

Supporting Information

Tissue-specific kinase expression and activity regulates flux through the pyruvate dehydrogenase complex.

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Table of contents:

Figure S1

Figure S2

Figure S3

Figure S4

Figure S5

Figure S6

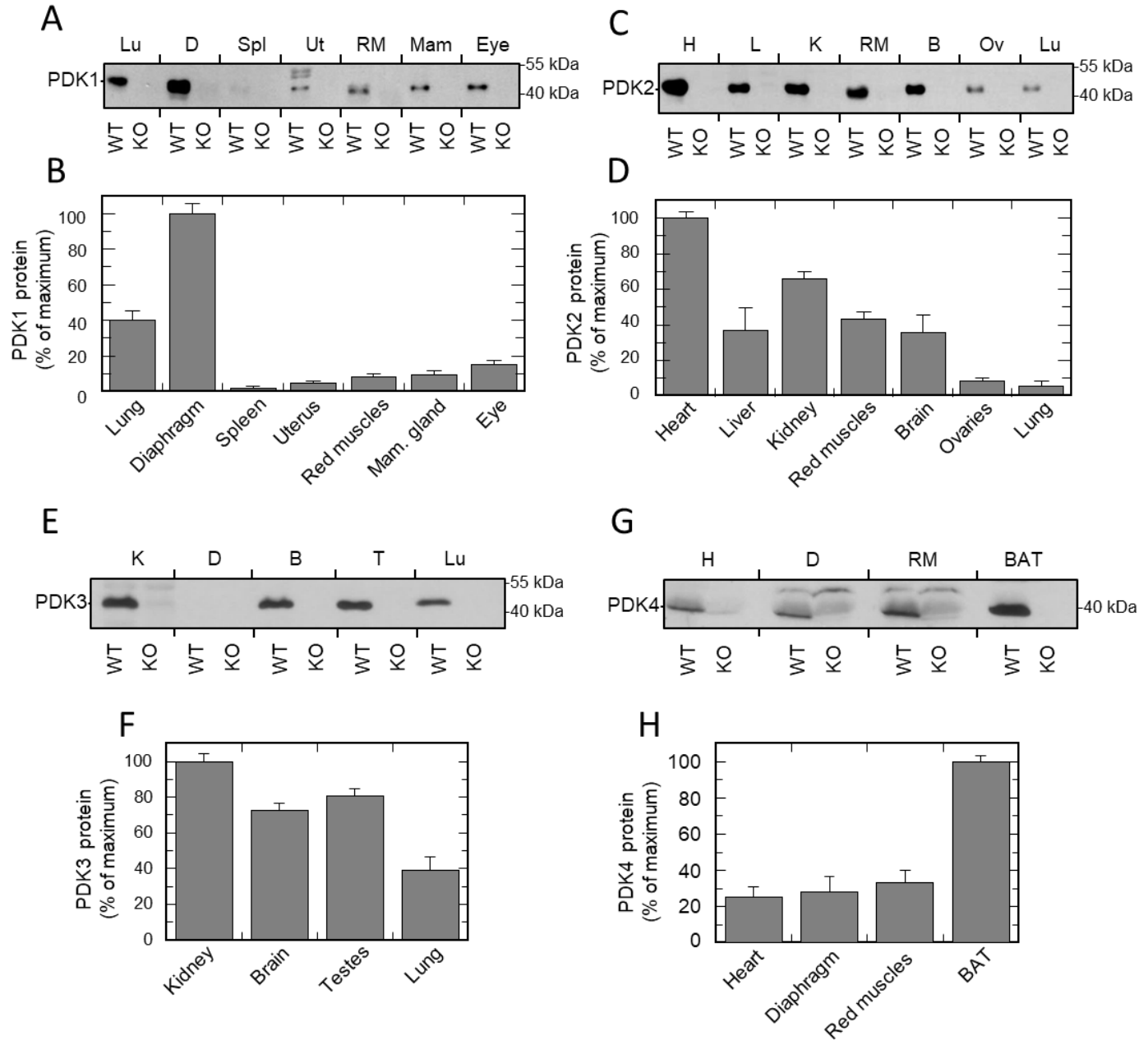


Figure S1. Protein levels of PDK isozymes in mouse tissues. Panels A, C, E, and G, show representative Western blots of PDK1, PDK2, PDK3, and PDK4 protein, respectively. Panels B, D, F, and H illustrate the relative abundance of PDK1, PDK2, PDK3, and PDK4 proteins in mouse tissues calculated based on the results of scanning densitometry. Data are expressed as percent of area of the strongest band. Western blotting was carried out as described under Experimental procedures. Each lane contains 35 μ g of total whole-tissue protein. Data points represent means \pm SD for three mice in each group. Abbreviations are: Lu – lung, D – diaphragm, Spl – spleen, Ut – uterus, RM – red skeletal muscle, Mam – mammary gland, H – heart, L – liver, K – kidney, B – brain, Ov – ovary, BAT – brown adipose tissue.

