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Supplemental Information

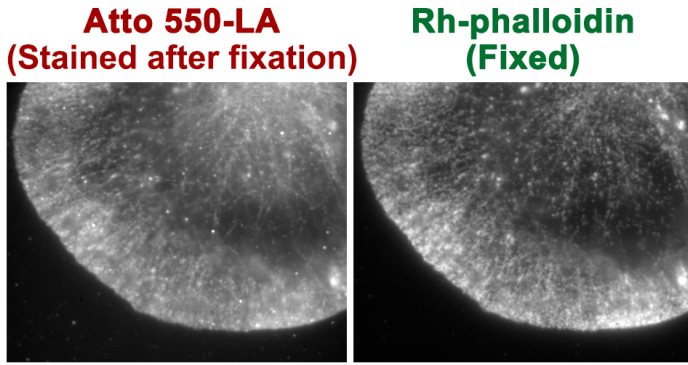
Convection-Induced Biased Distribution of Actin Probes in Live Cells

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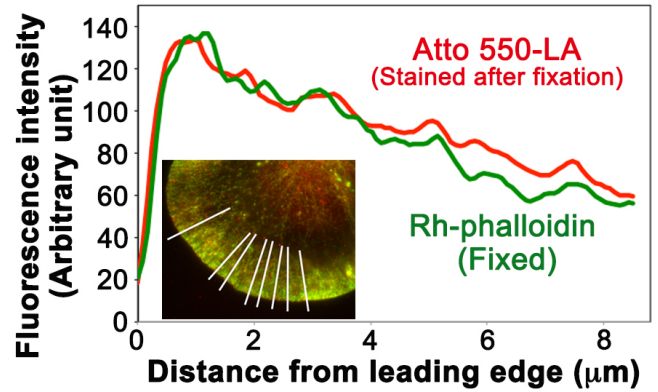
Supplementary Table S1**Model parameters**

Symbol	Description	Value
	Retrograde flow speed relative to cell	
	migration speed	
v	XTC cell	60 nm s ⁻¹
	keratocyte	100 nm s ⁻¹
	Diffusion coefficient of free actin probe	
D	Lifect-mCherry	6.8 μm ² s ⁻¹
	Alexa647-phalloidin	16.7 μm ² s ⁻¹
	Probe association rate	
k_{on}	Lifect-mCherry	2.28 μM ⁻¹ s ⁻¹
	Alexa647-phalloidin	2.9×10 ⁻² μM ⁻¹ s ⁻¹
	Probe dissociation rate	
k_{off}	Lifect-mCherry	30.1 s ⁻¹
	Alexa647-phalloidin	0.08 s ⁻¹
\bar{F}	Uniform F-actin concentration	1000 μM
a	Slope of non-uniform F-actin concentration	112.5 μM μm ⁻¹
b	Base of non-uniform F-actin concentration	100 μM
L	Lamellipodium length	8 μm

