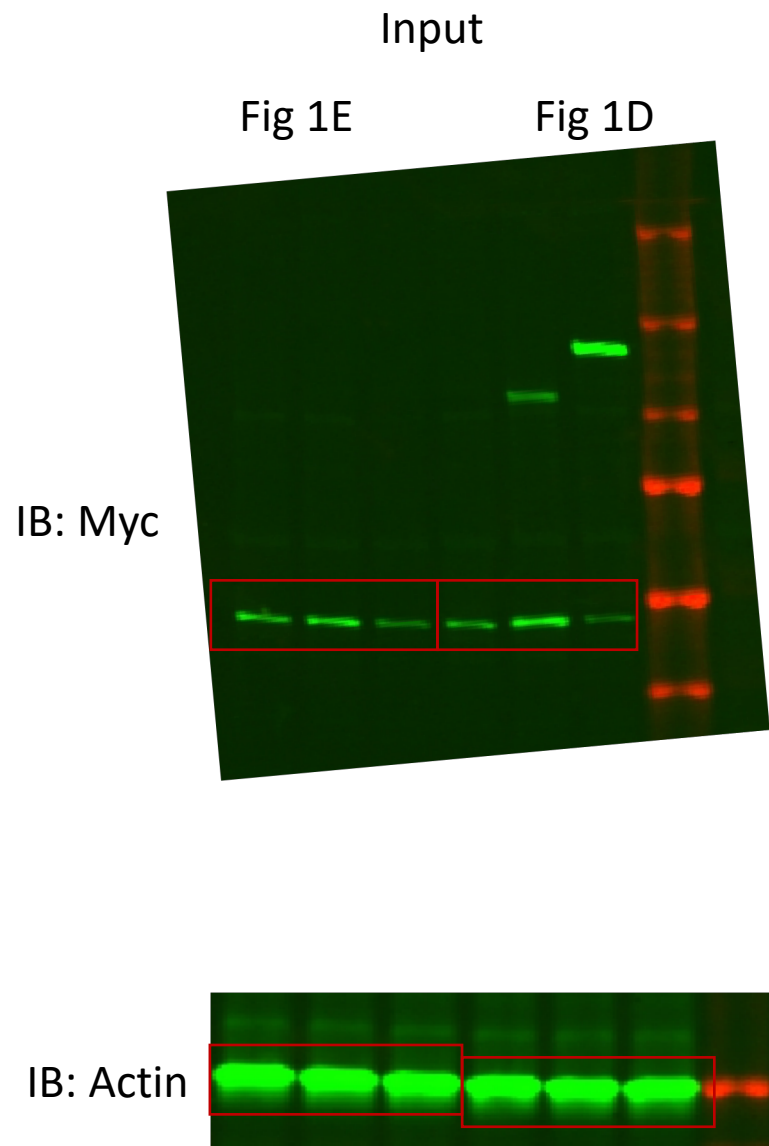
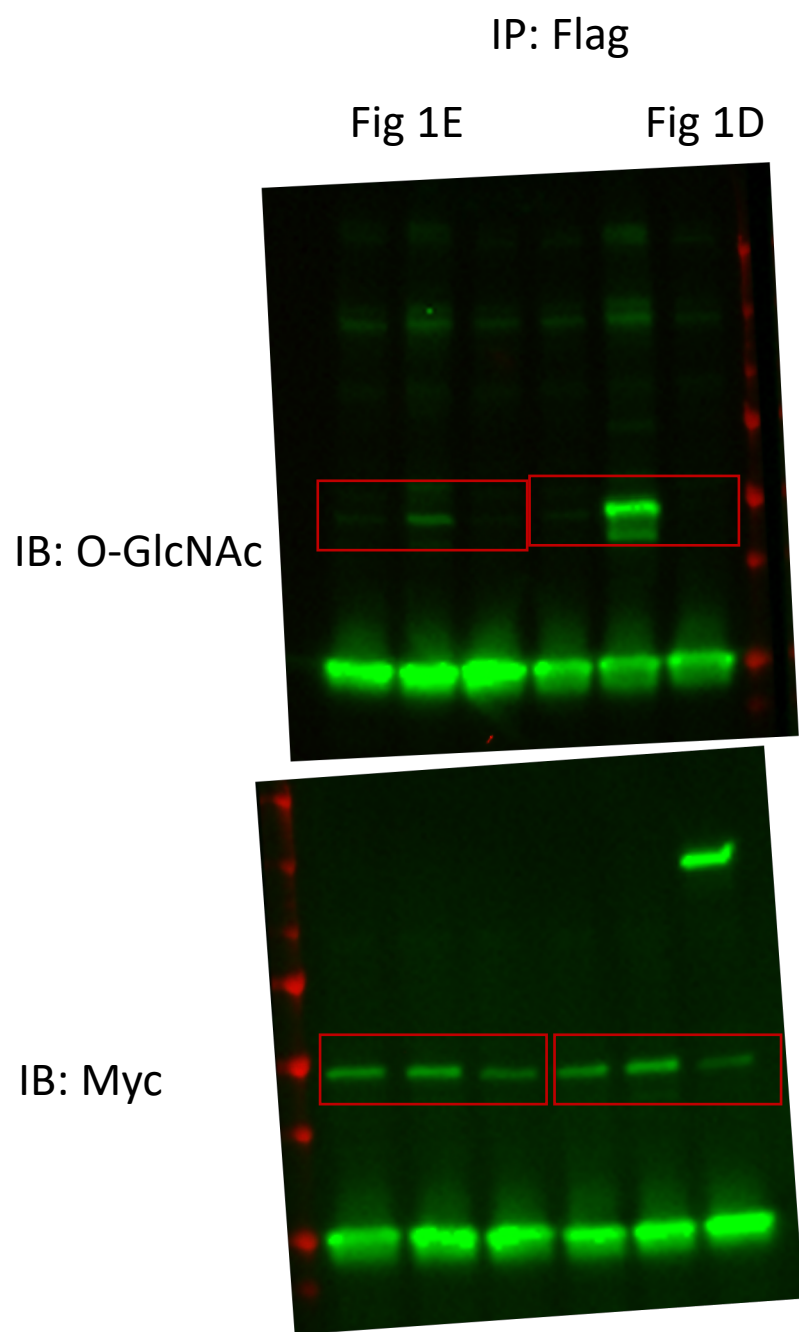