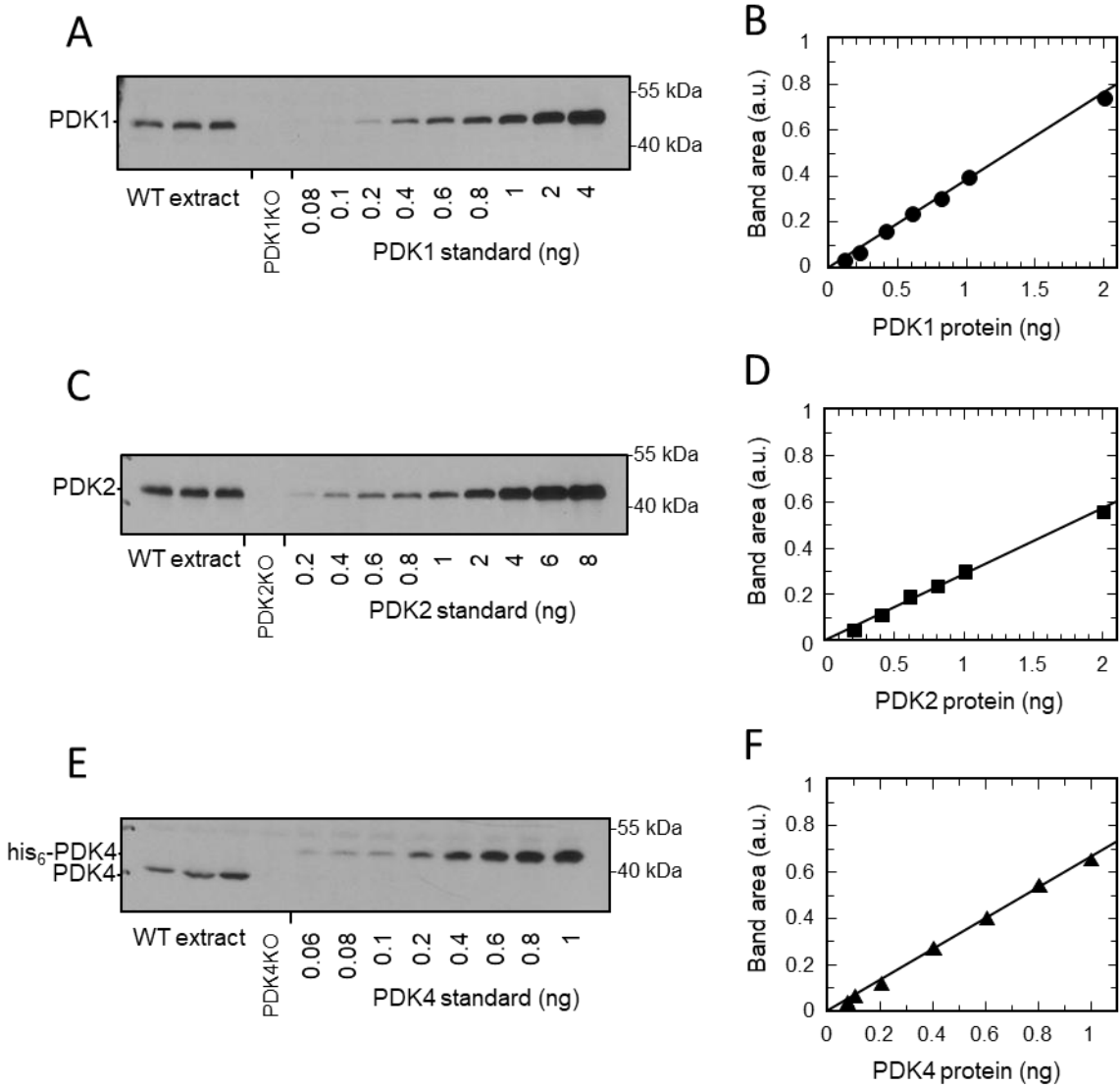


Figure S2. Quantitative Western blot analysis of PDK1, PDK2, and PDK4 in heart muscle. Panels A, C, and E - representative Western blots showing protein levels of PDK1, PDK2, and PDK4 in heart muscle from three animals along with the corresponding protein standards. Samples were prepared as described under Experimental procedures. For PDK1 and PDK2 staining, each lane contains 10 μ g of total protein extracted from hearts of wild-type mice. The lanes with protein standards contain 10 μ g of total protein extract from the appropriate PDK knockout mice. For PDK4, each lane contains 40 μ g of total protein extracted from heart of wild-type mice. The lanes with protein standards contain 40 μ g of total protein extract from PDK4 knockout mice. Panels B, D, and F – calibration curves for measuring protein levels of PDK1, PDK2, and PDK4 in heart muscle constructed based on the results of scanning densitometry of Western blots.

