

***Thermococcus eurythermalis* endonuclease IV can cleave various apurinic/apyrimidinic site analogues in ssDNA and dsDNA**

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Supplementary materials

Supplementary Figures

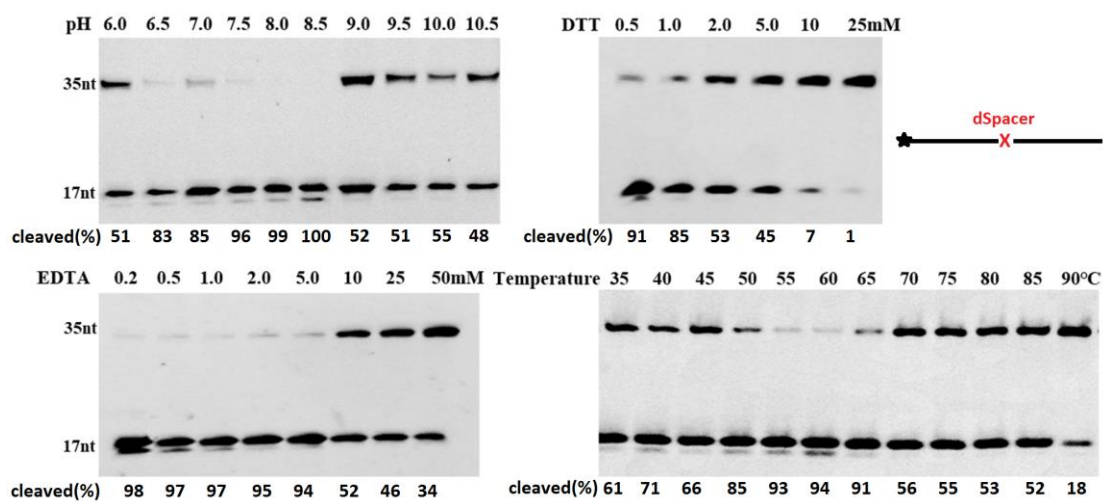


Figure S1 Optimization of reaction conditions. The pH value, concentrations of EDTA and DTT, and reaction temperature were sequentially optimized. The cleavage percentages of substrates are listed at the bottom of each panel.