**Supplementary Fig. 10** (A) Flow cytometry and quantification of the frequencies of mouse CD4<sup>+</sup>FOXP3<sup>+</sup> Treg cells treated with TMG, H<sub>2</sub>O treatment was used as a control, n = 5 each group. (B) Frequencies of human tTreg cells expansion treated with or without TMG for 7 days, n=3. (C) Representative plots showing the cell proliferation of CD8<sup>+</sup> cells labeled with CFSE. Data are shown as mean ± s.e.m. \*\*, p<0.01 by unpaired student's t-test.



**Supplementary Fig. 11** (A) Gating strategy to sort naïve T cells ( $CD4^+CD25^-FOXP3^-$ ) and Treg cells ( $CD4^+CD25^+FOXP3^+$ ) from unpurified lymphocytes presented on Fig. 3 & Fig. 7. (B) Gating strategy to sort Treg cells ( $CD4^+CD25^+FOXP3^+$ ) from wildtype mice treated with/without CD3/CD28 beads for in vitro purification and expansion presented on Fig. 1C. (C) Gating strategy to sort Treg cells ( $CD4^+FOXP3^+$ ) from *Ubc-CreER<sup>+</sup>Ogt<sup>fl/y</sup>* and control mice treated with 4-OHT for in vitro purification and expansion presented on Fig. 2A-D or wildtype mice treated with/without TMG for in vitro purification and expansion presented on Fig. 8A-C. (D) Gating strategy to sort Treg cells ( $CD4^+FOXP3^+$ ) from naïve T cells ( $CD4^+CD25^-$ ) infected with retroviruses for in vitro purification and expansion presented on Fig. 2G, H. (E) Gating strategy to sort ex-Treg cells ( $Td\text{-tomato}^+GITR^+FOXP3^-$ ) from *Foxp3<sup>eGFP-Cre-ERT2/y</sup>Ogt<sup>wt/y</sup>Rosa26<sup>tdTomato/wt</sup>* and *Foxp3<sup>eGFP-Cre-ERT2/y</sup>Ogt<sup>fl/y</sup>Rosa26<sup>tdTomato/wt</sup>* mice fed tamoxifen food presented in Fig. 4G-I. (F) Gating strategy to sort Treg cells ( $CD4^+FOXP3^{YFP+}$ ) for RNA-seq presented in Fig. 6A.