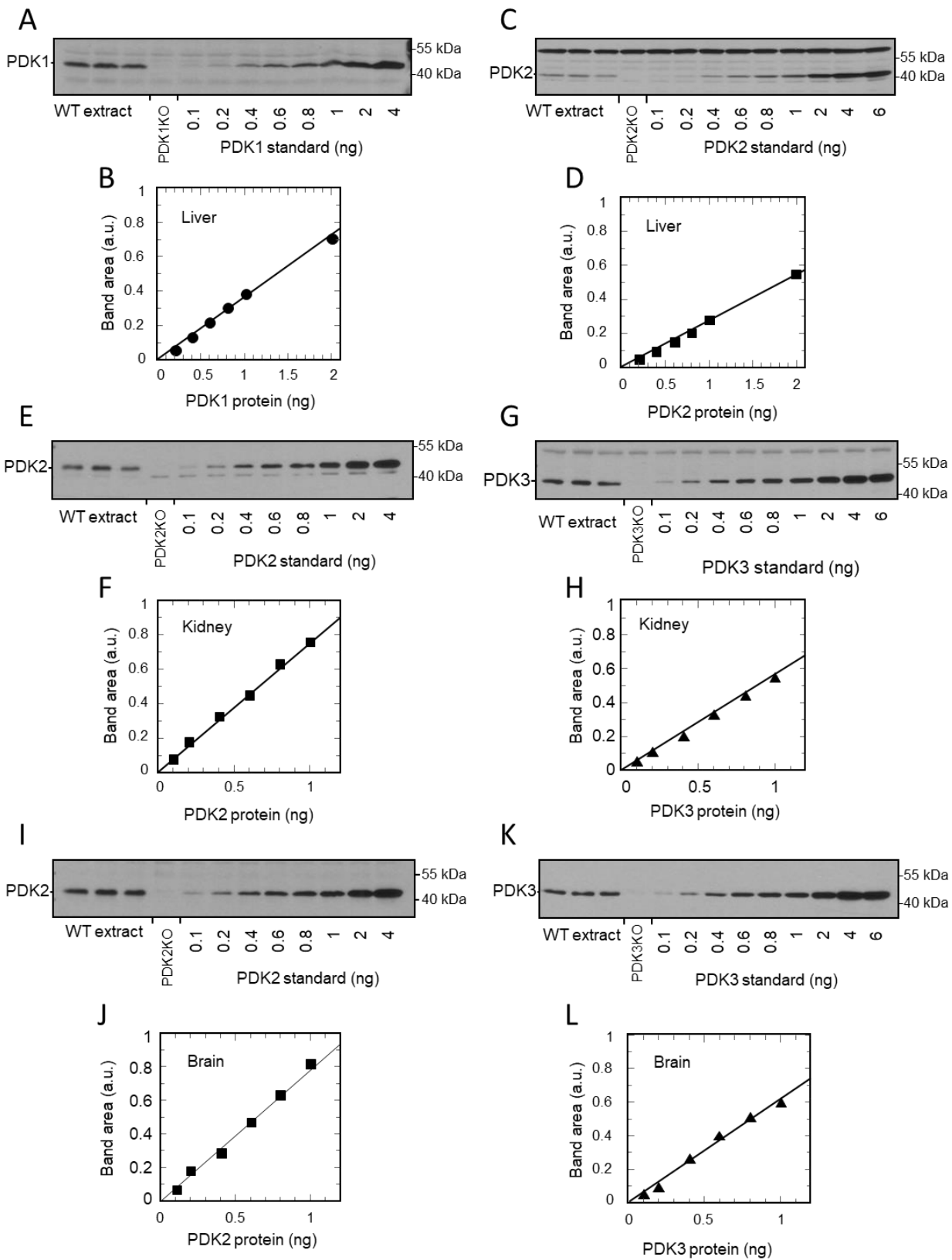


Figure S3. Quantitative Western blot analysis of PDK isozymes in liver, kidney, and brain. Panels A and C - representative Western blots showing protein levels of PDK1 and PDK2 in liver from three animals along with corresponding protein standards. For PDK1 staining, each lane contains 60 μg of total protein extracted from heart of wild-type mice. The lanes with protein standards contain 60 μg of total protein extract from PDK1 knockout mice. For PDK2, each lane contains 30 μg of total protein extracted from heart of wild-type mice. The lanes with protein standards contain 30 μg of total protein extract from PDK2 knockout mice. Panels B and D - calibration curves for measuring protein levels of PDK1 and PDK2 in liver constructed based on the results of scanning densitometry. Panels E and G - representative Western blots depicting protein levels of PDK2 and PDK3 in kidney from three animals along with corresponding protein standards. Each lane contains 20 μg of total protein extracted from kidney of wild-type mice or, for lanes containing protein standards, from appropriate PDK knockout mice. Panels F and H - calibration curves for measuring protein levels of PDK2 and PDK3 in kidney constructed based on the results of scanning densitometry. Panels I and K - representative Western blots illustrating protein levels of PDK2 and PDK3 in brain from three animals along with corresponding protein standards. Each lane contains 20 μg of total protein extracted from brain of wild-type mice or, for lanes containing protein standards, from appropriate PDK knockout mice. Panels J and L - calibration curves for measuring protein levels of PDK2 and PDK3 in brain constructed based on the results of scanning densitometry.