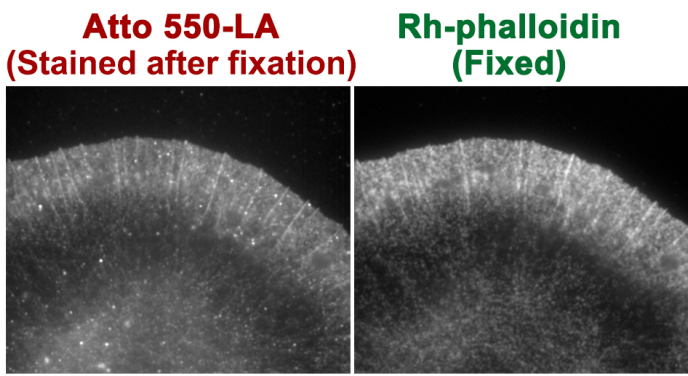
A Cell 1



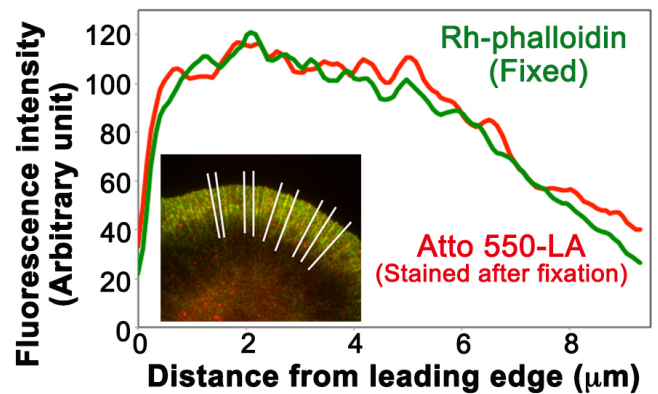
B Cell 1



C Cell 2



D Cell 2



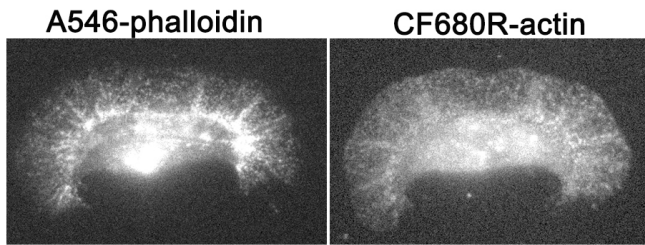
Suppl. Fig. S1

Figure S1. Two other examples of the experiments in Fig. 1 *D* and *E*. The data show similar distribution of Atto 550-Lifeact (Atto 550-LA) and rhodamine-phalloidin (Rh-phalloidin) in fixed XTC cells.

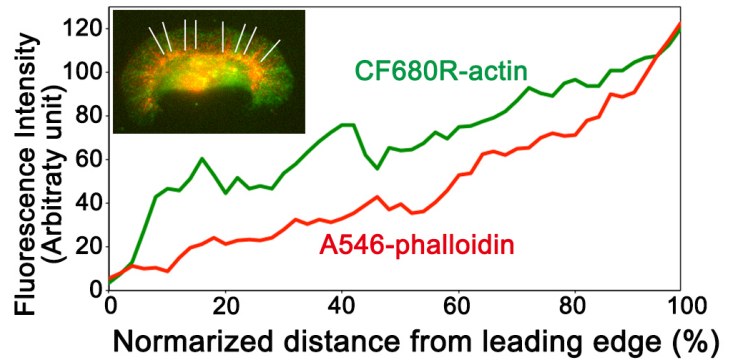
(*A*, and *C*) Images of fixed cells stained with Atto 550-LA (left) and Rh-phalloidin (right). Bars = 5 μm .

(*B*, and *D*) Average fluorescence intensity of the images in *A* or *C* along the white lines in the insets.