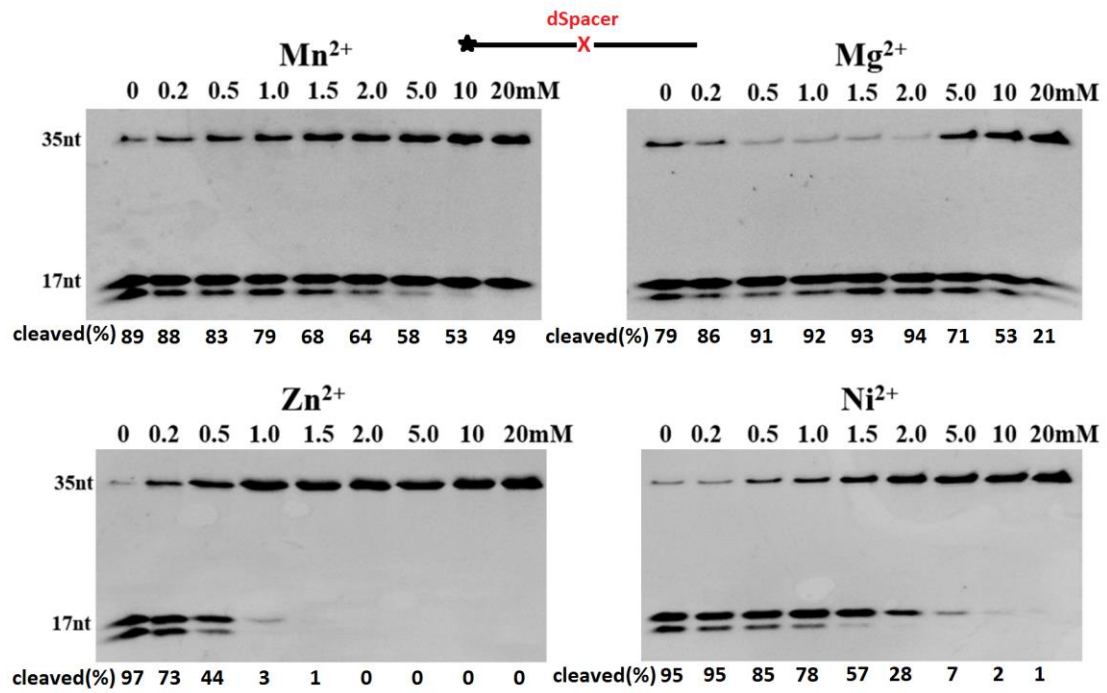


Figure S2 Optimization of divalent metal ions. Different concentrations of Zn²⁺, Mg²⁺, Ni²⁺, and Mn²⁺ were included in the optimized reaction buffer for the assays of AP endonuclease activity. The cleavage percentages of substrates are listed at the bottom of each panel.

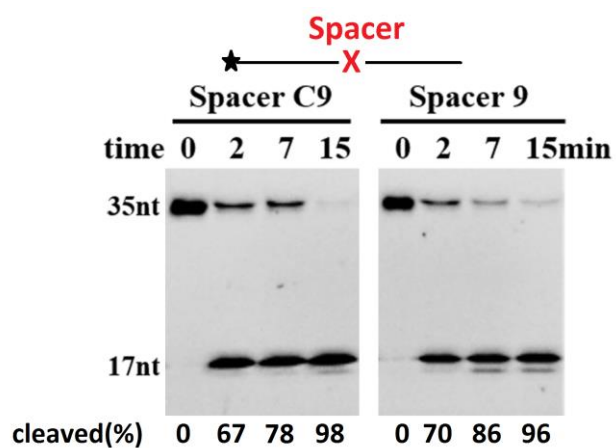


Figure S3 Cleavage of ssDNAs containing Spacer C9 or 9. SsDNAs (100 nM) containing an internal Spacer C9 or Spacer 9 were incubated with TeuendoIV (5 nM) at 55°C for the indicated time in the optimized reaction buffer. The cleavage percentages of substrates are listed at the bottom of each panel.

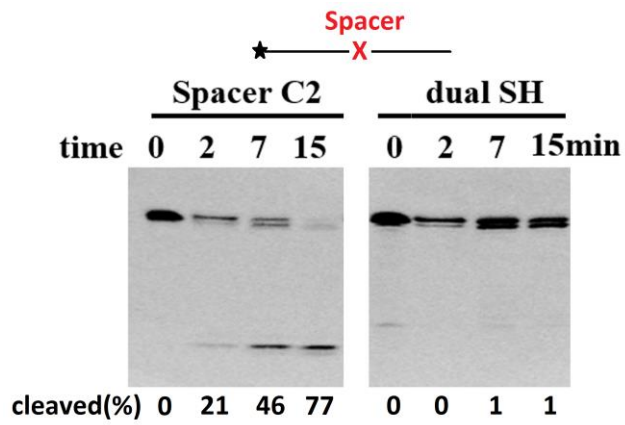


Figure S4 Cleavage of ssDNAs containing single Spacer C2 or Dual SH. SsDNAs (100 nM) containing one Spacer C2 or Dual SH were incubated with TeuendoIV (50 nM) at 55°C for the indicated time. The cleavage percentages of substrates are listed at the bottom of each panel.