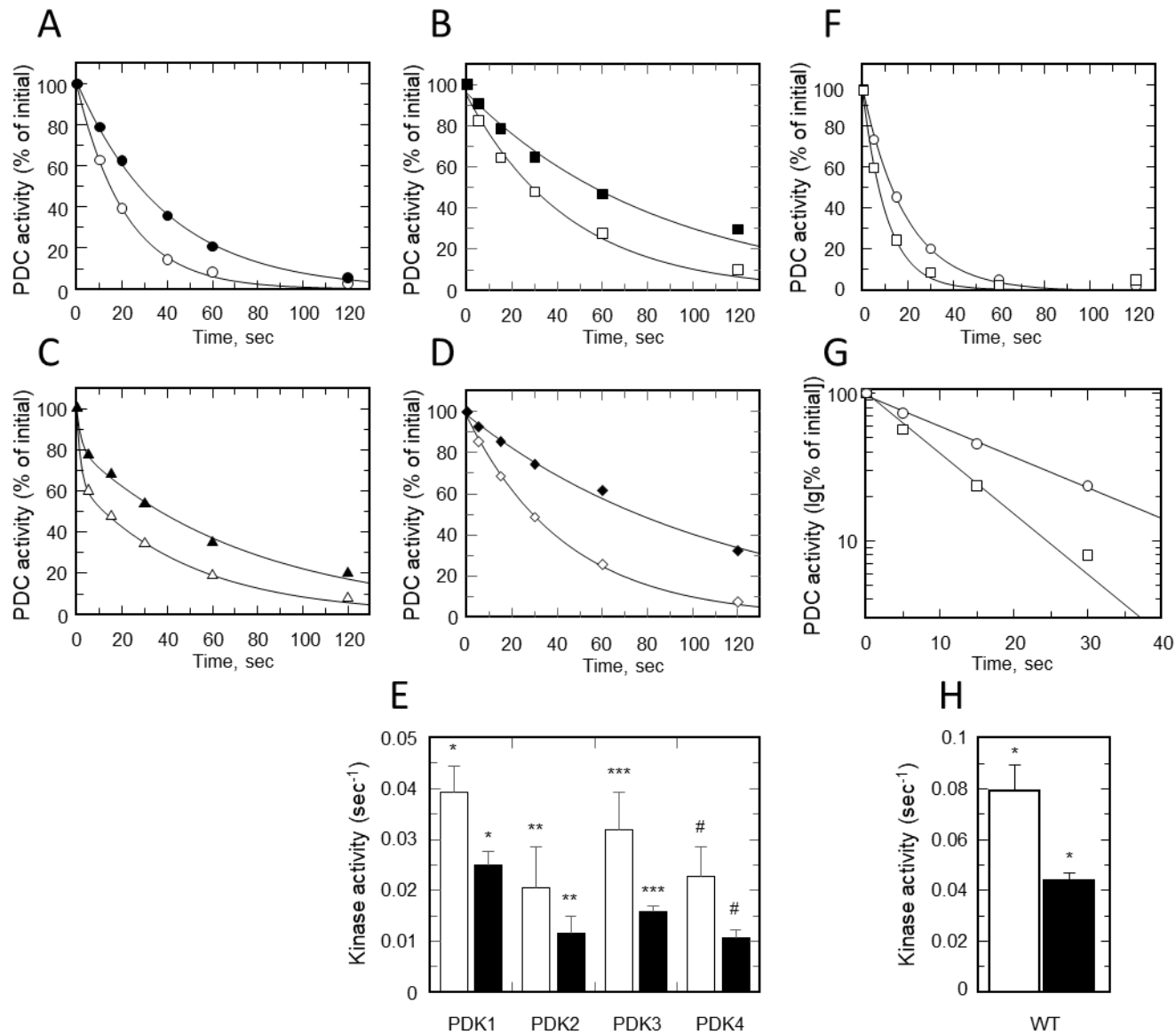


Figure S4. Kinase activity measurements utilizing ATP or ATP γ S as nucleotide substrate. Panels A, B, C, and D - representative ATP- and ATP γ S-dependent inactivation curves (open and closed symbols, respectively) of highly purified, recombinant PDC catalyzed by highly purified, recombinant PDK1, PDK2, PDK3, and PDK4, respectively. Experiments were carried out as described under Experimental procedures. Each phosphorylation reaction received highly purified recombinant PDC at the final protein concentration of 0.5 mg/ml and appropriate isozyme of kinase at the final protein concentration of 3 μ g/ml. Inactivation reactions were initiated by the addition of ATP or ATP γ S at saturating concentration of 0.4 mM. Panel E – kinase activities of recombinant PDK isozymes measured with ATP (open bars) or ATP γ S (closed bars). Activities are expressed in terms of pseudo-first order rate inactivation constants