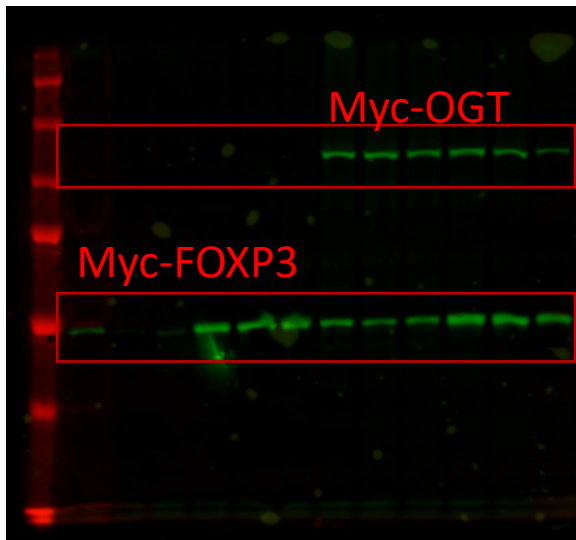




**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

Fig 1F

1<sup>st</sup> day for Myc incubation



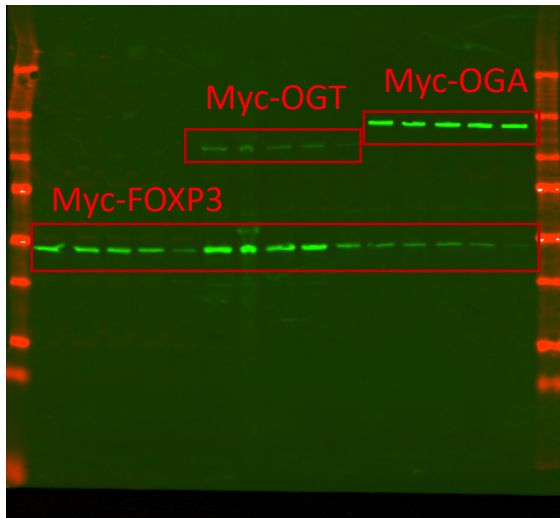
2<sup>nd</sup> day same membrane for Actin incubation



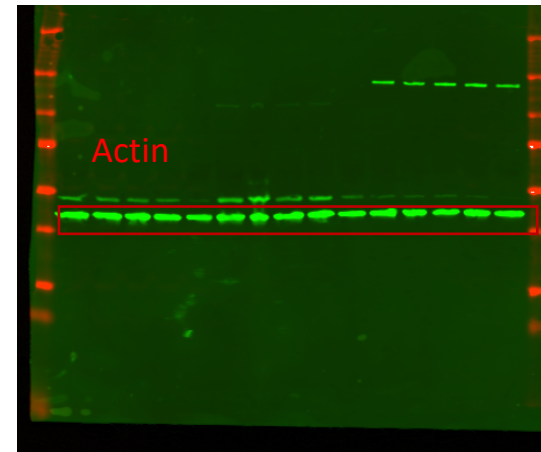
**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