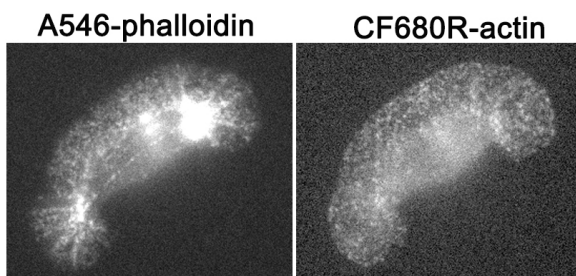
A Cell 1



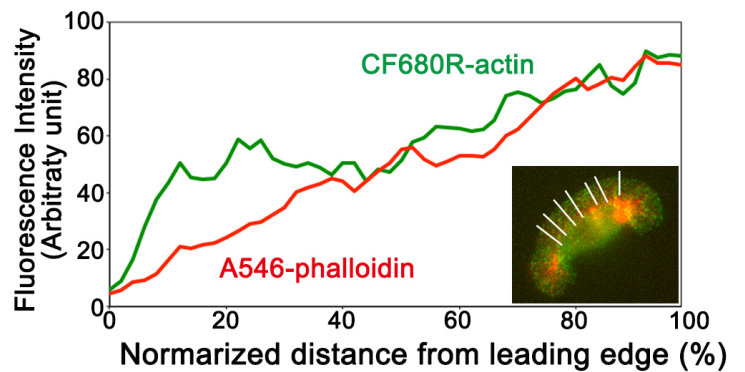
B Cell 1



C Cell 2

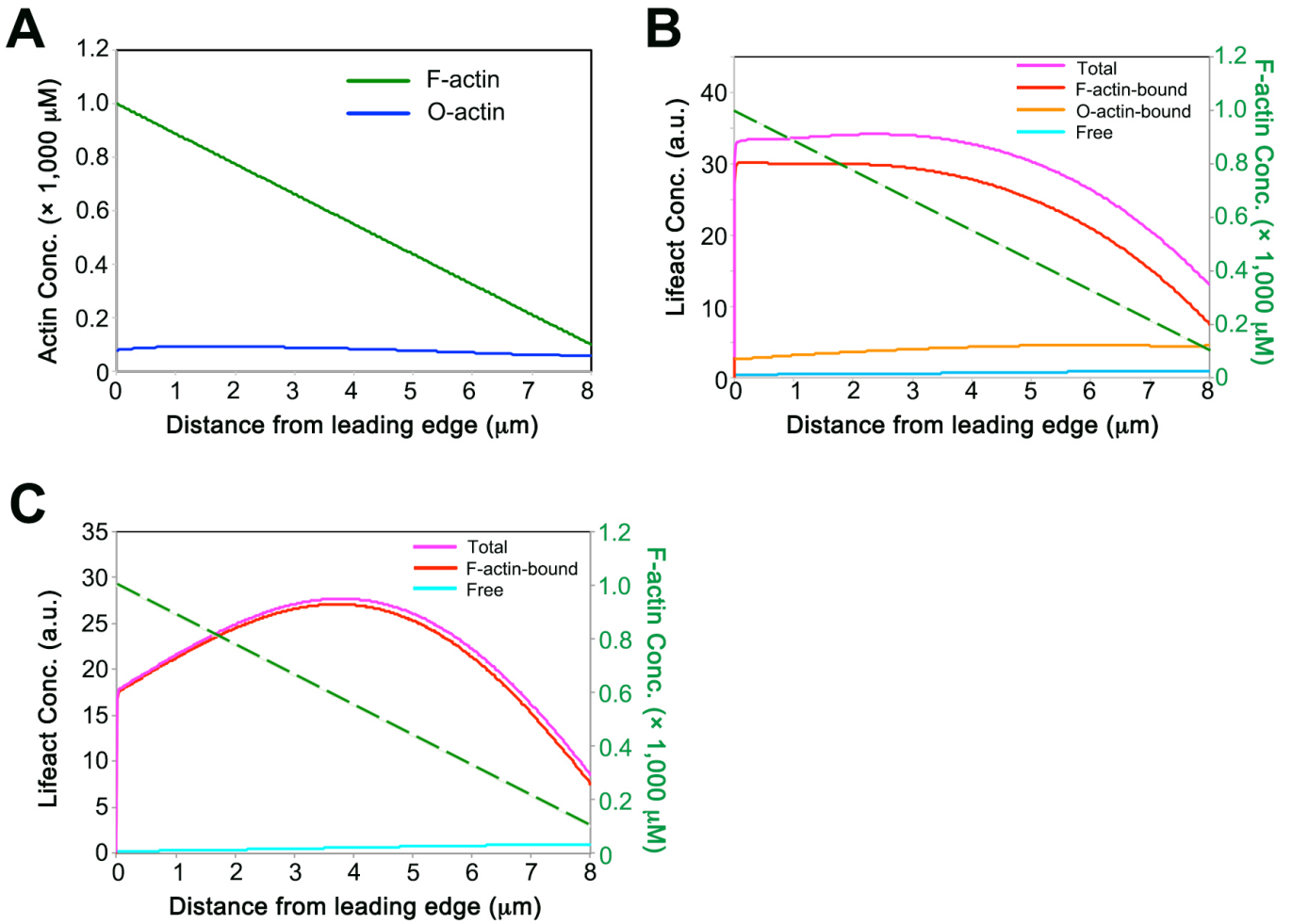


D Cell 2



Suppl. Fig. S2

Figure S2. Two other examples of the experiments in Fig. 2. (**A, and C**) Fluorescent speckle images of Alexa546-phalloidin (A546-phalloidin, left) and CF680R-actin (right) in live fish keratocytes. Bars = 10 μm . (**B, and D**) Average fluorescence intensity of the images in *A* or *C* along the white lines in the insets.