(sec⁻¹). Data points represent means ± SD for three independent experiments. P<0.05 between groups is considered statistically significant. Groups showing statistically significant differences are indicated by *, **, ***, and # symbols, respectively. Panel F – representative ATP- and ATPγS-dependent inactivation curves of partially purified PDC from heart (open squares or circles, respectively). Panel G - representative ATP- and ATPγS-dependent inactivation curves from Panel F re-plotted in semi logarithmic coordinates. Panel H - kinase activity in preparation of partially purified PDC obtained from heart measured with ATP or ATPγS as nucleotide substrate (open or closed bars, respectively). Data points represent means ± SD for three independent experiments. P<0.05 between groups is considered statistically significant. Groups showing statistically significant differences are indicated by * symbol.

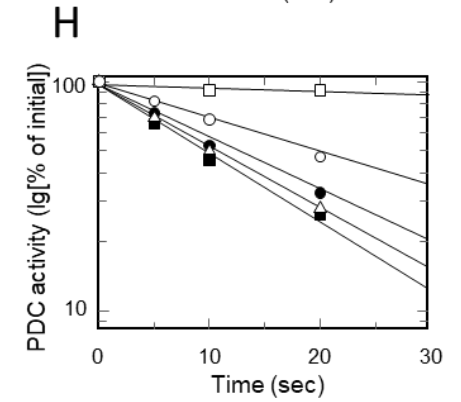
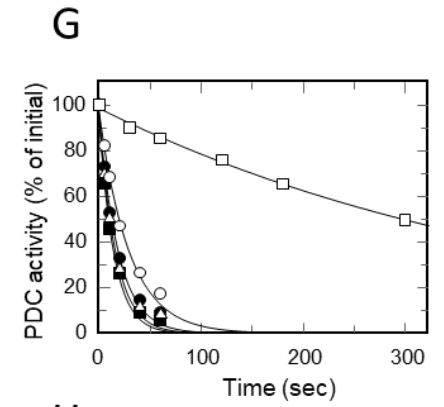
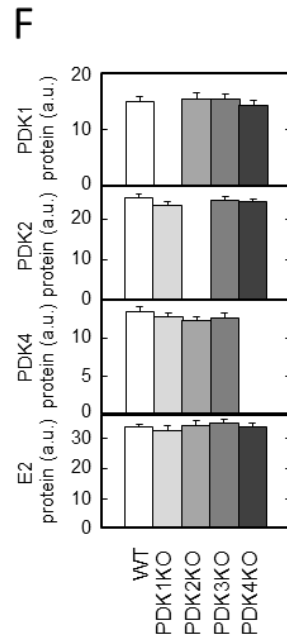
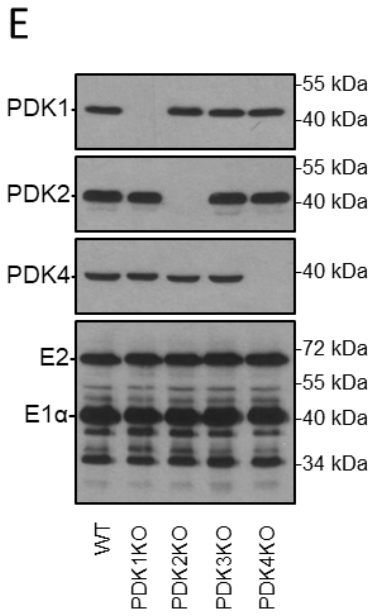
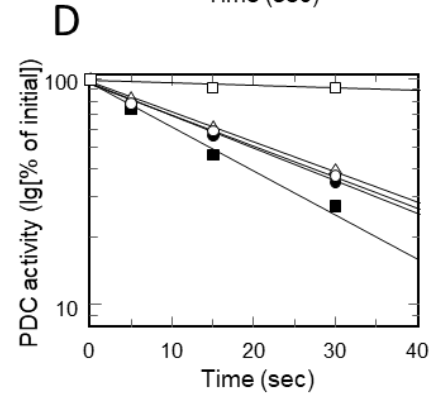
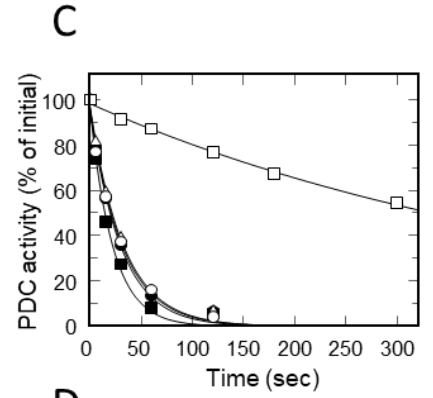
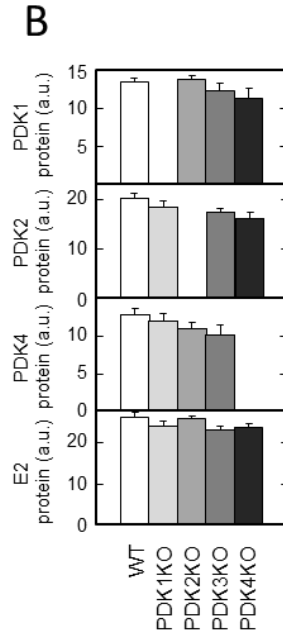
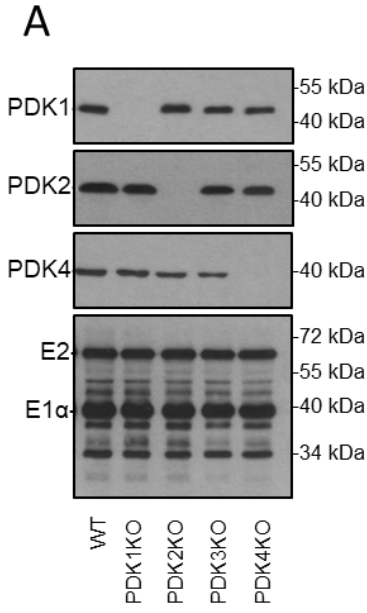