Fig 1G

1<sup>st</sup> day for Myc incubation



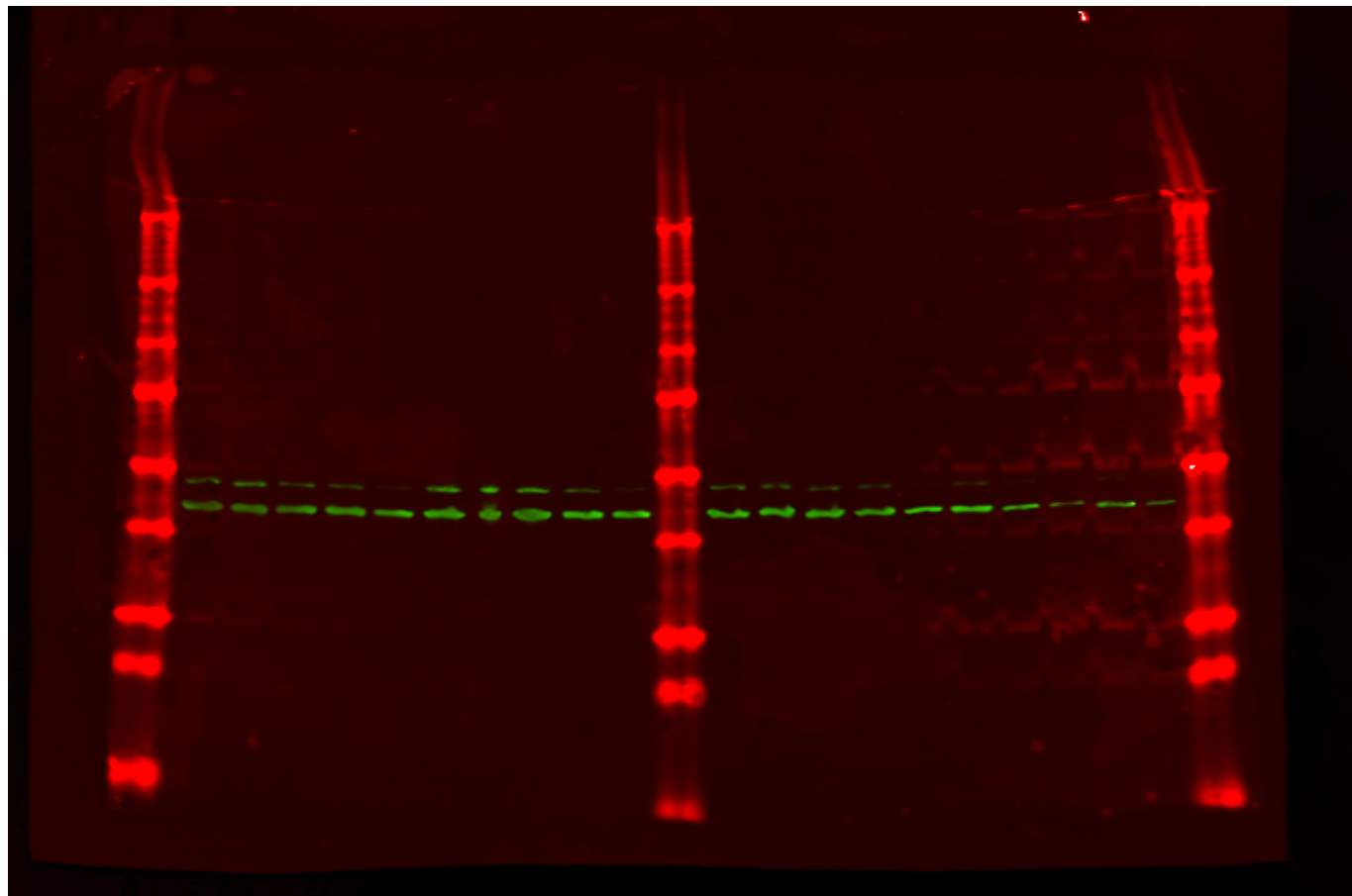
2<sup>nd</sup> day same membrane for Actin incubation