Suppl. Fig. S3

Figure S3. Simulated concentration profile of Lifect-mCherry in a model with both F-actin and actin oligomers (O-actin) as Lifect-binding species. **(A)** Concentration profiles of F- and O-actin in lamellipodia. The ratio of O-actin to F-actin is set as ~ 0.1 at leading edge (29). **(B)** Calculated distributions of Lifect-mCherry in the states of F-actin-bound (red), O-actin-bound (orange) and free (light blue). The distribution of total Lifect-mCherry is indicated by the pink line. A model lamellipodium with a linear decrease in F-actin concentration is indicated in a green dotted line. The supplementary method of simulation including oligomer actin is described below. **(C)** Calculated distribution of Lifect-mCherry in a model with F-actin as only Lifect-binding species. The method and the model parameters for simulation is same to that used in Fig. 3 C, except for the retrograde flow speed was set as 30 nm/s .

[Supplementary method of simulation including oligomer actin employed in Fig. S3B]

1 Model equations

The modified model to take into account the effect of oligomer actin is as follows

$$\frac{\partial C_f}{\partial t} = D_f \frac{\partial^2 C_f}{\partial x^2} + k_{\text{off}} C_{\text{fb}} - k_{\text{on}} F C_f + k_- C_{\text{ob}} - k_+ O C_f + k_d C_{\text{ob}} \quad (1)$$

$$\frac{\partial C_{\text{fb}}}{\partial t} = v \frac{\partial C_{\text{fb}}}{\partial x} - k_{\text{off}} C_{\text{fb}} + k_{\text{on}} F C_f - k_s C_{\text{fb}} + k_i F C_{\text{ob}} \quad (2)$$

$$\frac{\partial C_{\text{ob}}}{\partial t} = D_o \frac{\partial^2 C_{\text{ob}}}{\partial x^2} + k_s C_{\text{fb}} - k_i F C_{\text{ob}} - k_- C_{\text{ob}} + k_+ O C_f - k_d C_{\text{ob}} \quad (3)$$

$$\frac{\partial O}{\partial t} = D_o \frac{\partial^2 O}{\partial x^2} + k_s F - k_i F O - k_d O \quad (4)$$

where C_f , C_{fb} , and C_{ob} are respectively the concentration of free, F-actin-bound, oligomer actin-bound probes. F and O represent the concentration of F-actin and oligomer actin at 1-dimensional position x at time t , respectively. x ranges from 0 (lamellipodial base) from L (leading edge). Based on our experimental data of $F(x)$ in XTC cells, we assumed that the concentration profile of actin, $F(x)$, is prescribed as $F(x) = ax + b$. The model parameters are summarized in Supplementary Table S2.

Supplementary Table S2: Model parameters for Fig. S3B

Symbol	Meaning	Value
v	Retrograde flow speed	30 nm s ⁻¹
D_f	Diffusion coefficient of free Lifeact	6.8 μm ² s ⁻¹
D_o	Diffusion coefficient of oligomer actin	1.0 μm ² s ⁻¹
k_{on}	Rate at which free Lifeact associates with F-actin	2.28 μM ⁻¹ s ⁻¹
k_{off}	Rate at which F-actin-bound Lifeact dissociates from F-actin	30.1 s ⁻¹
k_+	Rate at which free Lifeact associates with O-actin	2.28 μM ⁻¹ s ⁻¹

k_-	Rate at which O-actin-bound Lifeact dissociates from O-actin	30.1 s^{-1}
k_s	F-actin severing rate	0.25 s^{-1}
k_i	Rate at which O-actin is incorporated into F-actin	$0.002 \text{ }\mu\text{M}^{-1} \text{ s}^{-1}$
k_d	Disassembly rate of O-actin	0.5 s^{-1}
a	Slope of linear F-actin concentration profile	$112.5 \text{ }\mu\text{M } \mu\text{m}^{-1}$
b	Base of linear F-actin concentration profile	$100 \text{ }\mu\text{M}$
L	Lamellipodial length	$8 \text{ }\mu\text{m}$