Figure S5. Isozymic composition of PDK and regulation of activity of PDC isolated from diaphragm and skeletal muscle. Panels A and E – representative Western blots illustrating protein levels of PDK1, PDK2, PDK4, as well as E2 and E1 α proteins of PDC in skeletal muscle and diaphragm, respectively. Panels B and F – relative abundance of PDK1, PDK2, PDK4 and E2 proteins in skeletal muscle and diaphragm, respectively, based on the results of scanning densitometry (data points represent means \pm SD for three mice in each group). White, light grey, grey, dark grey and black bars represent samples obtained from wild-type, PDK1 knockout, PDK2 knockout, PDK3 knockout, and PDK4 knockout mice, respectively. Isolation of PDC and Western blotting were carried out as described under Experimental procedures. Each lane contains 15 or 6 μ g of partially purified PDC protein for kinase or PDC staining, respectively. Panels C and G – representative ATP γ S-dependent inactivation curves of partially purified PDC from skeletal muscle and diaphragm of wild-type (dark squares), PDK1 knockout (dark circles), PDK2 knockout (open squares), PDK3 knockout (open triangles), and PDK4 knockout mice (open circles), respectively. ATP-dependent inactivation assay was carried out as described under Experimental procedures. Panels D and H - representative ATP γ S -dependent inactivation curves from Panels C and G re-plotted in semi logarithmic coordinates.