**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

Fig 1H

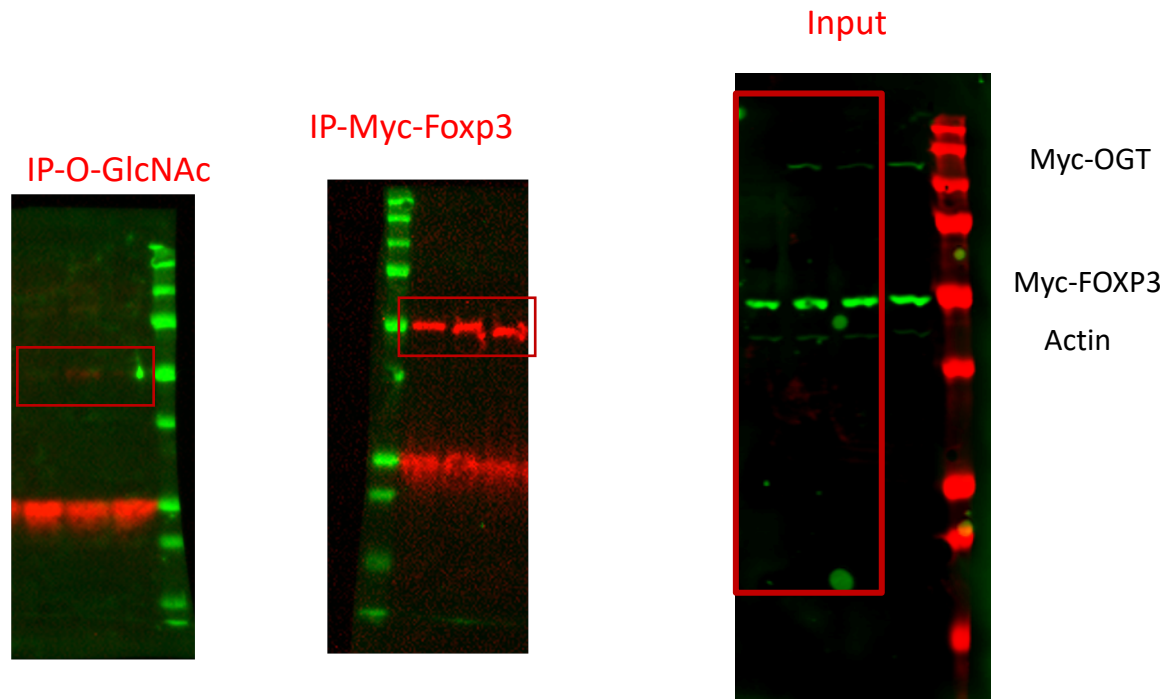
Fig 1I



Up-Myc  
Down-Actin

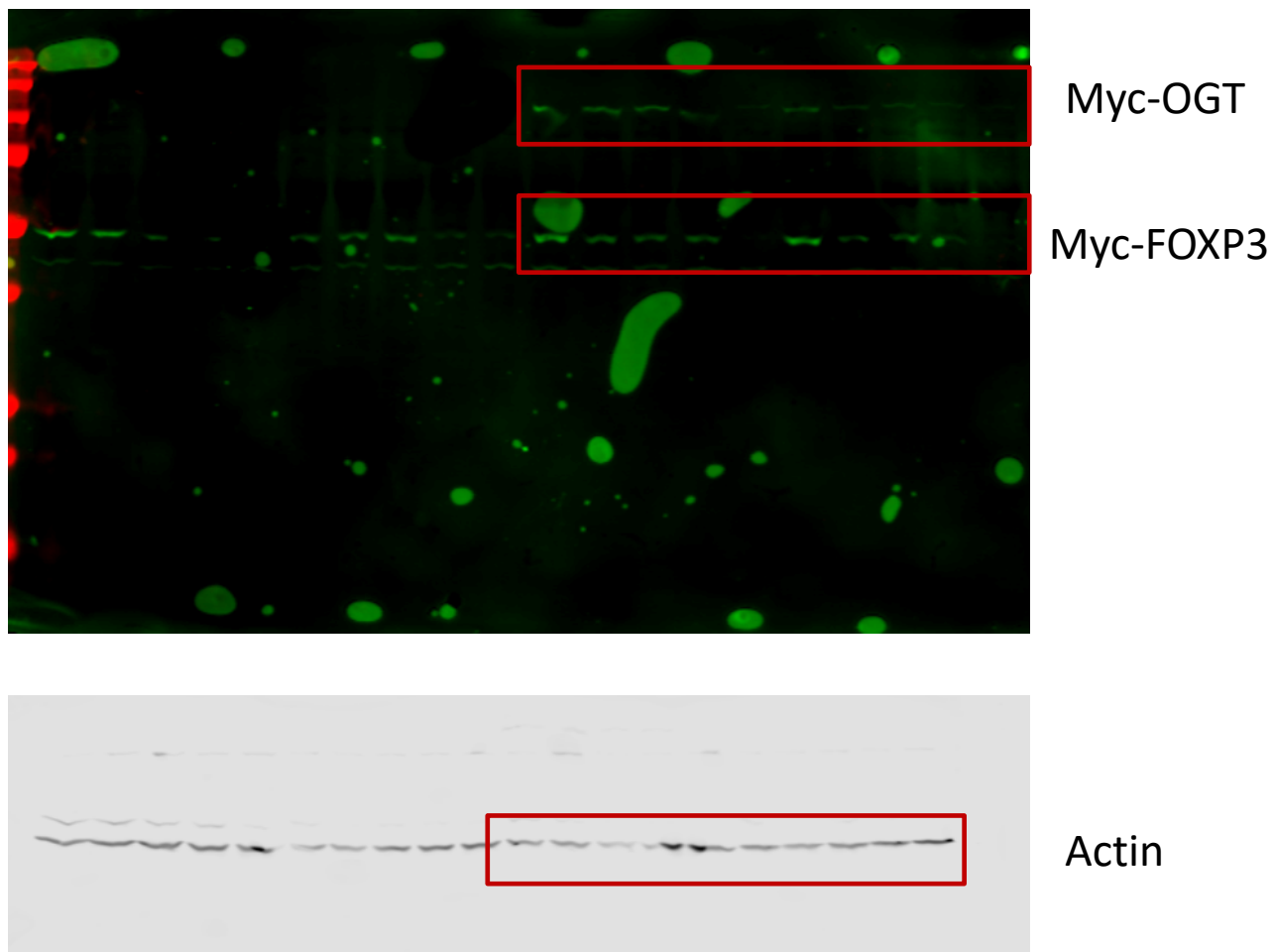
**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