The boundary conditions at the steady state are

$$C_f(0) = \text{const.}, \frac{\partial C_f(L)}{\partial x} = 0 \quad (5)$$

$$vC_{fb}(L) = 0 \quad (6)$$

$$D_o \frac{\partial C_{ob}(0)}{\partial x} = -vC_{fb}(0), D_o \frac{\partial C_{ob}(L)}{\partial x} = -vC_{fb}(L) \quad (7)$$

$$D_o \frac{\partial O(0)}{\partial x} = -vF(0), D_o \frac{\partial O(L)}{\partial x} = -vF(L) \quad (8)$$

In Eq. (5), const. is determined by the total amount of the probe.

2 Spatial discretization

Using the standard finite difference scheme, Eqs. (1)-(4) with the boundary conditions Eqs. (5)-(8) in the steady-state are discretized in space with a step Δx as

$$0 = D_f \frac{C_f(x_{i+1}) - 2C_f(x_i) + C_f(x_{i-1}))}{\Delta x^2} + k_{\text{off}}C_{\text{fb}}(x_i) - k_{\text{on}}F(x_i)C_f(x_i) + k_-C_{\text{ob}}(x_i) - k_+O(x_i)C_f(x_i) + k_dC_{\text{ob}}(x_i) \quad (2 \leq i \leq n-1) \quad (9)$$

$$C_f(x_1) = \text{const.} \quad (10)$$

$$C_f(x_n) = C_f(x_{n-1}) \quad (11)$$

$$0 = v \frac{C_{\text{fb}}(x_{i+1}) - C_{\text{fb}}(x_i)}{\Delta x} - k_{\text{off}}C_{\text{fb}}(x_i) + k_{\text{on}}F(x_i)C_f(x_i) - k_sC_{\text{fb}}(x_i) + k_iF(x_i)C_{\text{ob}}(x_i) \quad (1 \leq i \leq n-1) \quad (12)$$

$$C_{\text{fb}}(x_n) = 0 \quad (13)$$

$$0 = D_o \frac{C_{\text{ob}}(x_{i+1}) - 2C_{\text{ob}}(x_i) + C_{\text{ob}}(x_{i-1}))}{\Delta x^2} + k_sC_{\text{fb}}(x_i) - k_iF(x_i)C_{\text{ob}}(x_i) - k_-C_{\text{ob}}(x_i) + k_+O(x_i)C_f(x_i) - k_dC_{\text{ob}}(x_i) \quad (2 \leq i \leq n-1) \quad (14)$$

$$C_{\text{ob}}(x_1) = C_{\text{ob}}(x_2) + \frac{v\Delta x}{D_o}C_{\text{fb}}(x_1) \quad (15)$$

$$C_{\text{ob}}(x_n) = C_{\text{ob}}(x_{n-1}) - \frac{v\Delta x}{D_o}C_{\text{fb}}(x_n) \quad (16)$$

$$0 = D_o \frac{O(x_{i+1}) - 2O(x_i) + O(x_{i-1}))}{\Delta x^2} + k_sF(x_i) - k_iF(x_i)O(x_i) - k_dO(x_i) \quad (2 \leq i \leq n-1) \quad (17)$$

$$O(x_1) = O(x_2) + \frac{v\Delta x}{D_o}F(x_1) \quad (18)$$

$$O(x_n) = O(x_{n-1}) - \frac{v\Delta x}{D_o}F(x_n) \quad (19)$$

where $x_i = (i-1)\Delta x$ ($i = 1, \dots, n$) and $(n-1)\Delta x = L$. In the present study, we set $\Delta x = 0.01$ [μm].