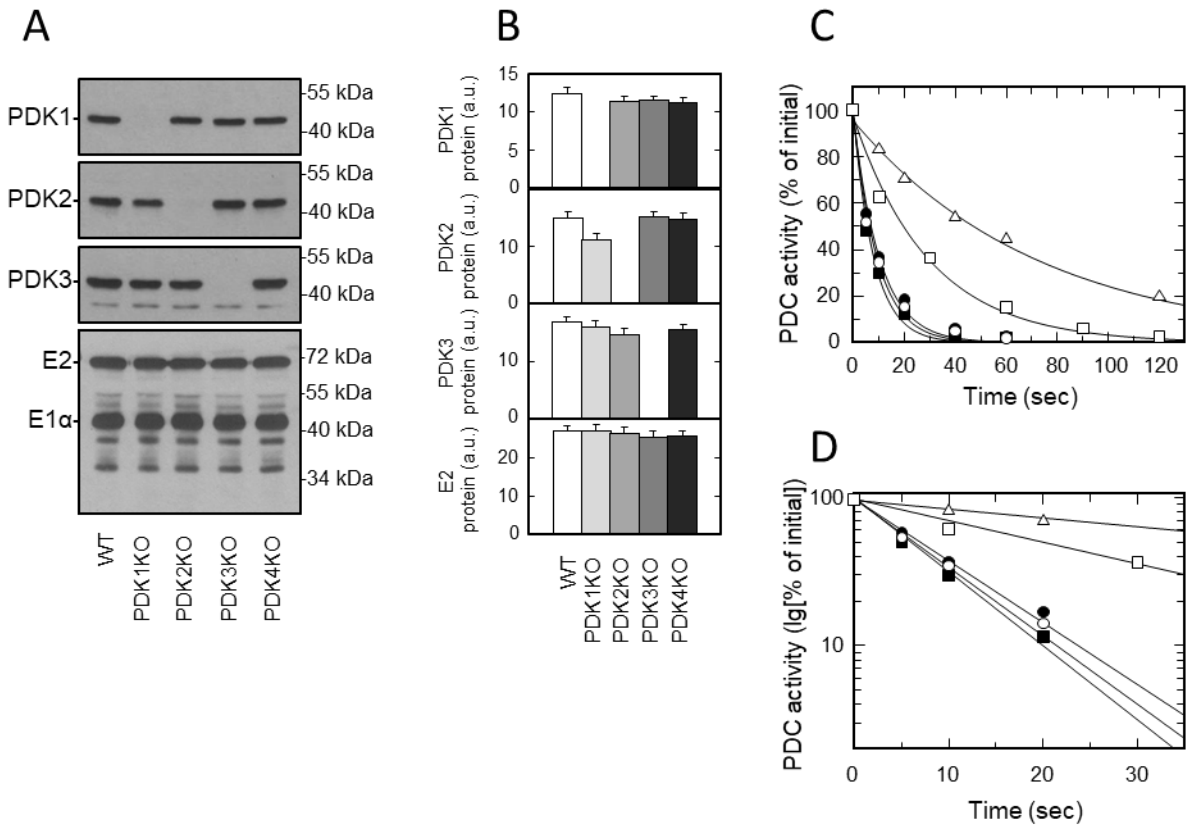


Figure S6. Isozymic composition of PDK and regulation of activity of PDC isolated from brain. Panel A – representative Western blots illustrating protein levels of PDK1, PDK2, PDK3, as well as E2 and E1 α proteins of PDC in brain. Panel B – relative abundance of PDK1, PDK2, PDK3, and E2 proteins based on the results of scanning densitometry (data points represent means \pm SD for three mice in each group). White, light grey, grey, dark grey and black bars represent samples obtained from wild-type, PDK1 knockout, PDK2 knockout, PDK3 knockout, PDK4 knockout mice, respectively. Each lane contains 15 or 6 μ g of partially purified PDC protein for kinase or PDC staining, respectively. Panel C – representative ATP-dependent inactivation curves of partially purified PDC from brain of wild-type (dark squares), PDK1 knockout (dark circles), PDK2 knockout (open squares), PDK3 knockout (open triangles), and PDK4 knockout mice (open circles), respectively. Panel D - representative ATP-dependent inactivation curves from Panel C re-plotted in semi logarithmic coordinates.