Fig 2E



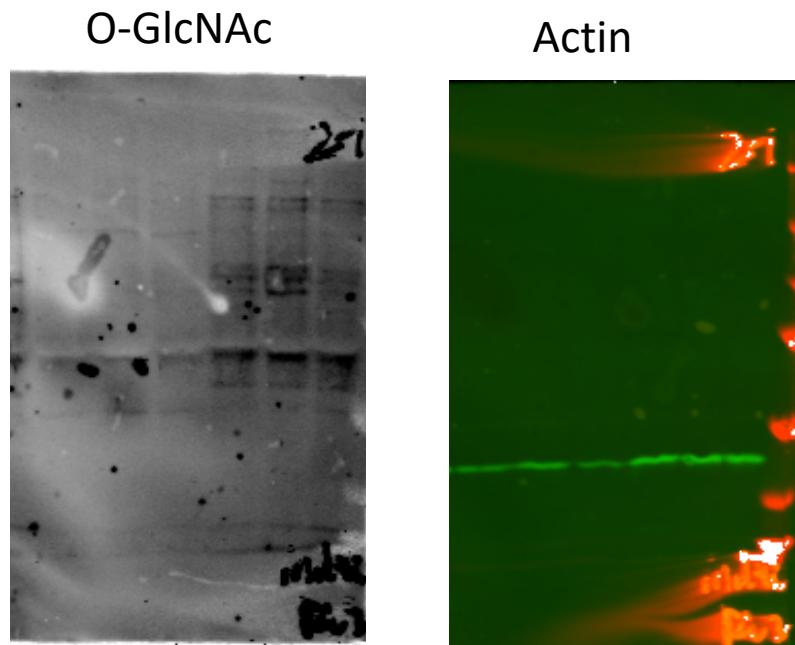
Supplementary Fig 12. Uncropped and unprocessed scans of blots

Fig 2F



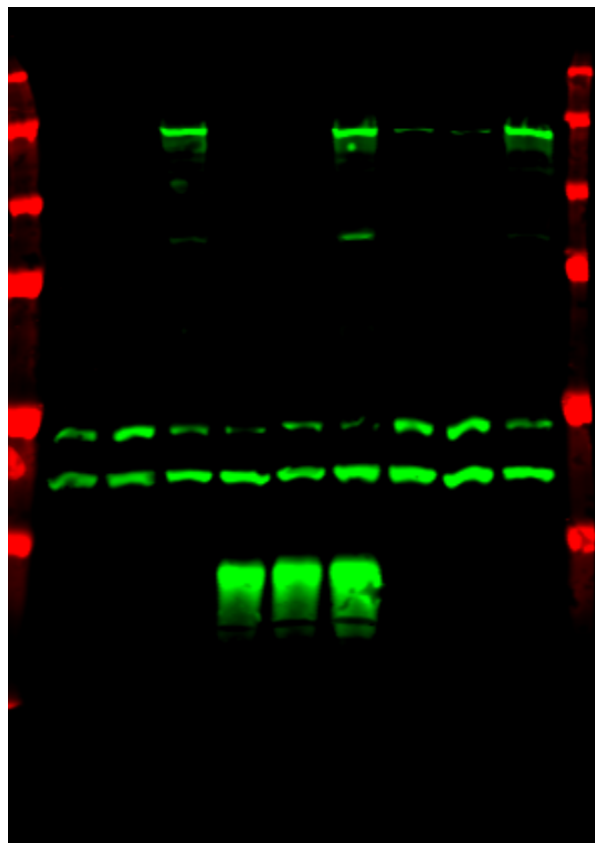
**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