Rearranging Eqs. (9)-(19) gives

$$C_f(x_i) = \frac{C_f(x_{i+1}) + C_f(x_{i-1}) + c_3C_{\text{fb}}(x_i) + c_4C_{\text{ob}}(x_i)}{c_1F(x_i) + c_2O(x_i) + 2} \quad (2 \leq i \leq n-1) \quad (20)$$

$$C_f(x_1) = \text{const.} \quad (21)$$

$$C_f(x_n) = C_f(x_{n-1}) \quad (22)$$

$$C_{\text{fb}}(x_i) = \frac{1}{c_5} [C_{\text{fb}}(x_{i+1}) + c_6F(x_i)C_f(x_i) + c_7F(x_i)C_{\text{ob}}(x_i)] \quad (1 \leq i \leq n-1) \quad (23)$$

$$C_{\text{fb}}(x_n) = 0 \quad (24)$$

$$C_{\text{ob}}(x_i) = \frac{C_{\text{ob}}(x_{i+1}) + C_{\text{ob}}(x_{i-1}) + c_{10}C_{\text{fb}}(x_i) + c_{11}O(x_i)C_f(x_i)}{c_8F(x_i) + c_9} \quad (2 \leq i \leq n-1) \quad (25)$$

$$C_{\text{ob}}(x_1) = C_{\text{ob}}(x_2) + c_{13}C_{\text{fb}}(x_1) \quad (26)$$

$$C_{\text{ob}}(x_n) = C_{\text{ob}}(x_{n-1}) \quad (27)$$

$$O(x_i) = \frac{O(x_{i+1}) + O(x_{i-1}) + c_{10}F(x_i)}{c_8F(x_i) + c_{12}} \quad (2 \leq i \leq n-1) \quad (28)$$

$$O(x_1) = O(x_2) + c_{13}F(x_1) \quad (29)$$

$$O(x_n) = O(x_{n-1}) - c_{13}F(x_n) \quad (30)$$

where

$$c_1 = \frac{k_{\text{on}}\Delta x^2}{D_f}, c_2 = \frac{k_+\Delta x^2}{D_f}, c_3 = \frac{k_{\text{off}}\Delta x^2}{D_f}, c_4 = \frac{(k_- + k_d)\Delta x^2}{D_f}, c_5 = 1 + \frac{(k_{\text{off}} + k_s)\Delta x}{v} \quad (31)$$

$$c_6 = \frac{k_{\text{on}}\Delta x}{v}, c_7 = \frac{k_i\Delta x}{v}, c_8 = \frac{k_i\Delta x^2}{D_o}, c_9 = 2 + \frac{(k_- + k_d)\Delta x^2}{D_o}, c_{10} = \frac{k_s\Delta x^2}{D_o} \quad (32)$$

$$c_{11} = \frac{k_+\Delta x^2}{D_o}, c_{12} = 2 + \frac{k_d\Delta x^2}{D_o}, c_{13} = \frac{v\Delta x}{D_o} \quad (33)$$

Notice that Eqs. (20)-(30) remain unsolved with respect to $C_f(x_i), C_{fb}(x_i), C_{ob}(x_i)$ and $O(x_i)$ because the RHS of Eqs. (20)-(30) also include unknown $C_f(x_{i+1}), C_f(x_{i-1}), C_{fb}(x_{i+1}), C_{ob}(x_{i+1}), C_{ob}(x_{i-1}), O(x_{i-1})$ and $O(x_{i-1})$.

3 Iterative method to obtain the steady-state solution

To fully solve Eqs. (20)-(30), we used the following iterative update of $C_f(x_i), C_{fb}(x_i), C_{ob}(x_i)$, and $O(x_i)$:

$$C_f^{[k+1]}(x_i) \leftarrow \frac{C_f^{[k]}(x_{i+1}) + C_f^{[k]}(x_{i-1}) + c_3 C_{fb}^{[k]}(x_i) + c_4 C_{ob}^{[k]}(x_i)}{c_1 F(x_i) + c_2 O^{[k]}(x_i) + 2} \quad (2 \leq i \leq n-1) \quad (34)$$

$$C_f^{[k+1]}(x_n) \leftarrow C_f^{[k+1]}(x_{n-1}) \quad (35)$$

$$C_{fb}^{[k+1]}(x_i) \leftarrow \frac{1}{c_5} \left[C_{fb}^{[k]}(x_{i+1}) + c_6 F(x_i) C_f^{[k]}(x_i) + c_7 F(x_i) C_{ob}^{[k]}(x_i) \right] \quad (1 \leq i \leq n-1) \quad (36)$$