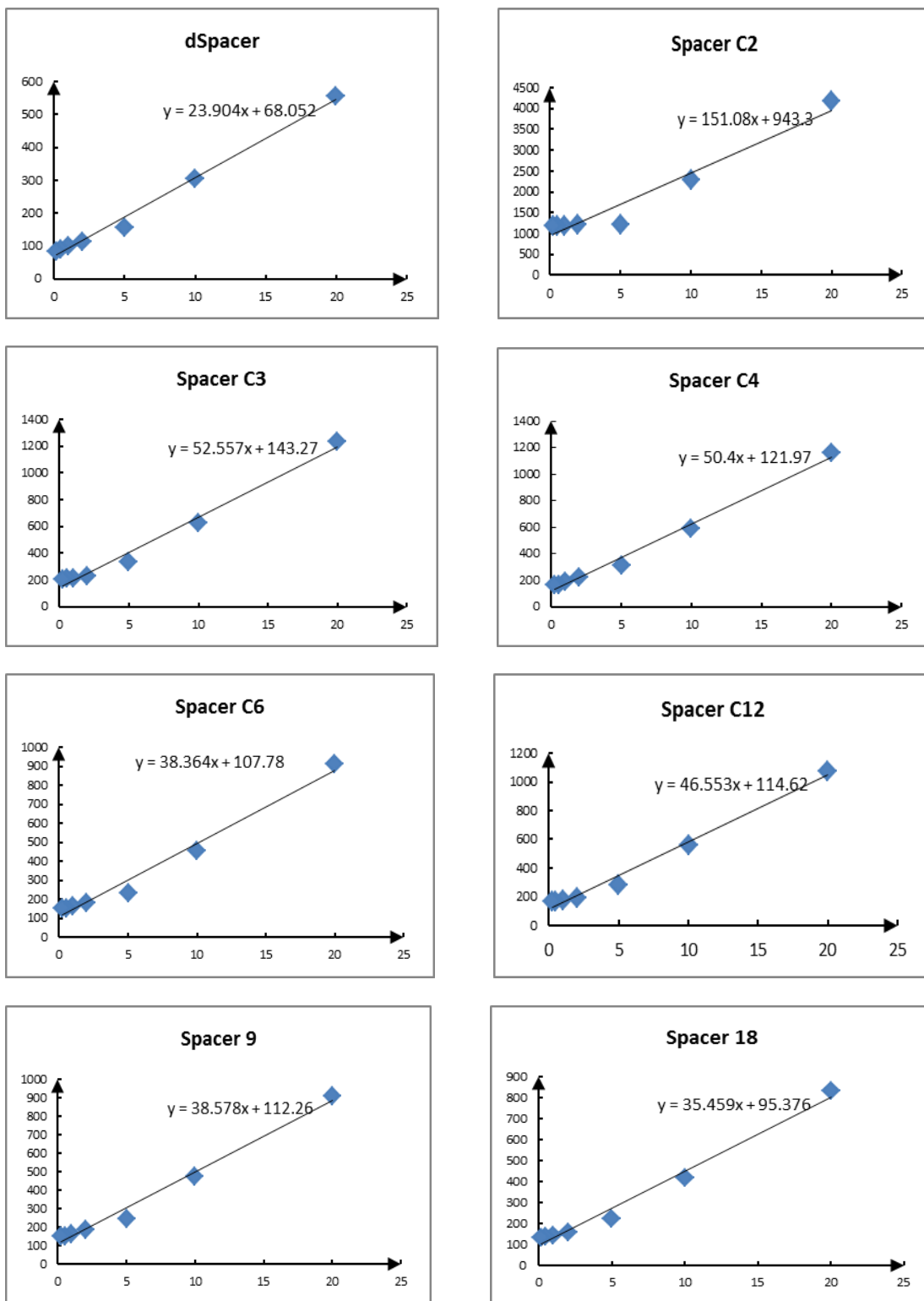


Fig. S5 Graphs of the double reciprocal plotting of various AP-site analogues.

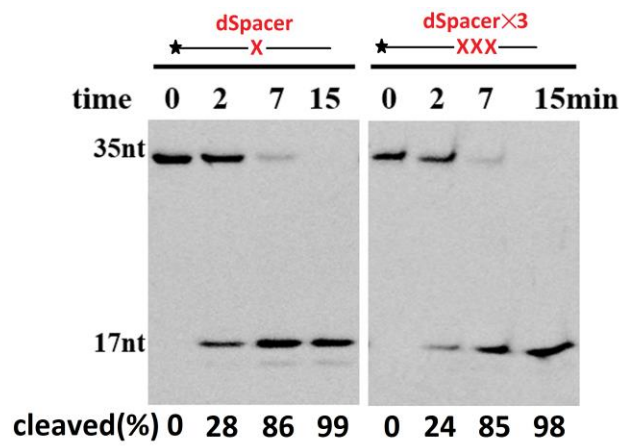


Figure S6 Cleavage of ssDNAs containing one or three clustered dSpacers. SsDNAs (100 nM) containing one or three internal dSpacers were incubated with TeuendoIV (5 nM) at 55°C for the indicated time in the optimized reaction buffer. The cleavage percentages of substrates are listed at the bottom of each panel.