## Supplementary Fig 1B



**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

Supplementary Fig 1C

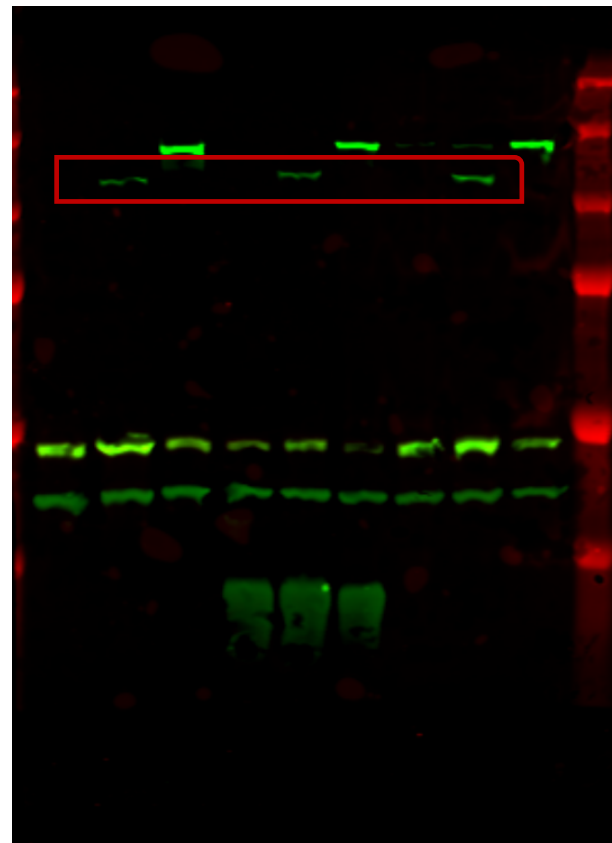


Flag-OGA & Flag-USP7

Flag-FOXP3

Actin

Flag-STUB1



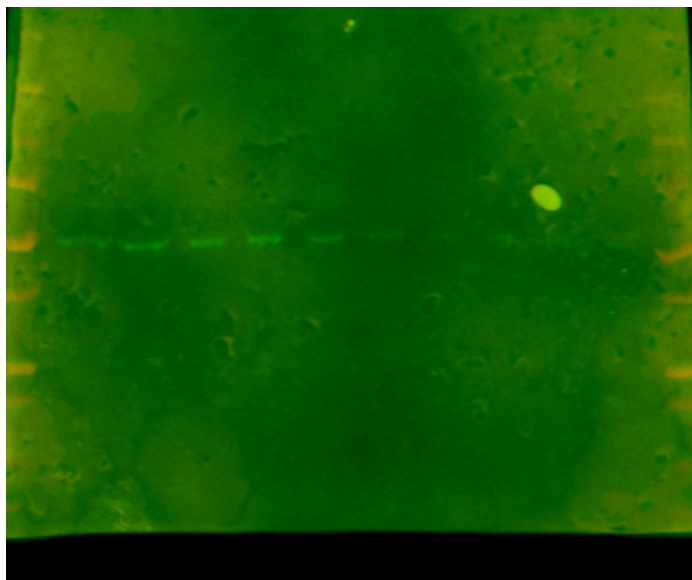
Myc-OGT

**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

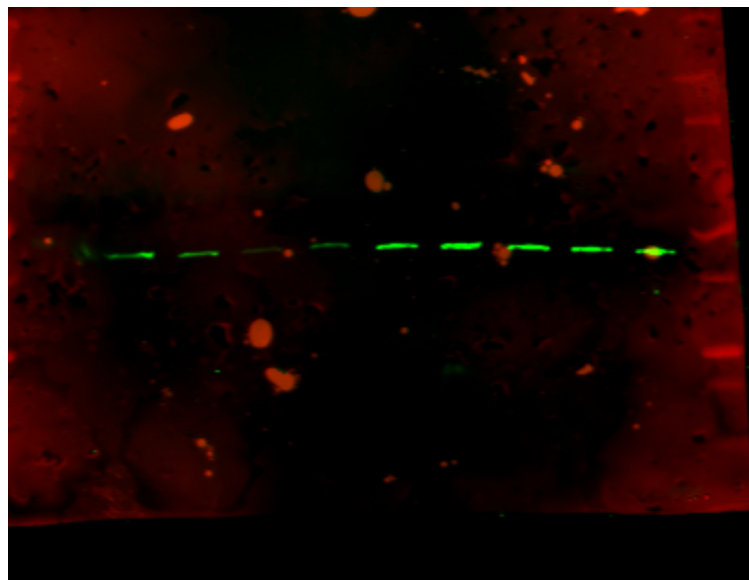


Supplementary Fig 4G

Foxp3



Actin



**Supplementary Fig 12.** Uncropped and unprocessed scans of blots

**Supplementary Table 1**

Primer name	Sequence
Human <i>SOCS1</i> sense	5'-CACGCACTTCCGCACATTC-3'
Human <i>SOCS1</i> antisense	5'-TAAGGGCGAAAAAGCAGTTCC-3'
Human <i>SOCS 3</i> sense	5'-CCTGCGCCTCAAGACCTTC-3'
Human <i>SOCS 3</i> antisense	5'-GTCACTGCGCTCCAGTAGAA-3'
Human <i>18S</i> sense	5'-GCTGGAGGACTCATGTTCAAC -3'
Human <i>18S</i> antisense	5'-CCTTGGGTCAAGTTCACAAGC-3'
Mouse <i>Klrg1</i> sense	5'-TTTGGGGCTTTTGACTGTGAT-3'
Mouse <i>Klrg1</i> antisense	5'-TGTAAGGAGATGTGAGCCTTTGT-3'
Mouse <i>Pd-1</i> sense	5'-ACCCTGGTCATTCATTGGG-3'
Mouse <i>Pd-1</i> antisense	5'-CATTTGCTCCCTCTGACACTG-3'
Mouse <i>Cd44</i> sense	5'-TCGATTTGAATGTAACCTGCCG-3'
Mouse <i>Cd44</i> antisense	5'-CAGTCCGGGAGATACTGTAGC-3'
Mouse <i>Cd73</i> sense	5'-GGACATTTGACCTCGTCCAAT-3'
Mouse <i>Cd73</i> antisense	5'-GGGCACTCGACACTTGGTG-3'
Mouse <i>Socs1</i> sense	5'-CTGCGGCTTCTATTGGGGAC-3'
Mouse <i>Socs1</i> antisense	5'-AAAAGGCAGTCGAAGTCTCG-3'
Mouse <i>Socs3</i> sense	5'-ATGGTCACCCACAGCAAGTTT-3'
Mouse <i>Socs3</i> antisense	5'-TCCAGTAGAATCCGCTCTCCT-3'
Mouse <i>Ogt</i> sense	5'-AAGAGGCACGCATTTTTGAC-3'
Mouse <i>Ogt</i> antisense	5'-ATGGGGTTGCAGTTCGATAG-3'
Mouse <i>18s</i> sense	5'-ACCGCAGCTAGGAATAATGGA-3'