$$C_{ob}^{[k+1]}(x_i) \leftarrow \frac{C_{ob}^{[k]}(x_{i+1}) + C_{ob}^{[k]}(x_{i-1}) + c_{10} C_{fb}^{[k]}(x_i) + c_{11} O^{[k]}(x_i) C_f^{[k]}(x_i)}{c_8 F(x_i) + c_9} \quad (2 \leq i \leq n-1) \quad (37)$$

$$C_{ob}^{[k+1]}(x_1) \leftarrow C_{ob}^{[k+1]}(x_2) + c_{13} C_{fb}^{[k+1]}(x_1) \quad (38)$$

$$C_{ob}^{[k+1]}(x_n) \leftarrow C_{ob}^{[k+1]}(x_{n-1}) \quad (39)$$

$$O^{[k+1]}(x_i) \leftarrow \frac{O^{[k]}(x_{i+1}) + O^{[k]}(x_{i-1}) + c_{10} F(x_i)}{c_8 F(x_i) + c_{12}} \quad (2 \leq i \leq n-1) \quad (40)$$

$$O^{[k+1]}(x_1) \leftarrow O^{[k+1]}(x_2) + c_{13} F(x_1) \quad (41)$$

$$O^{[k+1]}(x_n) \leftarrow O^{[k+1]}(x_{n-1}) - c_{13} F(x_n) \quad (42)$$

where the superscript $[k]$ indicates the number of iteration steps. It is clear that one can obtain the steady-state solution of Eqs. (1)-(4) after the convergence of the loop with respect to k . During the iterations, we kept $C_f(x_1)$ constant, namely, 1, and also maintained $C_{fb}(x_n) = 0$. We set the initial guess of C_f, C_{fb}, C_{ob} , and O as $C_f(x_1) = 1, C_f(x_{2 \sim n}) = 0, C_{fb}(x_{1 \sim n}) = 0, O(x_1) = O(x_2) + c_{13} F(x_1), O(x_{2 \sim n-1}) = k_s F(x_{2 \sim n-1}) / [k_i F(x_{2 \sim n-1}) + k_d]$, and $O(x_n) = O(x_{n-1}) - c_{13} F(x_n)$. We judged convergence of the loop if all of the following conditions are satisfied: $\| C_f^{[k+1]} - C_f^{[k]} \|^2 \leq \epsilon, \|$

$C_{fb}^{[k+1]} - C_{fb}^{[k]} \|^2 \leq \epsilon, \| C_{ob}^{[k+1]} - C_{ob}^{[k]} \|^2 \leq \epsilon$ and $\| O^{[k+1]} - O^{[k]} \|^2 \leq \epsilon$ ($\epsilon = 10^{-10}$). After the convergence, we normalized C_f, C_{fb} and C_{ob} such that they satisfy $\int_0^L (C_f + C_{fb} + C_{ob}) dx = C_{tot}$ where C_{tot} is the total amount of actin probes in lamellipodia. In the present study, we set $C_{tot} = 1$.

Instead of Eq. (12) where the forward finite difference scheme was used, one might use the central finite difference scheme to replace Eq. (36) with

$$C_{fb}^{[k+1]}(x_i) \leftarrow \frac{1}{c_5 - 1} \left[C_{fb}^{[k]}(x_{i+1}) - C_{fb}^{[k]}(x_{i-1}) + c_6 F(x_i) C_f^{[k]}(x_i) + c_7 F(x_i) C_{ob}^{[k]}(x_i) \right] \quad (43)$$

However, we found that iterative updating using Eq.(43) was unstable because the RHS of Eq (43) can sometimes become negative due to $-C_{fb}^{[k]}(x_{i-1})$ depending on model parameters and shape of $C_{fb}(x_i)$. To ensure positivity of C_{fb} during all updating steps, we used the forward difference scheme.