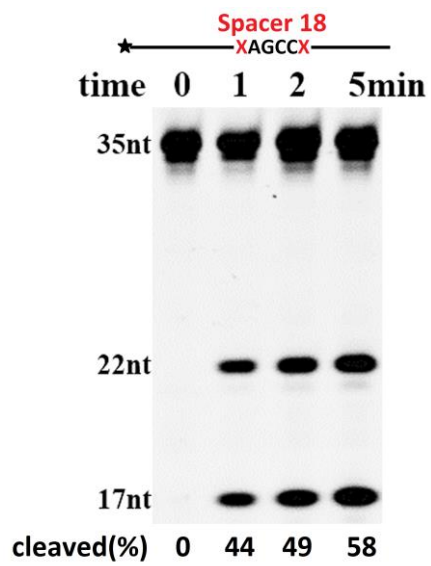


Figure S7 Cleavage of ssDNA containing two Spacer 18s separated by four normal nucleotides. The ssDNAs (100 nM) were incubated with TeuendoIV (2 nM) at 55°C for the indicated time in the optimized reaction buffer. The cleavage percentages of substrates are listed at the bottom of each panel.

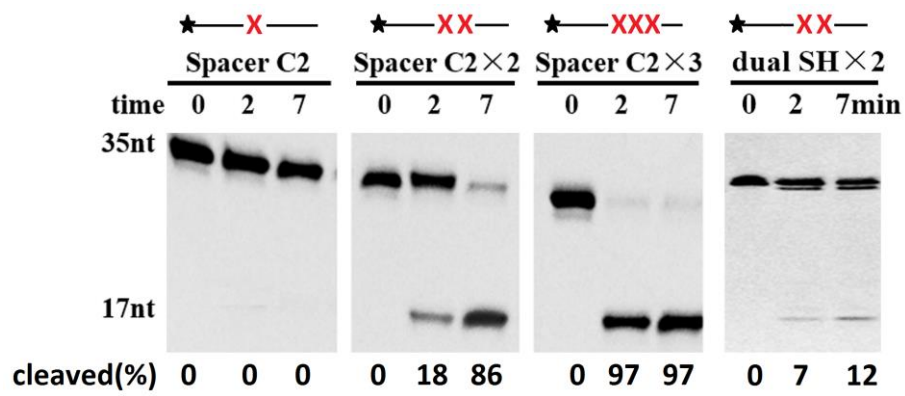


Figure S8 Cleavage of ssDNAs containing one or clustered Spacer C2 and dual SH. SsDNAs (100 nM) containing 1-3 internal Spacer C2s or dual SH were incubated with TeuendoIV (5 nM for Spacer C2 and 50 nM for Spacer dual SH) at 55°C for the indicated time. The cleavage percentages of substrates are listed at the bottom of each panel.