Mouse <i>18s</i> antisense	5'-GCCTCAGTTCCGAAAACCA-3'

**Supplementary Table 2**

Name	Vendor	Catalog no.	Dilution
CD4	BioLegend	100551	1:200
CD4	BioLegend	100548	1:200
CD4	eBioscience	48-0042-82	1:200
CD25	BioLegend	102041	1:200
CD8	BD Biosciences	563786	1:200
CD8	BD Biosciences	563068	1:200
PD-1	eBioscience	25-9985-80	1:200
PD-1	BioLegend	135208	1:200
CD44	BioLegend	103047	1:200
CD44	BioLegend	103012	1:200
CD62L	eBioscience	47-0621-80	1:200
KLRG1	BD Horizon	562897	1:200
Nrp-1	BioLegend	145209	1:200
TCR $\beta$ chain	BD pharmingen	560705	1:200
CD73	BioLegend	127214	1:200
CD103	BioLegend	121406	1:200
CD103	BioLegend	121415	1:200
Helios	BioLegend	137222	1:200
GITR	Thermo Fisher	25-5874-82	1:200
NK-1.1	BioLegend	108708	1:200
CD19	Tonbo Biosciences	65-0193-U100	1:200
CD45R/B220	BD Biosciences	563793	1:200
FOXP3	eBioscience	56-7773-82	1:50
O-GlcNAc	Abcam	201994	1:400
Ki-67	eBioscience	12-5698-82	1:100
T-bet	BioLegend	644824	1:100
GATA-3	eBioscience	12-9966-42	1:100
GATA-3	BD Horizon Biosciences	563349	1:20
ROR $\gamma$ T	BD Horizon	562684	1:100
ROR $\gamma$ T	BD Horizon	564723	1:100
GFP	Thermo Fisher	A-21311	1:400
Annexin V	BD Biosciences	550474	1:100
STAT5 (pY694)	BD Biosciences	612516	1:100
IFN $\gamma$	TONBO biosciences	60-7311-U100	1:100
IL4	BioLegend	504105	1:100
IL17A	BD Biosciences	559502	1:100
IL10	BD Biosciences Pharmingen	554467	1:100
IL5	Caltag Laboratories	RM9064	1:100
IL13	Invitrogen	47-7133-80	1:100
BLIMP-1	BioLegend	150007	1:100
pAKT (Ser473)	Cell Signaling Technology	#4060	1:100
pS6(Ser235, 236)	eBioscience	12-9007-41	1:100
rabbit IgG (H+L)	Life Technologies	A21429	1:100
GZMB	eBioscience	12-8899-71	1:100