A Appendix: analytical solution

In the special case where $k_i = 0$ is satisfied, the analytical solution of O can be obtained. we used the analytical solution to check the validity of the iterative method. The steady-state model equation about O-actin and its BCs are

$$\frac{\partial^2 O}{\partial x^2} + k_s F - k_i FO - k_d O = 0 \quad (44)$$

$$D_o \frac{\partial O(0)}{\partial x} = -vF(0), D_o \frac{\partial O(L)}{\partial x} = -vF(L), F(x) = ax + b \quad (45)$$

The solution of which is given by

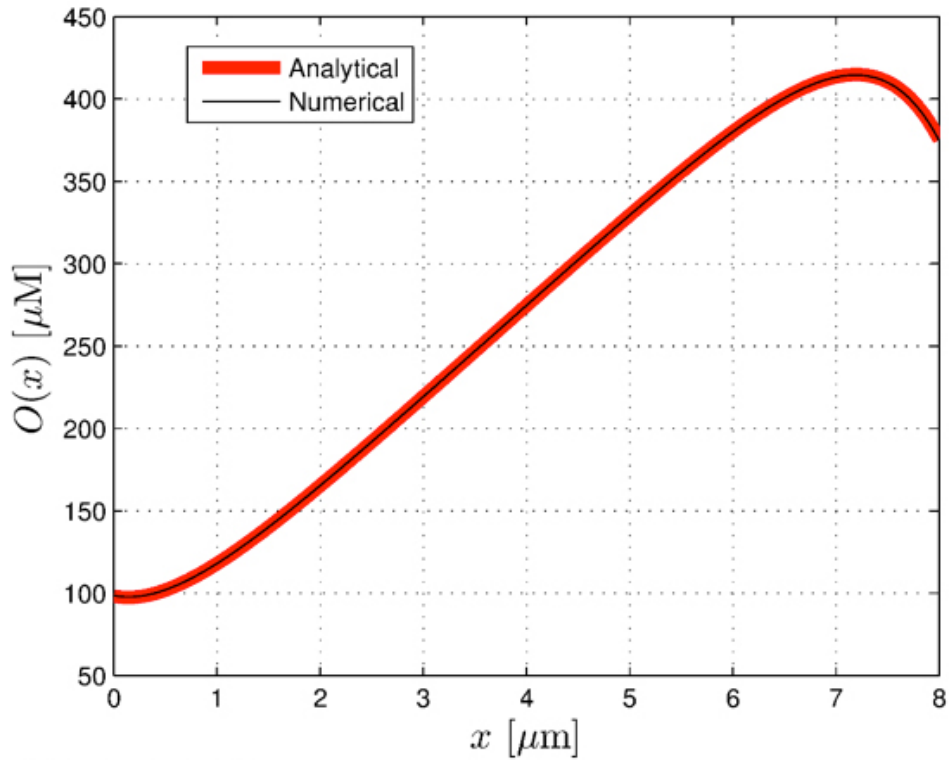
$$O(x) = \frac{k_s}{k_d}(ax + b) + C_1 \exp\left(-\frac{x}{\lambda}\right) + C_2 \exp\left(\frac{x}{\lambda}\right) \quad (46)$$

$$C_1 = \frac{\lambda}{\exp\left(-\frac{2L}{\lambda}\right) - 1} \left\{ \left(\frac{ak_s}{k_d} + \frac{bv}{D_o} \right) \left[\exp\left(-\frac{L}{\lambda}\right) - 1 \right] + \frac{avL}{D_o} \exp\left(-\frac{L}{\lambda}\right) \right\} \quad (47)$$

$$C_2 = C_1 - \frac{ak_s\lambda}{k_d} - \frac{bv\lambda}{D_o} \quad (48)$$

where $\lambda = \sqrt{D_o/k_d}$.

Supplementary Figure S4 compares the numerical solution and analytical solution at $D_o = 0.25$ [$\mu\text{m}^2\text{s}^{-1}$]. The numerical solution agrees well with the analytical one.



Suppl. Fig. S4

Figure S4 Comparison between analytical and numerical solutions. $D_o = 0.25$ [$\mu\text{m}^2\text{s}^{-1}$] and $k_i = 0$ [$\mu\text{M}^{-1}\text{s}^{-1}$].