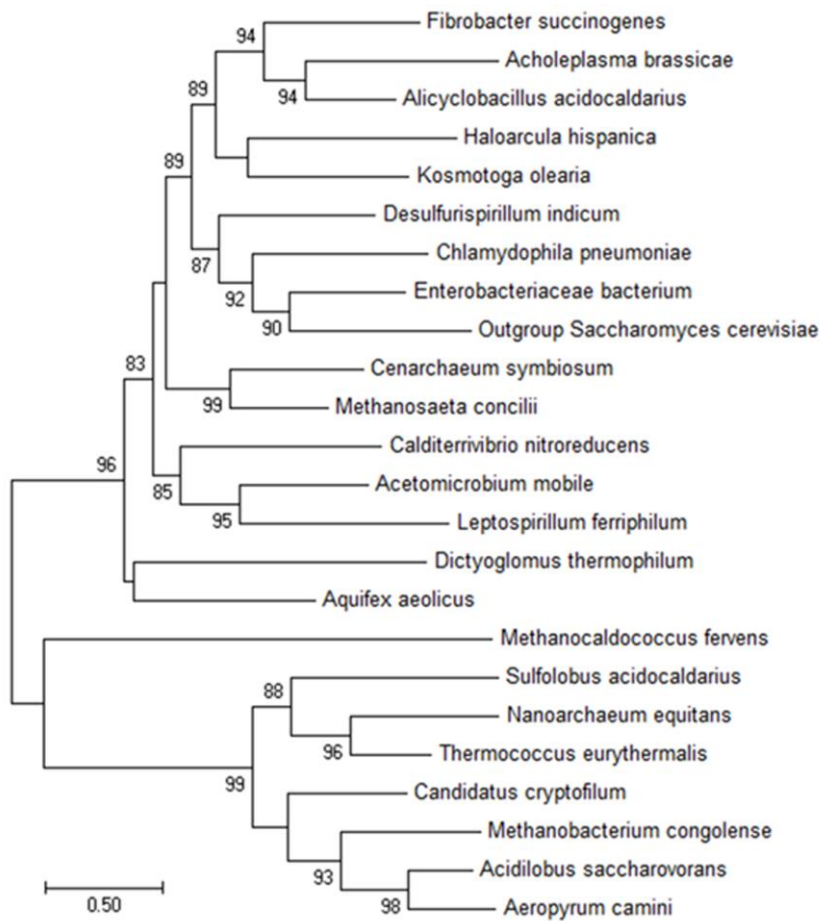


Figure S9 Phylogenetic tree of EndoIV. Selected EndoIV from typical bacteria and archaea were used to construct the phylogenetic tree. The bootstrap values are listed at the branch.

Supplementary Tables

Table S1 Oligonucleotides used for constructing plasmids

Oligos	Sequences (5' – 3') ^a	Comments
TeuendoIV-F	tgaaac catatg aaagtacttctcctc	TeuEndoIV
TeuendoIV-R	ttgcttc gggatcc aaaag cac	
H70A-F	ctgct cacagctGCc gcgccctact acat	H70A
H70A-R	agtagggcgc gGCagctgtg agcaggacg	
Y73A-F	tcac gcgcccGCct acatcaacct caacg	Y73A
Y73A-R	g aggttgatgt agGCgggcgc gtgagctg	
N76A-F	ccctact acatcGCcct caacgcgagc ga	N76A
N76A-R	tcgcgttg aggGCgatgt agtagggcgc g	
H110A-F	agcgt cgttttcGCc gccggctact acct	H110A
H110A-R	agtagccggc gGCgaaaacg acgctccag	
R231A-F	a gggcgagaag GCgcacctga acctccag	R231A
R231A-R	aggt tcaggtgcGC cttctcgccc ttatc	
H232N-F	gcgagaag aggGCcctga acctccagga g	H232N
H232N-R	tggaggt tcaggGCcct cttctcgccc tt	

^a The changed bases are shown in uppercase letters.

Table S2 Oligonucleotides used for analyzing enzymatic activity

Sequences (5'-3') ^a	Damages	Figures
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=dU	1b
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer C2	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer C3	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer C4	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer C6	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer C9	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer C12	3,4,5, S1-3,S4,S6-7
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=dSpacer	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer 9	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Spacer 18	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=Dual SH	
*CAGCCAGGTGTCTCACTXAGCCGACTCGCCACAGT	X=S-S	
*CAGCCAGGTGTCTCACTXXGCCGACTCGCCACAGT	X=Spacer C12X2	
*CAGCCAGGTGTCTCACTXXGCCGACTCGCCACAGT	X=Spacer 18X2	
*CAGCCAGGTGTCTCACTXXXCCGACTCGCCACAGT	X=Spacer 18X3	4, S5
*CAGCCAGGTGTCTCACTXXXXXXCTCGCCACAGT	X=Spacer 18X7	
*CAGCCAGGTGTCTCACTXYGCCGACTCGCCACAGT	X=Spacer 18, Y=Spacer C12	
*CAGCCAGGTGTCTCACTYXGCCGACTCGCCACAGT	X=Spacer 18, Y=Spacer C12	
*CAGCCAGGTGTCTCACTXAGCCXACTCGCCACAGT	X=Spacer 18	
*CAGCCAGGTGTCTCACTXYXCCGACTCGCCACAGT	X=Spacer 18, Y=Spacer C12	
*CAGCCAGGTGTCTCACTXYXCCGACTCGCCACAGT	X=Spacer 18, Y=Spacer 9	
*CAGCCAGGTGTCTCACTXYXCCGACTCGCCACAGT	X=Spacer 18, Y=Spacer C2	
*CAGCCAGGTGTCTCACTXXXCCGACTCGCCACAGT	X=dSpacer	S4
*CAGCCAGGTGTCTCACTXXGCCGACTCGCCACAGT	X=Spacer C2	S6
*CAGCCAGGTGTCTCACTXXXCCGACTCGCCACAGT	X=Spacer C2	
*CAGCCAGGTGTCTCACTXXGCCGACTCGCCACAGT	X=Dual SH	S7
TXAGCCGACTCGCCACAGT*	X=Spacer C6	
CTXAGCCGACTCGCCACAGT*	X=Spacer C6	
CACTXAGCCGACTCGCCACAGT*	X=Spacer C6	
TCTCACTXAGCCGACTCGCCACAGT*	X=Spacer C6	
*CAGCCAGGTGTCTCACTXA	X=Spacer C6	
*CAGCCAGGTGTCTCACTXAG	X=Spacer C6	6
*CAGCCAGGTGTCTCACTXAGCC	X=Spacer C6	
*CAGCCAGGTGTCTCAXXAGCCGACTCGCCACAGT	X=Spacer C6	
*CAGGTGTCTCACTXXX	X=Spacer C6	
*CAGGTGTCTCACTXXXC	X=Spacer C6	
*CAGGTGTCTCACTXXXCCG	X=Spacer C6	
*CAGCCAGGTGTCTCACTX	X=Spacer C3	
*CAGCCAGGTGTCTCACTX	X=Spacer C6	7
*CAGCCAGGTGTCTCACTX	X=Spacer C12	

*CAGCCAGGTGTCTCACT

ACTGTGGCGAGTCGGCTAAGTGAGACACCTGGCTG	complementary strand	
ACTGTGGCGAGTCGGCTTAGTGAGACACCTGGCTG	complementary strand	2b,5,7,8
ACTGTGGCGAGTCGGCTCAGTGAGACACCTGGCTG	complementary strand	
ACTGTGGCGAGTCGGCTGAGTGAGACACCTGGCTG	complementary strand	

^a Asterisks denote the fluorescein (6-FAM) group at the 5'- or 3'-end. The damaged bases (AP site analogues or dU) are denoted by the letters X and Y in the sequences and are annotated in the "Damage" column. The damage-containing strands are shown in blue with the Spacer damage in red, and the complementary strands are shown in black.

Table S3 EndoIV used for constructing phylogenetic tree

Strain name	Protein Name	Gene locus	Lineage
<i>Acidilobus saccharovorans</i>	Endonuclease IV	ASAC_1468	Archaea
<i>Aeropyrum camini</i>	Endonuclease IV	ACAM_1316	Archaea
<i>Korarchaeum cryptofilum</i>	Endonuclease IV	Kcr_0075	Archaea
<i>Cenarchaeum symbiosum</i>	Endonuclease IV	CENSYa_1925	Archaea
<i>Haloarcula hispanica</i>	Endonuclease IV	HAH_2520	Archaea
<i>Methanobacterium congolense</i>	Putative endonuclease 4	MCBB_0201	Archaea
<i>Methanocaldococcus fervens</i>	Putative endonuclease 4	Mefer_1095	Archaea
<i>Methanosaeta concilii</i>	Apurinic-apyrimidinic endonuclease	MCON_1877	Archaea
<i>Nanoarchaeum equitans</i>	NEQ077a	NEQ077a	Archaea
<i>Sulfolobus acidocaldarius</i>	Endonuclease IV	Saci_0015	Archaea
<i>Fibrobacter succinogenes</i>	Apurinic endonuclease Apn1	FSU_2936	Bacteria
<i>Acetomicrobium mobile</i>	Apurinic endonuclease APN1	Anamo_0665	Bacteria
<i>Acholeplasma brassicae</i>	Endodeoxyribonuclease IV	BN85311130	Bacteria
<i>Alicyclobacillus acidocaldarius</i>	Deoxyribonuclease IV	Aaci_1017	Bacteria
<i>Aquifex aeolicus</i>	Endonuclease IV	Aq_1629	Bacteria
<i>Calditerrivibrio nitroreducens</i>	Deoxyribonuclease IV	Calni_1384	Bacteria
<i>Chlamydophila pneumoniae</i>	Endonuclease IV	CP_0014	Bacteria
<i>Desulfurispirillum indicum</i>	Apurinic endonuclease Apn1	Selin_1546	Bacteria
<i>Dictyoglomus thermophilum</i>	Endonuclease IV	DICTH_1988	Bacteria
<i>Enterobacteriaceae bacterium</i>	Endonuclease IV	F652_1793	Bacteria
<i>Kosmotoga olearia</i>	Apurinic endonuclease Apn1	Kole_0273	Bacteria
<i>Leptospirillum ferriphilum</i>	Apurinic endonuclease	LFML04_0897	Bacteria
<i>Saccharomyces cerevisiae</i>	APN2	YBL019W	Eukaryota

Table S4 The maximum values of product amount and the initial rates during the incubation period

Substrate concentration (μM)		0.05	0.1	0.2	0.5	1.0	2.0	5.0
dSpacer	Cleaved substrate (%)	18.0	16.5	16.0	8.9	5.1	2.8	1.2
	Initial rate ($\mu\text{M}/\text{min}$)	0.0018	0.0033	0.0064	0.0089	0.01	0.011	0.012
Spacer C2	Cleaved substrate (%)	14.0	13.2	12.2	4.9	2.5	1.3	0.5
	Initial rate ($\mu\text{M}/\text{min}$)	0.00023	0.00044	0.00081	0.00082	0.00083	0.00087	0.00083
Spacer C3	Cleaved substrate (%)	8.1	8.1	7.3	4.4	2.4	1.2	0.5
	Initial rate ($\mu\text{M}/\text{min}$)	0.00081	0.0016	0.003	0.0044	0.0048	0.0048	0.005
Spacer C4	Cleaved substrate (%)	8.6	8.4	8.0	4.5	2.7	1.5	0.6
	Initial rate ($\mu\text{M}/\text{min}$)	0.00086	0.0017	0.0032	0.0045	0.0054	0.006	0.006
Spacer C6	Cleaved substrate (%)	11.1	11.0	10.6	5.6	3.1	1.6	0.7
	Initial rate ($\mu\text{M}/\text{min}$)	0.0011	0.0022	0.0043	0.0056	0.0062	0.0065	0.0067
Spacer C12	Cleaved substrate (%)	9.3	9.2	9.2	5.2	2.9	1.5	0.6
	Initial rate ($\mu\text{M}/\text{min}$)	0.00093	0.0018	0.0036	0.0052	0.0058	0.0061	0.006
Spacer 9	Cleaved substrate (%)	11.0	10.5	10.2	5.4	3.1	1.6	0.7
	Initial rate ($\mu\text{M}/\text{min}$)	0.0011	0.0021	0.0041	0.0054	0.0062	0.0065	0.0067
Spacer 18	Cleaved substrate (%)	11.9	11.8	11.3	6.4	3.5	1.8	0.8
	Initial rate ($\mu\text{M}/\text{min}$)	0.0012	0.0024	0.0045	0.0064	0.007	0.0073	0.0076

The percentage of substrate converted to product during the incubation period, and the initial rates at the increasing substrate concentrations of various AP-site analogues are listed.